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Original article

Increased incidence of Parkinsonism among Chinese with β -glucosidase mutation in central Taiwan

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ARTICLE INFO

Article history:

Received 21 December 2012

Received in revised form

22 January 2013

Accepted 31 January 2013

Available online 6 March 2013

Keywords:

β -glucosidase

L444P

mutation

Parkinson's disease

Taiwanese

ABSTRACT

Background: Parkinson's disease (PD) is characterized by progressive neuronal cell loss and decline in movement. Recently, much attention has been focused on the dopaminergic neurotoxicity, which causes neurodegeneration in the nervous system, thus implicating dysfunction in predisposition to PD pathogenesis. β -glucosidase (GBA), the enzyme deficient in Gaucher's disease, has been cited as linked with Parkinson's disease. We investigated whether GBA mutation is associated with PD patients in middle Taiwan.

Methods: GBA mutation was detected by polymerase chain reaction and sequencing in 148 patients with PD and 120 normal controls.

Results: The results revealed a significant difference between PD patients and normal controls in GBA mutation and a statistic correlation between GBA mutation L444P and PD formation. It could be concluded that patients who carry the T/C of GBA mutation, L444P, have a higher risk of developing PD in Chinese patients in middle Taiwan.

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1. Introduction

Parkinson's disease (PD) is characterized by the progressive loss of dopaminergic neurons and the presence of intracellular Lewy body inclusions within the *substantia nigra*. The presence of age-associated cytopathogenic process is required for a definite diagnosis. Clinically, PD patients exhibit at least

one of resting tremor, muscle rigidity, bradykinesia, and postural instability, which are features of neurodegenerative disorder [1,2]. Patients with familial PD provide a good insight for examination of candidate genes; however, the incidence of PD pathogenesis is still too complex to unify the common mechanisms of etiologies. In particular, the major causes of the disease occur in a sporadic manner, possibly suggesting

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that the susceptibility of PD is involved in a wide spectrum of genetic and environmental factors.

Current theories of PD pathogenesis are based on available data from familiar histories of the disease. Mutations in five causal genes— α -synuclein, parkin, DJ-1, PINK1, and leucine-rich repeat kinase 2 (LRRK2) [3–9]—have been identified as molecular events increasing risk of disease. Evidence for the association of the PD candidate gene LRRK2, which encodes the protein kinase dardarin and is located within PARK8 locus on chromosome 12p11.2-q13.1 [4,5], is growing. Genetic evidence of its linkage to hereditary late-onset PD was originally detected in a Japanese family [4,10]. Then, as currently identified from a rare familial pedigree of PD, finding several LRRK2 gene mutations revealed it as causing autosomal-dominant familiar Parkinsonism [4,5].

The gene encoding β -glucosidase (also known as glucocerebrosidase; GBA1) has been localized to chromosome 1q21 [11]; a highly homologous pseudogene sequence exists at 16 kb downstream [11,12]. More than 200 mutations, point mutations, deletions, insertions, splicing aberrations, and various rearrangements have been identified within the GBA gene [13–15]. Environmental factors are known to play a role in PD pathogenesis [16,17]. This case–control association study explored a hypothesis of common coding variation within GBA gene also tending toward susceptibility of sporadic PD among Chinese in central Taiwan. In the search for variants, we undertook the potentially functional mutation for frequent analysis in sporadic forms of PD cases and ethnically-matched controls among the Taiwanese population.

2. Patients and methods

2.1. Patients

A standard clinical examination was performed on every participant, and PD diagnosis was confirmed according to published neurological criteria [1,2,18]. The cohort enrolled 148 PD cases and 120 healthy controls from the same population in central Taiwan. Patients were of Taiwanese ethnicity, with sporadic PD based on pedigree assay, as each member in the absence of family history was observed.

2.2. PCR amplification

PCR amplification of the DNA fragments covering mutation L444P of GBA was conducted from genomic DNA of patients and controls; PCR amplified the DNA fragments of GBA according to Tsai et al [9]. Briefly, PCR reaction was performed under the conditions: PCR carried out in total volume of 50 μ L, containing 50 ng genomic DNA, 1 \times Taq polymerase buffer, 5 pmol of each primer, and 0.25 U of AmpliTaq DNA polymerase (Perkin Elmer, Foster City, CA, USA). Amplification conditions were: (1) denaturation at 95°C for 4 minutes; (2) 30 cycles of denaturation at 95°C for 30 seconds, annealing at 55°C for 30 seconds, and extension at 72°C for 45 seconds; and (3) final elongation at 72°C for 10 minutes. Preventive contamination entailed PCR reaction mixture without DNA (negative control) in each run of amplification.

2.3. DNA sequencing

All PCR fragments were purified by QIAEX II (Qiaagen, Hilden, Germany) and directly sequenced to identify mutations. The direct sequencing was achieved by an ABI Prism 3100 DNA sequencer (PE Applied Biosystems) with the BigDye Dideoxy Terminator Sequencing Kit (PE Applied Biosystems).

2.4. Statistical analysis

The parameters were compared using Fisher’s exact test, with $p < 0.05$ statistically significant. SPSS software (version 10.0; SPSS Inc., Chicago, IL, USA) was used to performed calculations.

3. Results

The L444P mutation was screened by PCR and sequencing. The screening of L444P mutation showed all patients carrying PD hetero- or homozygote genotype in their GBA gene. Table 1 plots genotype distribution and allelic frequencies for L444P mutation in patients and controls. Significant differences arose between patients and controls ($p < 0.001$) in L444P genotype distribution, with T/C homozygote distribution in patients (4.51%) higher than in controls (0%). Also, allelic frequency of L444P mutation showed statistical significance, distinguishing PD patients from controls ($p = 0.048$; Fisher’s exact test).

4. Discussion

This study analyzes the GBA gene in PD among the Chinese population of central Taiwan. Data indicate GBA mutation L444P is associated with PD. Since significance is dominant, this association is not likely to stem from a chance event. Earlier study showed L444P mutation as second most common Gaucher’s disease (GD) mutation among non-Jewish patients, accounting for 37% of total mutations surveyed [19,20]. In our research, L444P accounts for 4.51% of PD cases. A cause for the linkage of mutant GBA with Parkinsonism remains unclear; further studies must identify the pathological mechanism. Recently, the mutation in GBA was noted in relation to

Table 1 – Distribution of genotypes among Parkinson’s disease (PD) patients and healthy controls.

Genotype	PD patients	Normal controls	<i>p</i>
	<i>n</i> (%)	<i>n</i> (%)	
T/T	148 (95.48)	120 (100)	<0.001
T/C	7 (4.51)	0 (0)	
C/C	0 (0)	0 (0)	
Allelic frequency			0.048
Allele T	303 (97.74)	240 (100)	
Allele C	7 (2.26)	0 (0)	

p values calculated by Fisher’s exact test.

synucleinopathies. Some studies pay attention to this association [21,22]; others find a correlation between Parkinsonism and the GBA mutation, the gene encoding the enzyme deficient in autosomal, recessively-inherited lysosomal storage, also known as GD. Rare GD cases developing Parkinsonism have been identified [23,24]. Neuropathological evaluation of several such patients showed intraneuronal inclusions with α -synuclein in brain regions related to GD [24,25]. Additional studies focusing on diverse cohorts with Parkinsonism indicate higher frequency of specific GBA mutations. Increasing GBA mutations in PD, identification of mutations in the spectrum of synucleinopathy [26], and higher frequency of Parkinsonism among GD patients [25] led us to research this correlation further, via the genetic approach. Our analysis lends compelling evidence of the GBA gene L444P mutation contributing significant association, thereby implicating it as one possible genetic risk factor for sporadic PD and accounting for higher prevalence of disease in allele frequencies. These data may extend the survey of GBA to other populations for sporadic form of PD. Detailed study of molecular mechanisms must explore the role of GBA and related drug development in PD.

Acknowledgments

This work was funded by grants from both the National Science Council in Taiwan (NSC 97-2320-B-039-021-MY3) and China Medical University Hospital (CMU100-S-27, DMR-100-132, and CMU98-P-004). This study was also supported in part by the Taiwan Department of Health Clinical Trial and Research Center of Excellence (DOH101-TD-B-111-004; DOH102-TD-B-111-004).

REFERENCES

- [1] Gelb DJ, Oliver E, Gilman S. Diagnostic criteria for Parkinson disease. *Arch Neurol* 1999;56:33–9.
- [2] Hughes AJ, Daniel SE, Kilford L, Lees AJ. Accuracy of clinical diagnosis of idiopathic Parkinson's disease: a clinicopathological study of 100 cases. *J Neurol Neurosurg Psychiatr* 1992;55:181–4.
- [3] Deng H, Le WD, Xie WJ, Pan TH, Zhang X, Jankovic J. Genetic analysis of parkin co-regulated gene (PACRG) in patients with early-onset parkinsonism. *Neurosci Lett* 2005;382:297–9.
- [4] Zimprich A, Biskup S, Leitner P, Lichtner P, Farrer M, Lincoln S, et al. Mutations in LRRK2 cause autosomal-dominant parkinsonism with pleomorphic pathology. *Neuron* 2004;44:601–7.
- [5] Paisín-Ruiz C, Jain S, Evans EW, Gilks WP, Simón J, van der Brug M, et al. Cloning of the gene containing mutations that cause PARK8-linked Parkinson's disease. *Neuron* 2004;44:595–600.
- [6] Polymeropoulos MH, Lavedan C, Leroy E, Ide SE, Dehejia A, Dutra A, et al. Mutation in the alpha-synuclein gene identified in families with Parkinson's disease. *Science* 1997;276:2045–7.
- [7] Kitada T, Asakawa S, Hattori N, Matsumine H, Yamamura Y, Minoshima S, et al. Mutations in the parkin gene cause autosomal recessive juvenile parkinsonism. *Nature* 1998;392:605–8.
- [8] Bonifati V, Rizzu P, Squitieri F, Krieger E, Vanacore N, van Swieten JC, et al. DJ-1 (PARK7), a novel gene for autosomal recessive, early onset parkinsonism. *Neurol Sci* 2003;24:159–60.
- [9] Valente EM, Abou-Sleiman PM, Caputo V, Muqit MM, Harvey K, Gispert S, et al. Hereditary early-onset Parkinson's disease caused by mutations in PINK1. *Science* 2004;304:1158–60.
- [10] Funayama M, Hasegawa K, Kowa H, Saito M, Tsuji S, Obata F. A new locus for Parkinson's disease (PARK8) maps to chromosome 12p11.2-q13.1. *Ann Neurol* 2002;51:296–301.
- [11] Horowitz M, Wilder S, Horowitz Z, Reiner O, Gelbart T, Beutler E. The human glucocerebrosidase gene and pseudogene: structure and evolution. *Genomics* 1989;4:87–96.
- [12] Winfield SL, Tayebi N, Martin BM, Ginns EI, Sidransky E. Identification of three additional genes contiguous to the glucocerebrosidase locus on chromosome 1q21: implications for Gaucher disease. *Genome Res* 1997;7:1020–6.
- [13] Filocamo M, Mazzotti R, Stroppiano M, Seri M, Giona F, Parenti G, et al. Analysis of the glucocerebrosidase gene and mutation profile in 144 Italian Gaucher patients. *Hum Mutat* 2002;20:234–5.
- [14] Beutler E, Gelbart T, Scott CR. Hematologically important mutations: Gaucher disease. *Blood Cells Mol Dis* 2005;35:355–64.
- [15] Stone DL, Tayebi N, Orvisky E, Stubblefield B, Madike V, Sidransky E. Glucocerebrosidase gene mutations in patients with type 2 Gaucher disease. *Hum Mutat* 2000;15:181–8.
- [16] Singh M, Khan AJ, Shah PP, Shukla R, Khanna VK, Parmar D. Polymorphism in environment responsive genes and association with Parkinson disease. *Mol Cell Biochem* 2008;312:131–8.
- [17] Kuehn BM. Scientists probe role of genes, environment in Parkinson disease. *JAMA* 2006;295:1883–5.
- [18] Hughes AJ, Daniel SE, Lees AJ. Improved accuracy of clinical diagnosis of Lewy body Parkinson's disease. *Neurology* 2001;57:1497–9.
- [19] Ida H, Iwasawa K, Kawame H, Rennert OM, Maekawa K, Eto Y. Characteristics of gene mutations among 32 unrelated Japanese Gaucher disease patients: absence of the common Jewish 84GG and 1226G mutations. *Hum Genet* 1995;95:717–20.
- [20] Ida H, Rennert OM, Iwasawa K, Kobayashi M, Eto Y. Clinical and genetic studies of Japanese homozygotes for the Gaucher disease L444P mutation. *Hum Genet* 1999;105:120–6.
- [21] Cullen V, Sardi SP, Ng J, Xu YH, Sun Y, Tomlinson JJ, et al. Acid beta-glucosidase mutants linked to Gaucher disease, Parkinson disease, and Lewy body dementia alter alpha-synuclein processing. *Ann Neurol* 2011;69:940–53.
- [22] Kim MJ, Suh JT, Lee HJ, Lee WI, Moon A, Lee J, et al. Simultaneous detection of Gaucher's disease and renal involvement of non-Hodgkin's lymphoma: the first Asian case report and a review of literature. *Ann Clin Lab Sci* 2012;42:293–301.
- [23] Neudorfer O, Giladi N, Elstein D, Abrahamov A, Turezkite T, Aghai E, et al. Occurrence of Parkinson's syndrome in type I Gaucher disease. *QJM* 1996;89:691–4.
- [24] Tayebi N, Walker J, Stubblefield B, Orvisky E, LaMarca ME, Wong K, et al. Gaucher disease with parkinsonian manifestations: does glucocerebrosidase deficiency contribute to a vulnerability to parkinsonism? *Mol Genet Metab* 2003;79:104–9.
- [25] Wong K, Sidransky E, Verma A, Mixon T, Sandberg GD, Wakefield LK, et al. Neuropathology provides clues to the pathophysiology of Gaucher disease. *Mol Genet Metab* 2004;82:192–207.
- [26] Goker-Alpan O, Giasson BI, Eblan MJ, Nguyen J, Hurtig HI, Lee VM, et al. Glucocerebrosidase mutations are an important risk factor for Lewy body disorders. *Neurology* 2006;67:908–10.