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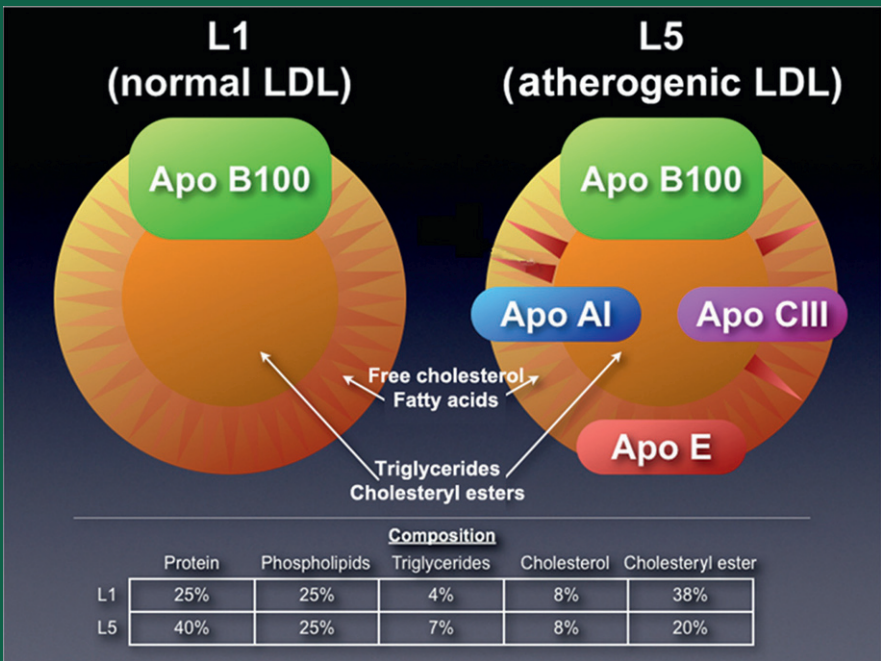
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## Editorial

# Cardiovascular disease and cancer progression—A brief insight

Apart from the environmental factors, the complex interaction of genes plays a central role in the development of diseases. Growing evidence indicates that death due to cardiovascular complications and cancer is increasing and survival rate decreasing due to the availability of fewer therapeutic options. The current issue of *Biomedicine* focuses on some of the risk factors that influence cardiovascular disease, and on the functional roles of androgenic signals, very long noncoding RNAs (vlncRNAs), and betel quid chewing in cancer progression.

The first paper reviews the current literature on angiotensin-converting enzyme inhibitory peptides, focusing on their structure–activity relationship and inhibitory mechanisms. In addition, an *in vitro* simulated gastrointestinal digestion model for assessing bioavailability and an *in vivo* examination of the antihypertensive effects of the peptides are also summarized.

The second review mainly focuses on the types of hyperlipidemia, digestion, and absorption of lipids, as well as their consequences on human health, and on potentially effective new therapeutic targets for treating hyperlipidemia.

Low-density lipoprotein (LDL) abnormality is a central cause of atherosclerosis and is associated with complications such as coronary artery disease. Negatively charged LDL (L5), a subclass of circulating LDL, is a nonoxidized or merely oxidized LDL, but it is potentially atherogenic. L5 was isolated from human plasma for mechanistic scrutiny. This review article elucidates the potential role of L5 in endothelial dysfunction and atherosclerosis formation.

Androgenic signals (androgen/androgen receptor; A/AR) function in a biphasic manner in the progression of hepatocellular carcinoma. In this review, the author discusses the roles of the A/AR in hepatic host immunity and the hepatic damage/regenerating state, and how the A/AR signals influence hepatitis B virus replication in cirrhotic livers.

Long noncoding RNAs (lncRNAs) and vlncRNAs, which are more than 5000 nt long, are associated with numerous bio-

logical functions. XIST, MALAT1, PCA3, PCGEM1, and PCNCR1 are some of the vlncRNAs expressed during the progression of cancer. This review focuses on vlncRNAs, a specific group of lncRNAs that are more than 5000 nt long, and explored their roles in the development of diseases.

Although extensive research findings have linked betel quid chewing with oral cancer and precancerous conditions, to date, no pharmacological or behavioral treatment exists for betel quid cessation. In this review, the author reports on the prevalence of betel quid use in Taiwan and the link between betel quid chewing, smoking, and oral cancer among Taiwanese men. The author also elaborately discusses the defaults about smoking and betel quid addiction.

Overall, this issue widens and deepens our knowledge of cardiovascular disease and cancer research by providing new perspectives. Further studies are required to facilitate the development of novel therapeutic applications to treat the abovementioned diseases.

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## Review article

# Angiotensin I-converting enzyme inhibitory peptides: Inhibition mode, bioavailability, and antihypertensive effects

Chia-Ling Jao<sup>a</sup>, Shih-Li Huang<sup>b</sup>, Kuo-Chiang Hsu<sup>c,\*</sup>

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## ABSTRACT

Bioactive peptides within the original food-derived proteins are inactive but can be activated by releasing them during food processing (by enzymatic hydrolysis or fermentation) or during gastrointestinal (GI) digestion. Among all the bioactive peptides, the antihypertensive peptides attract particular attention owing to the prevalence of high blood pressure, which plays an important role in cardiovascular diseases. These peptides have the ability to act as angiotensin I-converting enzyme (ACE) inhibitors. Previous studies have shown that the ACE inhibitory peptides functioned as competitive, noncompetitive, or uncompetitive inhibitors, and therefore, the structure–activity relationship of the peptides with various inhibition modes needs to be clarified. Besides, the ACE inhibitory activity of these peptides *in vitro* does not always suggest its antihypertensive effect *in vivo*, which is based on its fate to encounter GI enzymes and brush-border membrane peptidases, after oral administration. This paper reviews the current literature on ACE inhibitory peptides, focusing on the structure–activity relationship and inhibition mechanisms due to their inhibition modes. In addition, the *in vitro*–simulated GI digestion for assessing bioavailability and *in vivo* antihypertensive effects of the peptides are also summarized.

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## 1. Introduction

Cardiovascular disease (CVD), a class of diseases that affects the heart and blood vessels, has been recognized as the biggest cause of death worldwide. High blood pressure, or hypertension, is a condition of sustained increase in blood pressure levels and is the primary risk factor for CVD. It is reported that >25% of the population worldwide

(approximately 1 billion) had been affected by hypertension in 2000 and according to the findings of Kearney et al this figure is predicted to increase to 1.56 billion by 2025 [1].

The renin–angiotensin system is a hormone system that regulates blood pressure and fluid balance, and plays an important role in the pathophysiology of CVDs such as congestive heart failure and hypertension [2]. Plasma renin is responsible for the conversion of angiotensinogen released by

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the liver into angiotensin I, which subsequently undergoes proteolytic cleavage, in the presence of angiotensin I-converting enzyme (ACE), to form angiotensin II in the lungs. The hormone angiotensin II is a vasoconstrictor and its formation increases blood pressure. In addition, ACE also degrades bradykinin which has vasodilatation properties. Therefore, the use of ACE inhibitors is believed to lower hypertension and further prevent CVDs.

ACE inhibitors are originally synthesized from compounds found in pit viper venom, and synthetic ACE inhibitors such as captopril, enalapril, lisinopril, and ramipril are currently used in the treatment of hypertension in humans [3,4]. Although these synthetic inhibitors show a remarkable effect in treating hypertension, they also cause adverse side effects, such as cough, allergic reactions, taste disturbances, and skin rashes. Thus, the development of safe and natural ACE inhibitors is necessary for future treatment and prevention of hypertension.

Many studies have successfully produced and isolated ACE inhibitory peptides from various food proteins such as gelatin [5], milk [6], maize [7], sunflower [8], ovalbumin [9], and wheat germ [10]. These peptides have frequently been reported to act as competitive inhibitors of ACE [11–13]; however, in recent years, some noncompetitive and uncompetitive ACE inhibitory peptides have also been isolated [14–17]. Although some studies have demonstrated the relationship between ACE inhibitory activities and peptide structures, only few studies discussed about the activity and inhibition mode of these peptides. On the other hand, in order to reduce increased blood pressure levels after oral administration, the peptides possessing *in vitro* ACE inhibitory activity have to reach the target organ in the organism in an active form. However, because these peptides are degraded by gastrointestinal (GI) enzymes, there is an inconsistency between their *in vitro* ACE inhibitory activity and *in vivo* antihypertensive activity. Although some researchers have used *in vitro*-simulated GI digestion to evaluate the bioavailability and bioactivity of ACE inhibitory peptides [18–20], the correspondence between both *in vitro* and *in vivo* effects has not been investigated in many studies. In this review, the relationship between ACE inhibitory activity and inhibition mode is introduced first. Then, the discrepancy between *in vitro* and *in vivo* activity is examined in the view of bioavailability. Finally, some future perspectives of these peptides are also discussed.

## 2. Inhibition mode of ACE inhibitory peptides

Proteins are well-known precursors of a range of biologically active peptides. The biologically active (also called “bioactive”) peptides are derived from food proteins that have a physiological effect in the body in addition to their nutritional value. The fact that proteins are precursors of biologically active molecules is particularly attractive for the development of functional foods, because the bioactive peptides are commonly-used food ingredients and are of natural origin. As compared with chemosynthetic drugs, food protein-derived peptides can be used as potent pharmaceuticals as alternatives to synthetic drugs because of the increasing interest for safe and economical use of drugs. The bioactive peptides are

activated when released from proteins by enzymatic or acidic hydrolysis, and their biological activity is determined by their native amino acid composition and sequence [21].

Since the isolation of the first ACE inhibitory peptide from snake venom [22], many other ACE inhibitory peptides have been discovered in the enzymatic hydrolysates of various food proteins, including animal-, plant-, and microorganism-derived peptides. Table 1 [12,16,17,19,23–27,30,31,33–36] shows a summary of ACE inhibitory peptides from different food proteins categorized by their inhibition mode.

### 2.1. Competitive inhibitor

The inhibition mode of ACE inhibitory peptides is evaluated using Lineweaver–Burk plots. The competitive inhibitors can bind to the active site to block it or to the inhibitor-binding site that is remote from the active site so as to alter the enzyme conformation such that the substrate no longer binds to the active site (Fig. 1) [23]. For example, NLP [24], NG [24], YN [25], LGFPTTKTYFPHF [26], VVYPWT [26], LNVPGEIVE [27], NIPPLTQTPV [27], DKIHPP [27], LF [28], WA [28], WM [28], were reported to be competitive inhibitors. A previous study has reported that the active sites of two domains of somatic ACE are structurally and functionally homologous to a dipeptidyl carboxypeptidase, and that the zinc-coordination geometry is critical for their hydrolytic action [3]. However, the two catalytic sites are differentially activated by chloride ions and the physiological substrate angiotensin I is preferentially bound to the C-domain catalytic site. The substrate also makes a contribution to the chloride-mediated activation of the active site. Therefore, these differences indicate that despite the higher level of primary sequence homology, structural and functional differences do exist between two active sites of C and N domains. Three subsites, S1 (antepenultimate), S1' (penultimate), and S2' (ultimate), with unshared distinct characteristics for the binding of C-terminal amino acids of substrates or inhibitor are located on two homologous active sites. For the inhibitor–enzyme binding and interaction, three main subsites on the active site of the enzyme with different amino acid sequence should be bound with the substrate. Binding of inhibitor or the natural substrate to the enzyme takes place predominantly via the C-terminal tripeptide residues. The peptides with high ACE inhibitory activity have Trp, Phe, Tyr, or Pro at their C terminus and the branched aliphatic amino acids at the N terminus, and ACE is known to have little affinity toward inhibitors with C-terminal dicarboxylic amino acid, such as Glu [29]. More specifically, the presence of aromatic amino acids, Pro, Ala, Val, and Leu are most favorable for the antepenultimate position (S1), while Ile is most favorable for the penultimate position (S1'). Pro and Leu in the substrate sequence are most favorable for the ultimate position (S2') with regard to the affinity exerted on the enzyme [30,31]. However, there are several peptides such as NNTGHNFTGEAM [32] and WM [28] that do not fit into the model.

### 2.2. Noncompetitive inhibitor

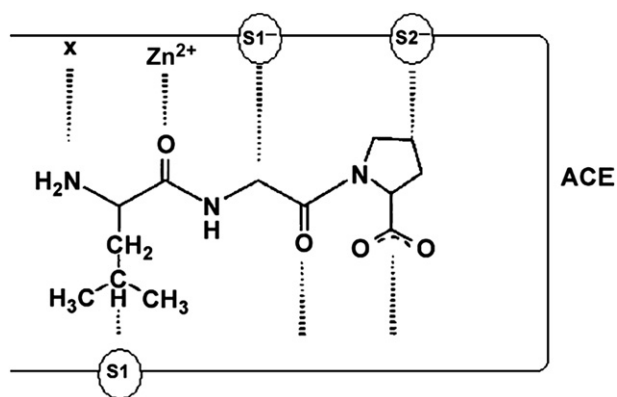
The noncompetitive inhibition system shows that both the inhibitor and the substrate can be bound to the enzyme at any given point of time. When both the substrate and the inhibitor

**Table 1 – Protein-derived ACE inhibitory peptides categorized by inhibition modes.**

Source	Preparation	Peptide	IC <sub>50</sub> (μM)	Reference
1. Competitive inhibitors				
Soy	Alcalase	NLP	4.8	[23]
		NG	12.3	
Hard clam meat	Protamex	YN	51	[24]
Porcine hemoglobin	Pepsin	LGFPPTTKTYFPHF	4.92	[25]
		VVYPWT	6.02	
Milk	Fermentation	LNVPGEIVE	300.1	[26]
		NIPPLTQTPV	173.3	
		DKIHPF	256.8	
Mushroom	Hot water extraction	VIEKYP	129.7	[30]
Chum salmon muscle	Thermolysin	LF	383.2	[27]
		WA	277.3	
		WM	98.6	[31]
		Marine rotifer	Alcalase	
2. Noncompetitive inhibitors				
Tofuyo (fermented soybean)	Fermentation	IFL	44.8	[19]
		WL	29.9	
Hen ovotransferrin	Chymotrypsin	KVREGTTY	102.8	[33]
	ACE	KVREGT	9.1	
Oyster	Pepsin	VVYPWTQRF	66	[34]
Oyster	Trypsin	DLTDY	143	[35]
Algae protein waste	Pepsin	VECYGPNRPQF	29.6	[36]
Yellowfin sole frame	α-Chymotrypsin	MIFPGAGGPEL	26.4	[37]
Tuna frame	Pepsin	GNLGKTTTVSNWSPPKWKNTP	11.28	[12]
Chum salmon muscle	Thermolysin	FL	13.6	[27]
		AW	6.4	
3. Uncompetitive inhibitor				
Wakame	Protease S	IW	1.5	[16]
		FY	42.3	
		AW	18.8	
Human plasma	Trypsin	YLYEIARR	86	[17]
ACE = angiotensin I-converting enzyme; IC <sub>50</sub> = median inhibitory concentration.				

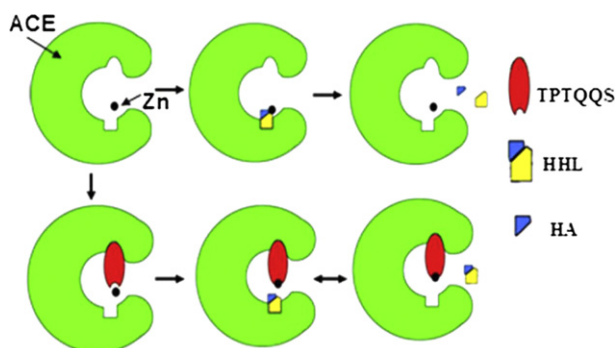
ACE = angiotensin I-converting enzyme; IC<sub>50</sub> = median inhibitory concentration.

are bound, the enzyme–substrate–inhibitor complex cannot form a product but can only be converted back into the enzyme–substrate complex or the enzyme–inhibitor complex [33]. Some food protein–derived peptides were reported to be noncompetitive inhibitors of ACE. These include IFL [19], WL [19], KVREGTTY [34], KVREGT [34], VVYPWTQRF [20], DLTDY [35], VECYGNRPQF [36], and MIFPGAGGPEL [37].



**Fig. 1 – Active site of angiotensin I-converting enzyme (ACE) showing the interactions between ACE inhibitory peptides and ACE [23].**

The inhibition site of these peptides is not specified because of the various structures of the peptides derived from different parent proteins, and the noncompetitive inhibition mechanism of ACE inhibitory peptides is not clear yet. In order to understand the inhibition mechanism, TPTQQS, a hexapeptide that acts as a noncompetitive inhibitor of ACE to prevent the formation of the reaction product His-Ala, was used to investigate the interactions between ACE, TPTQQS, and the nonphysiological substrate, hippuryl-His-Leu [38]. The results obtained showed that when ACE is in the unbound form, the zinc ion and HEXXH (the key amino acid residues of ACE active site) motif compose the complete active site of ACE, and hippuryl-histidyl-leucine can enter the active site and be converted into the reaction product. After TPTQQS enters ACE, the Thr1, Thr3, and Gln4 residues of TPTQQS allow the peptide to interact with the lid structure of testis ACE (tACE), and the C-terminal Ser6 pushes the zinc ion away from the active site through the coordination bonds between the Ser and the zinc ion, resulting in noncompetitive inhibition of ACE by TPTQQS (Fig. 2) [38]. Although the noncompetitive inhibition model of ACE using TPTQQS has been established, the other inhibitory peptides that function as noncompetitive inhibitors may not fit into this model because of their various peptide lengths and composition. Further investigations are being carried out to identify the relationship between the inhibition mechanism



**Fig. 2 – Model of the inhibition of angiotensin I-converting enzyme (ACE) by TPTQQS. The model shows that TPTQQS moves the zinc ion away from the active site to inhibit ACE [38]. HHL = hippuryl-histidyl-leucine.**

and the structure of these peptides. In any case, this was the first study to report the noncompetitive inhibition mechanism of ACE inhibitory peptides, and this study has also provided fresh thoughts for designing drugs or functional foods against enzyme targets.

### 2.3. Uncompetitive inhibitor

In the uncompetitive inhibition system, the inhibitor can bind only to substrate–enzyme complex and decrease the maximum enzyme activity, so that it takes longer for the substrate or product to leave the active site. Peptides such as IW [16], FY [16], AW [16], and YLYEIARR [17] were reported to act as the uncompetitive inhibitors of ACE. However, the inhibition mechanism of this mode is not clear yet.

The fact that ACE is a dipeptidase makes it plausible for further hydrolysis of ACE inhibitory peptides and affects their antihypertensive activity *in vivo*. Depending on the outcome of GI digestion and other enzyme action *in vivo*, ACE inhibitory peptides can be classified as either true inhibitor (i.e., ACE inhibitory activity remains unchanged), prodrug (i.e., increased ACE inhibitory activity), or substrate (i.e., decreased ACE inhibitory activity) types of ACE inhibitors [39]. However, the active sites or these peptides which act as competitive, noncompetitive, or uncompetitive inhibitors are not specified, and the exact inhibition mechanism of ACE inhibitory peptides remains unclear. Further studies are necessary to figure out the correlation between the inhibition mode and the structure of the peptides.

## 3. Bioavailability

In order to administer the ACE inhibitory peptides orally in hypertensive patients, these peptides have to pass through the digestive tract to be absorbed through the intestinal epithelium. Digestion of proteins and peptides starts in the stomach by the action of pepsin at acidic pH, and then the polypeptides are further truncated by the pancreatic proteases, trypsin,  $\alpha$ -chymotrypsin, elastase, and carboxypeptidases A and B at more alkaline pH. The *in vitro* bioavailability of the bioactive peptides is usually tested by sequential

hydrolysis of pepsin and mimicking the actions of pancreatic enzymes so as to simulate the conditions of the GI. The inhibitory activity of some ACE inhibitory peptides was reported to decrease during simulated GI digestion. After sequential treatment with pepsin, chymotrypsin, and trypsin, the median inhibitory concentration ( $IC_{50}$ ) values of IFL and WL (both isolated from tofuyo extract) against ACE varied from 44.8 to 117.9  $\mu$ M and from 29.9 to 103.1  $\mu$ M, respectively [19]. The ACE inhibitory activity of TQVY isolated from rice protein also showed a slight decrease after simulated GI digestion [40]. Some studies also demonstrated that several peptides are resistant to the digestive proteases, such as VIEKYP (from mushroom) [31], LNVPGIVE, NIPPLTQTPV, DKIHPP (from fermented milk) [27], WVPSV, YTVF, VVYPW (from porcine hemoglobin) [41], and IPP, VPP (from  $\beta$ -casein) [42]. Furthermore, DLTDY was hydrolyzed by the simulated GI digestion to a shorter active form, DY, and the  $IC_{50}$  value against ACE decreased from 143  $\mu$ M for DLTDY to 28  $\mu$ M for DY [35]. In one of the studies, egg-derived ACE inhibitory peptides YAEERYPIL and RADHPFL were hydrolyzed to other active forms after simulated GI digestion [43]. There was also a study in which the peptide KVLVPVQ derived from  $\beta$ -casein showed low ACE inhibitory activity ( $IC_{50}$  = 1000  $\mu$ M); however, the shorter peptide KVLVPV obtained after pancreatic digestion showed greater ACE inhibitory activity at the  $IC_{50}$  value of 5  $\mu$ M [44]. In the same study, YKVPQL with strong ACE inhibitory activity ( $IC_{50}$  = 22  $\mu$ M) failed to act as a potent ACE inhibitor with  $IC_{50}$  > 1000  $\mu$ M after pancreatic digestion.

In general, Pro- and hydroxyproline-containing peptides are resistant to degradation by digestive proteases, such that the tripeptides with the C-terminal Pro–Pro are reported to be resistant to Pro-specific peptidases [45,46]. This might be the reason that some ACE inhibitory peptides derived from casein and gelatin have been shown to exert *in vivo* antihypertensive effect, as the Pro content in these two proteins is high.

## 4. Antihypertensive effect

The inhibitory activity of the peptides against ACE does not always correlate with their *in vivo* antihypertensive effects. There are two ways in which the ACE inhibitory peptides exert an antihypertensive effect *in vivo* after oral administration. First, the peptides retain their intact structure and second, they are hydrolyzed into active products, after the action of digestive enzymes, absorbed in the intestine and finally reach their target sites. For this purpose, *in vitro*–simulated GI digestion is a simple and cheap experimental method that is usually used for mimicking the *in vivo* effect. However, the relationship between the *in vitro* ACE inhibitory activity of the peptides via simulated GI digestion and *in vivo* antihypertensive activity is not clear.

An ACE-inhibitory peptide KVLVPVQ derived from casein was degraded to KVLVPV after the simulated GI digestion, and they showed the similar ACE inhibitory activity. After oral administration at a dose of 2 mg/kg, the two peptides KVLVPV and KVLVPVQ got a systolic blood pressure (SBP) reduction of 32.2 and 31.5 mmHg, respectively [44]. In the same study, YLVPQL possessing a strong ACE inhibitory activity was hydrolyzed by simulated GI digestion to YLVP with low ACE

inhibitory activity, and the YLVP showed no significant antihypertensive effect for SHR. Another ACE inhibitory peptide DLTDY with an  $IC_{50}$  value of 143  $\mu$ M was degraded by simulated GI digestion to a dipeptide DL with an  $IC_{50}$  value of 28  $\mu$ M. After both of these ACE inhibitory peptides were orally administrated to SHRs (at dosages of 8 and 10 mg/kg, respectively), their SBPs significantly decreased by about 15 and 20 mmHg at 3–6 and 3 hours [35], respectively. In another study, the ACE inhibitory activity of the peptide TQVY that had an initial  $IC_{50}$  value of 18.2  $\mu$ M slightly decreased after simulated GI digestion. The peptide also showed the antihypertensive activity (by decreasing SBP to 40 mmHg) in SHR when a dosage of 30 mg/kg was administered 6 hours after the oral administration [40].

However, some studies showed inconsistent results. In one of the studies, two peptides YPI and RADHP obtained from the ACE inhibitory peptides YAEERYPI and RADHPFL, respectively, by simulated GI digestion were not as active as ACE inhibitors ( $IC_{50} > 1000$  and = 153  $\mu$ g/mL), but showed a great antihypertensive effect on SHR at a dosage of 2 mg/kg 2–4 hours after oral administration (SBP: 31.6 and 34.0 mmHg, respectively) [43]. In another study, three ACE inhibitory peptides WVPSV, YTVF, and VVYPW had initial  $IC_{50}$  values of 0.368, 0.226, and 0.254 mg/mL, respectively [41]. After the simulated GI digestion, the ACE inhibitory activity of WVPSV and YTVF slightly increased, while that of VVYPW remained almost unchanged. Three hours after the oral administration of the former peptides, the SBPs of SHRs decreased by 22.5 and 18.5 mmHg, respectively, whereas after administration of the latter peptide it decreased only by 9.6 mmHg. The possible explanation is that these peptides need further activation by intestinal brush-border or plasma peptidases [43].

In fact, it is difficult to establish a model that directly correlates with *in vitro* ACE inhibitory activity and *in vivo* antihypertensive activity of peptides. The first reason is the bioavailability after oral administration, and the other is that some other antihypertensive mechanisms other than ACE inhibition may be of interest. Although the ACE inhibitory activity of the peptides through *in vitro*–simulated GI digestion may not always ensure their *in vivo* antihypertensive activity, simulated GI digestion is still a good method for preliminary test that is carried out to understanding the possible changes of the peptide structure and screening the potent antihypertensive peptides. Some studies have demonstrated the existence of vasorelaxant peptides that exert their effects through the stimulation of opioid receptors [47]. Moreover, these peptides exhibited a direct or indirect action on vascular smooth muscles [48,49]. In addition, strong evidence indicates that oxidative stress and associated oxidative damage are mediators in cardiovascular pathologies, and thus, antioxidant activity can also be responsible for antihypertensive effects [50].

## 5. Conclusions and future perspectives

Much work has been done with food protein–derived ACE inhibitory peptides and evidence of their *in vivo* antihypertensive effect has been built in animal and clinical studies. However, the ACE inhibition mechanisms of the peptides with

different inhibition modes are still vague due to the various lengths and sequences of these peptides derived from different parent proteins. It is difficult to establish a model and a rule to describe the structure–activity relationship. “BIOPEP” database (<http://www.uwm.edu.pl/biochemia/index.php/en/biopep>) is responsible for collecting all the information about bioactive peptides from academic literature, and currently the database lists the sequences of 556 ACE inhibitory peptides. Further studies on the establishment of the model of structure–activity relationship are necessary, which can be achieved by using computing methods.

The most challenging task in the antihypertensive peptide researches is the establishment of a detection model for the identification of possible mechanisms by which they can exert *in vivo* antihypertensive activity. Although the simulated GI digestion is a kind of model mimicking the actions of human GI enzymes, the antihypertensive effect of the peptides released from the ACE inhibitory peptides through simulated GI digestion is not guaranteed to be similar to that observed in the GI tract *in vivo*. Therefore, the possible strategies for increasing the resistance to digestive enzymes and cellular permeability of antihypertensive peptides should be also investigated.

## REFERENCES

- [1] Kearney PM, Whelton M, Reynolds K, Muntner P, Whelton PK, He J. Global burden of hypertension: analysis of worldwide data. *Lancet* 2005;365:217–23.
- [2] Brunner HR, Laragh JH, Baer L, Newton MA, Goodwin FT, Krakoff LR, et al. Essential hypertension: renin and aldosterone, heart attack and stroke. *N Engl J Med* 1972;286: 441–9.
- [3] Ondetti MA, Rubin B, Cushman DW. Design of specific inhibitors of angiotensin-converting enzyme: new class of orally active antihypertensive agents. *Science* 1977;4288: 441–4.
- [4] Patchett AA, Harris E, Tristram EW, Wyvratt MJ, Wu MT, Taub D, et al. A new class of angiotensin-converting enzyme inhibitors. *Nature* 1980;298:280–3.
- [5] Oshima G, Shimabukuro H, Nagasawa K. Peptide inhibitors of angiotensin I-converting enzyme in digests of gelatin by bacterial collagenase. *Biochim Biophys Acta* 1979;566: 128–37.
- [6] Mullally MM, Meisel H, FitzGerald RJ. Identification of a novel angiotensin-I-converting enzyme inhibitory peptide corresponding to a tryptic fragment of bovine beta-lactoglobulin. *FEBS Lett* 1997;402:99–101.
- [7] Yano S, Suzuki K, Funatsu G. Isolation from alpha-zein of thermolysin peptides with angiotensin I-converting enzyme inhibitory activity. *Biosci Biotechnol Biochem* 1996;60:661–3.
- [8] Megías C, del Mar Yust M, Pedroche J, Lquari H, Girón-Calle J, Alaiz M, et al. Purification of an ACE inhibitory peptide after hydrolysis of sunflower (*Helianthus annuus* L.) protein isolates. *J Agric Food Chem* 2004;52:1928–32.
- [9] Yoshikawa M, Fujita H, Matoba N, Takenaka Y, Yamamoto T, Yamauchi R, et al. Bioactive peptides derived from food proteins preventing lifestyle-related diseases. *Biofactors* 2000;12:143–6.
- [10] Matsui T, Li CH, Osajima Y. Preparation and characterization of novel bioactive peptides responsible for angiotensin I-converting enzyme inhibition from wheat germ. *J Pept Sci* 1999;5:289–97.



- [11] Je JY, Park JY, Jung WK, Park PJ, Kim SK. Isolation of angiotensin I converting enzyme (ACE) inhibitor from fermented oyster sauce, *Crassostrea gigas*. Food Chem 2005; 90:809–14.
- [12] Lee SH, Qian ZJ, Kim SK. A novel angiotensin I converting enzyme inhibitory peptide from tuna frame protein hydrolysate and its antihypertensive effect in spontaneously hypertensive rats. Food Chem 2010;118:96–102.
- [13] Zhao Y, Li B, Dong S, Liu Z, Zhao X, Wang J, et al. A novel ACE inhibitory peptide isolated from *Acaudina molpadioides* hydrolysate. Peptides 2009;30:1028–33.
- [14] Qian ZJ, Je JY, Kim SK. Antihypertensive effect of angiotensin I converting enzyme-inhibitory peptide from hydrolysates of Bigeye tuna dark muscle, *Thunnus obesus*. J Agric Food Chem 2007;55:8398–403.
- [15] Suetsuna K, Nakano T. Identification of an antihypertensive peptide from peptic digest of wakame (*Undaria pinnatifida*). J Nutr Biochem 2000;11:450–4.
- [16] Sato M, Hosokawa T, Yamaguchi T, Nakano T, Muramoto K, Kahara T, et al. Angiotensin I-converting enzyme inhibitory peptides derived from wakame (*Undaria pinnatifida*) and their antihypertensive effect in spontaneously hypertensive rats. J Agric Food Chem 2002;50:6245–52.
- [17] Nakagomi K, Fujimura A, Ebisu H, Sakai T, Sadakane Y, Fujii N, et al. Acein-1, a novel angiotensin-I-converting enzyme inhibitory peptide isolated from tryptic hydrolysate of human plasma. FEBS Lett 1998;438:255–7.
- [18] Samaranayaka AGP, Kitts DD, Li-Chan ECY. Antioxidative and angiotensin-I-converting enzyme inhibitory potential of a Pacific hake (*Merluccius productus*) fish protein hydrolysate subjected to simulated gastrointestinal digestion and Caco-2 cell permeation. J Agric Food Chem 2010;58:1535–42.
- [19] Kuba M, Tanaka K, Tawata S, Takeda Y, Yasuda M. Angiotensin I-converting enzyme inhibitory peptides isolated from tofuyo fermented soybean food. Biosci Biotechnol Biochem 2003;67:1278–83.
- [20] Wang JP, Hu J, Cui J, Bai X, Du Y, Miyaguchi Y, et al. Purification and identification of a ACE inhibitory peptide from oyster proteins hydrolysate and the antihypertensive effect of hydrolysate in spontaneously hypertensive rats. Food Chem 2008;111:302–8.
- [21] Meisel H. Biochemical properties of regulatory peptides derived from milk proteins. Biopolymers 1997;43:119–28.
- [22] Ferreira SH, Bartelt DC, Greene LJ. Isolation of bradykinin-potentiating peptides from *Bothrops jararaca* venom. Biochemistry 1970;9:2583–93.
- [23] Hong F, Ming L, Yi S, Zhanxia L, Yongquan W, Chi L. The antihypertensive effect of peptides: a novel alternative to drugs? Peptides 2008;29:1062–71.
- [24] Wu J, Ding X. Characterization of inhibition and stability of soy-protein-derived angiotensin I-converting enzyme inhibitory peptides. Food Res Int 2002;35:367–75.
- [25] Tsai JS, Chen JL, Pan BS. ACE-inhibitory peptides identified from the muscle protein hydrolysate of hard clam (*Meretrix lusoria*). Process Biochem 2008;43:743–7.
- [26] Yu Y, Hu J, Miyaguchi Y, Bai X, Du Y, Lin B. Isolation and characterization of angiotensin I-converting enzyme inhibitory peptides derived from porcine hemoglobin. Peptides 2006;27:2950–6.
- [27] Gobetti M, Ferranti P, Smacchi E, Goffredi F, Addeo F. Production of angiotensin-I-converting-enzyme-inhibitory peptides in fermented milks started by *Lactobacillus delbrueckii* subsp. *bulgaricus* SS1 and *Lactococcus lactis* subsp. *cremoris* FT4. Appl Environ Microbiol 2000;66:3898–904.
- [28] Ono S, Hosokawa M, Miyashita K, Takahashi K. Inhibition properties of dipeptides from salmon muscle hydrolysate on angiotensin I-converting enzyme. Int J Food Sci Technol 2006;41:383–6.
- [29] Li GH, Le GW, Shi YH, Shrestha S. Angiotensin I – converting enzyme inhibitory peptides derived from food proteins and their physiological and pharmacological effects. Nutr Res 2004;24:469–86.
- [30] Ondetti MA, Cushman DW. Enzymes of the renin-angiotensin system and their inhibitors. Annu Rev Biochem 1982;51:283–308.
- [31] Choi HS, Cho HY, Yang HC, Ra KS, Suh HJ. Angiotensin I-converting enzyme inhibitor from *Grifola frondosa*. Food Res Int 2001;34:177–82.
- [32] Lee JK, Hong S, Jeon JK, Kim SK, Byun HG. Purification and characterization of angiotensin I converting enzyme inhibitory peptides from the rotifer, *Brachionus rotundiformis*. Bioresour Technol 2009;100:5255–9.
- [33] Si D, Wang Y, Zhou YH, Guo Y, Wang J, Zhou H, et al. Mechanism of CYP2C9 inhibition by flavones and flavonols. Drug Metab Dispos 2009;37:629–34.
- [34] Lee NY, Cheng JT, Enomoto T, Nakano Y. One peptide derived from hen ovotransferrin as pro-drug to inhibit angiotensin converting enzyme. J Food Drug Anal 2006;14: 31–5.
- [35] Shiozaki K, Shiozaki M, Masuda J, Yamauchi A, Ohwada S, Nakano T, et al. Identification of oyster-derived hypotensive peptide acting as angiotensin-I-converting enzyme inhibitor. Fish Sci 2010;76:865–72.
- [36] Sheih IC, Fang TJ, Wu TK. Isolation and characterization of a novel angiotensin I-converting enzyme (ACE) inhibitory peptide from the algae protein waste. Food Chem 2009;115: 279–84.
- [37] Jung WK, Mendis E, Je JY, Park PJ, Son BW, Kim HC, et al. Angiotensin I-converting enzyme inhibitory peptide from yellowfin sole (*Limanda aspera*) frame protein and its antihypertensive effect in spontaneously hypertensive rats. Food Chem 2006;94:26–32.
- [38] Ni H, Li L, Liu G, Hu SQ. Inhibition mechanism and model of an angiotensin I-converting enzyme (ACE)-inhibitory hexapeptide from yeast (*Saccharomyces cerevisiae*). PLoS One 2012;7:e37077.
- [39] Fujita H, Yokoyama K, Yoshikawa M. Classification and antihypertensive activity of angiotensin I-converting enzyme inhibitory peptides derived from food proteins. J Food Sci 2000; 65:564–9.
- [40] Li GH, Qu MR, Wan JZ, You JM. Antihypertensive effect of rice protein hydrolysate with *in vitro* angiotensin I-converting enzyme inhibitory activity in spontaneously hypertensive rats. Asia Pac J Clin Nutr 2007;16(Suppl. 1): 275–80.
- [41] Ren Y, Wan D-G, Lu X-M, Chen L, Zhang T, Guo J- L. Isolation and characterization of angiotensin I-converting enzyme inhibitor peptides derived from porcine hemoglobin. Sci Res Essays 2011;6:6262–9.
- [42] Ohsawa K, Satsu H, Ohki K, Enjoh M, Takano T, Shimizu M. Producibility and digestibility of antihypertensive  $\beta$ -casein tripeptides, Val-Pro-Pro and Ile-Pro-Pro, in the gastrointestinal tract: analyses using an *in vitro* model of mammalian gastrointestinal digestion. J Agric Food Chem 2008;56:854–8.
- [43] Miguel M, Aleixandre MA, Ramos M, López-Fandiño R. Effect of simulated gastrointestinal digestion on the antihypertensive properties of ACE-inhibitory peptides derived from ovalbumin. J Agric Food Chem 2006;54:726–31.
- [44] Maeno M, Yamamoto N, Takano T. Identification of an antihypertensive peptide from casein hydrolysate produced by a proteinase from *Lactobacillus helveticus* CP790. J Dairy Sci 1996;79:1316–21.
- [45] Vanhoof G, Goossens F, De Meester I, Hendriks D, Scharpé S. Proline motifs in peptides and their biological processing. FASEB J 1995;9:736–44.

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- [46] FitzGerald RJ, Meisel H. Milk protein-derived peptide inhibitors of angiotensin-I-converting enzyme. *Br J Nutr* 2000;84(Suppl. 1):S33–7.
- [47] Sipola M, Finckenberg P, Vapaatalo H, Pihlanto-Leppälä A, Korhonen H, Korpela R, et al.  $\alpha$ -Lactorphin and  $\beta$ -lactorphin improve arterial function in spontaneously hypertensive rats. *Life Sci* 2002;71:1245–53.
- [48] Kouno K, Hirano S, Kuboki H, Kasai M, Hatae K. Effects of dried bonito (*katsuobushi*) and captopril, an angiotensin I-converting enzyme inhibitor, on rat isolated aorta: a possible mechanism of antihypertensive action. *Biosci Biotechnol Biochem* 2005;69:911–5.
- [49] Maes W, Van Camp J, Vermeirssen V, Hemeryck M, Ketelslegers JM, Schrezenmeir J, et al. Influence of the lactokinin Ala-Leu-Pro-Met-His-Ile-Arg (ALPMHIR) on the release of endothelin-1 by endothelial cells. *Regul Pept* 2004;118:105–9.
- [50] Touyz RM. Reactive oxygen species, vascular oxidative stress, and redox signaling in hypertension: what is the clinical significance? *Hypertension* 2004;44:248–52.


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## Review article

# Therapeutic approaches to drug targets in hyperlipidemia

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## ABSTRACT

Hyperlipidemia is a metabolic syndrome characterized by diverse lipid profiles (e.g. hypercholesterolemia, hypertriglyceridemia, and familial combined hyperlipidemia) and may have significant adverse effects on health (e.g. atherosclerosis, cardiovascular diseases, diabetes, insulin resistance, obesity). Both genetic and environmental components are associated with hyperlipidemia sub-types. Effective drugs targeting hyperlipidemia sub-types are thus required. In the present review, we mainly focus on types of hyperlipidemia, digestion, and absorption of lipids as well as on their consequences on human health and on potential effective drug targets against hyperlipidemia. Omega-3 fatty acids have favorable effect on reducing postprandial triglyceride levels and will be beneficial if combined with statins.

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## 1. Introduction

Hyperlipidemia is a heterogeneous disorder commonly characterized by an increased flux of free fatty acids (FFAs), raised triglycerides (TGs), low-density lipoprotein-cholesterol (LDL-c) (aka “bad cholesterol”) and apolipoprotein B (apoB) levels, as well as by a reduced plasma high-density lipoprotein (HDL)-cholesterol concentration (aka “good cholesterol”), because of metabolic effects, or dietary and lifestyle habits [1]. The lipid abnormality in hyperlipidemia is an increase in circulating (nonesterified) FFAs originating from adipose tissue, and an inadequate esterification and FFA metabolism [2]. The reduced retention of fatty acids (FAs) by adipose tissue leads to an increased flux of FFA returning to the liver, which stimulates

hepatic TG synthesis, promoting the production of apoB and the assembly and secretion of very low-density lipoprotein (VLDL). When plasma TG concentration subsequently increased, TG-rich HDL particles are formed and undergo catabolism. Elevated VLDL particles are lysed and hence fail to bind efficiently to LDL receptors, while the exchange of cholesterol esters with TGs forms TG-rich lipoproteins, resulting in formation of small dense LDL-c particles [3,4]. A strong association exists between elevated LDL-c levels and increased incidence of coronary artery disease [5]. The development of atherosclerotic plaques is associated with elevated levels of LDL-c, reduced receptor-mediated clearance, increased arterial wall retention and an increased susceptibility [6]. Cardiovascular risk factors such as hyperlipidemia,

**Abbreviations:** ACAT, Acyl-Co A: cholesterol acyltransferase; AMPK, AMP-activated protein kinase; apoB, Apolipoprotein B; ATP III, adult Treatment Panel III; CETP, cholesteryl ester transfer protein; CM, chylomicrons; DGAT, diacylglycerol acyltransferase; FCH, familial combined hyperlipidemia; FFA, free fatty acid; HC, hypercholesterolemia; HDL, high-density lipoprotein; HTG, hypertriglyceridemia; LDL, low density lipoprotein; NCEP, National Cholesterol Education Program; PPAR, peroxisome proliferator-activated receptors; TG, triglyceride; VLDL, very low density lipoprotein.

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hypertension, and thrombosis contribute to the underlying mechanisms of atherosclerotic disease, promoting endothelial dysfunction, oxidative stress, and proinflammatory pathways to peroxidation [4,6]. Lipid guidelines from the National Heart Foundation of Australia place great emphasis on LDL-c and HDL-c as atherogenic and antiatherogenic components, respectively. Indeed, high LDL-cholesterolemia is considered as one of the major modifiable risk factors for coronary heart disease, which continues to be the leading cause of death and morbidity in the United States [7]. Conversely to the Australian lipid guidelines, the Adult Treatment Panel III (ATP III) guidelines of the US National Cholesterol Education Program (NCEP) place greater emphasis on TG levels [4,8]. According to the National Health and Nutrition Examination Survey III, 24% of individuals aged >20 years had metabolic syndrome [9]. Metabolic syndrome is characterized by the coexistence of hyperinsulinemia, obesity, dyslipidemia, and hypertension. Dyslipidemia, the hallmark of the metabolic syndrome, is summarized by: (1) increased flux of FFAs; (2) raised TG values; (3) low HDL-c values; (4) increased of LDL-c values; and (5) raised apoB values [10]. Dyslipidemia is an independent risk factor for cardiovascular disease [11]. Low HDL-c and hypertriglyceridemia (HTG) have been found to be independently and significantly related to myocardial infarction/stroke in patients with metabolic syndrome [12]. The combination of high fasting glucose and low HDL-c were shown to have primary predictive ability for coronary heart disease [13]. Dyslipidemia may be caused by a combination of overproduction of VLDL, apoB-100, decreased catabolism of apoB containing particles, and increased catabolism of HDL-apoA-I particles. Insulin resistance may be the consequence of this abnormality [1]. Dyslipidemia may arise from genetic components (e.g. mutated LDL receptors, mutated apoB-100, mutated proprotein convertase subtilisin/kexintype-9) [14], with or without environmental component (e.g. improper diet, familial history of hypercholesterolemia, hyperlipidemia, and/or hypertriglyceridemia) [15]. Causes of secondary hyperlipidemia include diabetes, hypothyroidism, obstructive liver disease, chronic renal failure, and drugs that increase LDL cholesterol and decrease HDL cholesterol, such as progestine and corticosteroids [16].

## 2. Digestion and absorption of lipids

Lipid digestion begins in the oral cavity by the use of lingual lipase, an enzyme secreted by lingual gland in the tongue, and continues in the stomach with the both lingual and gastric enzymes. Lipids undergo emulsification in the stomach under the influence of peristalsis. Fine lipid droplets enter the duodenum, where they mix with bile and pancreatic juice to undergo marked changes in physical and chemical form. For absorption across the intestinal walls, hydrolysis and micellization take place in duodenum [17,18]. Diacylglycerol and FFAs are the major digestion products of this gastric phase, and facilitate the intestinal phase of digestion acting as emulsifying agents [19]. Pancreatic lipase cleaves the TG, yielding 2-monoglycerides (2-MGs) and FFAs. Pancreatic cholesterol esters hydrolase completely hydrolyzes cholesterol esters into FFAs and free cholesterol [20]. Dietary

phospholipids are hydrolyzed by activated pancreatic phospholipase A2, yielding 1-lysophospholipids and FFAs [21]. FFAs and 2-MGs enter into bile micelles, which helps polar lipids to go through the unstirred water layer and reach the microvillous membrane where they are absorbed. Absorbed lipids are re-esterified to newly form TGs and in the smooth endoplasmic reticulum (ER). TGs can be synthesized via 2-MG or via 3-glycerol-phosphate. TGs, phospholipids, cholesterol, and apoproteins are used to synthesize chylomicrons (CMs), which are secreted to the lymph, and then to the blood stream through the thoracic duct. In the peripheral tissues, they are cleaved by lipoprotein lipase losing TG and giving CM remnants, which are taken up by the liver [21–23].

## 3. Hyperlipidemia profiles/sub-types

The classification of hyperlipidemia according to WHO is in Table 1 [17] and the constitution, composition and role of lipids in Table 2 [23].

### 3.1. HTG

Plasma TGs represent an important mechanism of whole body fatty acid delivery for tissue utilization or storage [23,24]. HTG is defined as an abnormally high concentration of TG in the blood. According to the NCEP ATP III guidelines, a normal TG level is <150 mg/dL [17]. In the United States, the prevalence of HTG, defined as a TG level >150 mg/dL, is 30%.

HTG is a risk factor for pancreatitis and it accounts for 1% to 4% of cases of acute pancreatitis [25]. HTG may be primary or secondary in nature. Primary HTG is the result of various genetic defects while the secondary causes are high fat diet, obesity, diabetes, hypothyroidism, and certain medications [26].

## 4. Patterns of HTG

Familial HTG is commonly seen in clinical practice and can have various lipid patterns [27].

### 4.1. Hyperlipoproteinemia

Most commonly, patients demonstrate type IV hyperlipoproteinemia, which includes elevated TG levels (250–500 mg/dL) and elevated VLDL levels that transport them, whereas normal LDL-c and apoB levels are observed [28].

### 4.2. Chylomicronemia syndrome

Familial chylomicronemia syndrome is a rare disorder of lipoprotein metabolism due to familial lipoprotein lipase (LPL) or apolipoprotein C-II deficiency or the presence of inhibitors to lipoprotein lipase [29]. The chylomicronemia syndrome is a disorder characterized by severe HTG and massive accumulation of CMs in plasma [30]. Finally, HTG may contribute to additional pathologic processes associated with metabolic syndrome and cardiovascular risk, including increased

**Table 1 – World Health Organization (modified Fredrickson) classification of hyperlipidemias [17].**

Type	Total cholesterol	LDL cholesterol	Plasma TGs	Lipoprotein abnormality	Primary causes	Secondary causes
I	Elevated	Low or normal	Elevated	Excess chylomicrons	Lipoprotein lipase deficiency, apoC-II deficiency	Systemic lupus erythematosus
II a	Elevated or normal	Elevated	Normal	Excess LDL	Familial hypercholesterolemia	Hypothyroidism
II b	Elevated	Elevated	Elevated	Excess LDL and VLDL	Familial combined hyperlipidemia	Nephrotic syndrome, diabetes, anorexia nervosa
III	Elevated	Low or normal	Elevated	Excess chylomicron remnants and Intermediate density lipoproteins	Familial type III Hyperlipoproteinemia	Hypothyroidis diabetes, obesity
IV	Elevated or normal	Normal	Elevated	Excess VLDL	Familial combined hyperlipidemia, Familial Hypertriglyceridemia	Diabetes, chronic renal diseases
V	Elevated	Normal	Elevated	Excess chylomicrons and VLDL	Familial hypertriglyceridemia, apoC-II deficiency	Alcohol, diuretics, $\beta$ blockers, oral

apoC-II = apolipoprotein-C II; LDL = low-density lipoprotein; TG = triglyceride; VLDL = very low-density lipoprotein.

coagulability, impaired fibrinolysis, impaired endothelial function, and increased inflammation, although this remains uncertain [27,28,31].

#### 4.3. HC

HC is one of the major causes of atherosclerosis and characterized by elevation of total cholesterol and usually normal levels of TG [32,33]. The population is considered to be unhealthy when its plasma concentration exceeds 5 mM and the incidence of CHD is usually low where plasma cholesterol concentration is low [34]. HC usually results from nutritional factors such as obesity and diet high in saturated fats along with genetic causes. The deficiency of adaptor protein Dab 2, or the clathrin coat adaptor AP-2 also leads to HC [32,35]. Patients with HC have plasma TG concentration of >10 mM owing to increase in both CMs and VLDL and, in such patients, plasma shows milky appearance [32]. Familial HC comprises a group of genetic disorders characterized by elevated plasma concentrations of LDL-c and premature cardiovascular disease due to a defective (mainly hepatic) metabolism of LDL [36]. Major genetic backgrounds of familial HC include loss-of-function mutations in the genes of LDL receptor, its ligand

apoB or, gain-of-function mutations in the facilitator gene for hepatic LDL receptor degradation, the proprotein convertase subtilisin/kexin type-9 [35,37]. LDL receptors are predominantly found on hepatocytes and steroid hormone producing cells and are responsible for removal of cholesterol carrying LDL from plasma by a process of receptor-mediated endocytosis. The most important feature of untreated familial HC is the development of premature and extensive atherosclerosis leading to coronary artery diseases [38].

#### 4.4. Familial combined hyperlipidemia

Familial combined hyperlipidemia (FCH) is the most common genetic hyperlipidemia in man and affects up to 5% of the general population [39]. HC, HTG, and elevated levels of apoB are the characteristics of FCH [40]. Other phenotypes of FCH are elevated levels of both LDL-c and VLDL, the presence of small dense LDL, and decreased levels of HDL-c. In addition, FCH is associated with obesity and insulin resistance [41,42]. Obesity results in an increase in number and size of adipocytes, which secrete leptin, a hormone involved in the regulation of the energy expenditure and appetite via hypothalamic receptors [43,44]. Both obesity and insulin

**Table 2 – The constitution, composition, and role of lipids [23].**

Constituent	Composition	Effect/role
Lipoproteins	95% TG and 5% cholesterol	Mobilize dietary lipids, deliver dietary triglycerides to adipose tissues, muscles and dietary
Chylomicrons	80% TG and 20% cholesterol	cholesterol to liver
VLDL	50% TG and 50% cholesterol	Transport triglycerols to extra hepatic tissues
IDL	10% TG and 90% cholesterol	They are either converted to LDL or taken up by the liver
LDL	5% TG and 95% cholesterol	Principal plasma carriers of cholesterol for delivering to peripheral tissues
HDL		The apolipoprotein-E in HDLs leads to an increase uptake of cholesterol and its catabolism by the liver to lower the levels of intracellular cholesterol

HDL = high-density lipoprotein; IDL = intermediate-density lipoprotein; LDL = low-density lipoprotein; TG = triglyceride; VLDL = very low-density lipoprotein.

resistance are characteristics of FCH, and therefore, it is likely that leptin is elevated in people with FCH [45]. Recently, in a large observational study, the calculated plasma non-HDL-c concentration was a stronger predictor of cardiovascular events than plasma cholesterol alone [46,47]. Partially lipolyzed TRL remnants (i.e. remnant-like particle cholesterol) are considered to be more atherogenic than larger newly secreted TRL because they can more readily penetrate the endothelial lining of the arterial wall [48]. In the metabolic syndrome, elevated levels of remnant-like particle cholesterol were a risk factor for cardiovascular disease and endothelial dysfunction, a predictor of coronary events [39,49]. FCH might be the result of a combination of an increased production of VLDL particles together with disturbances in their lipoprotein catabolism, such as a decreased LPL-activity [41]. The resulting partially hydrolyzed TG-rich remnant particles are more atherogenic than larger (newly-secreted) TG-rich lipoprotein particles, since the particles are smaller and thereby able to penetrate the endothelial barrier more easily [48,50]. A striking feature of FCHL is the presence of small and dense LDL particles, possibly consequent to hepatic overproduction of apoB [51–53]. Nondenaturing polyacrylamide gradient gel electrophoresis, which separates lipoprotein particles according to their size, has shown that the majority of the population can be characterized into two distinct, genetically determined, LDL subclass phenotypes [48]. Phenotype A is the most common phenotype and is found in individuals with a predominance of large LDL particles, whereas those with a predominance of small LDL particles have phenotype B [54]. Phenotype B often coexists with other lipoprotein abnormalities, notably raised plasma TGs and low HDL-c, in a condition that has been called 'atherogenic lipoprotein phenotype' [55]. Several regions on chromosomes including 2p, 6q, 8p, 9p, 10p, 11p, 16q, 19q, and 21q, have been reported to be associated with FCH [56]. The association of upstream stimulatory factor 1 with FCH however, was strongest in males with increased levels of TGs [57].

## 5. Current drug targets against hyperlipidemia

Conventional therapy for hyperlipidemia is as listed in Table 3 [28,58–62].

### 5.1. Activators of peroxisome proliferator-activated receptor

The peroxisome proliferator-activated receptors (PPARs) are members of the nuclear receptor superfamily that function as fatty acid-activated transcription factors [63]. PPARs are regulators of numerous metabolic pathways; hence there is huge increase in the development and use of agonists of these receptors as therapeutics for diabetes, dyslipidemia, and atherosclerosis [64]. Three different PPAR genes ( $\alpha$ ,  $\beta/\delta$ , and  $\gamma$ ) have been identified, each isotype displaying distinct patterns of tissues distribution and specific pharmacological activators, performing their distinct functions in different cell types [65]. PPAR $\alpha$  is mostly expressed in the tissues involved in lipid oxidation, such as liver, kidney, skeletal, cardiac

muscle, and adrenal glands. PPAR $\alpha$  potentiates FAs oxidation in the liver, heart, kidney, and skeletal muscle. Activation of PPAR $\alpha$  leads to an increase in expression of lipoprotein lipase and apoA-V and to a decrease in hepatic apoC-III. These actions lower plasma TGs in chylomicrons and VLDL particles, thus liberating FAs, which are taken up and stored as fat in adipocytes or metabolized in skeletal muscle [66]. In addition, PPAR $\alpha$  activation increases hepatic apoA-I and -II expression, which raises HDL cholesterol levels, and promotes HDL-mediated cholesterol efflux from macrophages by inducing ATP-binding cassette A1 transporter [67]. PPAR $\gamma$  is expressed in adipose tissue, macrophages, and vascular smooth muscles, while PPAR $\delta$  is mainly expressed in skeletal muscle and adipose tissues [68]. PPAR $\beta/\delta$  is best known for its role in skin homeostasis, and has recently been shown to play a role in HDL metabolism [64]. A combination of PPAR $\alpha$  and PPAR $\gamma$  agonists would be expected to achieve beneficial effects on restoring metabolic disorders. Hence, a number of PPAR $\alpha/\gamma$  dual agonists have been designed and developed. However, recently identified PPAR $\alpha/\gamma$  dual agonists were ineffective because of undesirable side effects during preclinical or clinical trials. For example, muraglitazar, a synthetic PPAR $\alpha/\gamma$  dual agonist, was aborted during clinical trials because of increased mortality, fluid retention, edema, and cancer [69]. PPAR $\alpha$  regulates genes involved in FA uptake,  $\beta$ -oxidation, and  $\omega$ -oxidation and down-regulates apolipoprotein C-III, a protein that inhibits TG hydrolysis by lipoprotein lipase, and it also regulates genes involved in reverse cholesterol transport, such as apolipoprotein A-I and A-II [68]. PPAR $\alpha$  and PPAR $\gamma$  are the molecular targets of number of marketed drugs such as fibrates, the activator of PPAR  $\alpha$  and the thiazolidinediones, the activators of PPAR  $\gamma$  [59].

### 5.2. Cholesteryl ester transfer protein inhibitors

Cholesteryl ester transfer protein (CETP) is a plasma glycoprotein that facilitates the movement of cholesteryl esters and triglycerides between the various lipoproteins in the blood by mediating the transfer of cholesteryl esters from the cardioprotective HDL-c to the proatherogenic LDL-c and VLDL-c [70]. Thus, the movement of cholesteryl esters from HDL-c to LDL-c by CETP has the overall undesirable effect of lowering HDL-c. It therefore follows that inhibition of CETP should lead to elevation of plasma HDL-c and lowering of plasma LDL-c, thereby providing a therapeutically beneficial plasma lipid profile [71]. Elevation in HDL levels is equally favored by diminished CETP-mediated transfer of CE and HDL to atherogenic acceptor lipoproteins (i.e. VLDL, LDL). Elevated CETP activity is a major player whose action underlies the atherogenic particle profile of both LDL and HDL in Type II diabetes [72]. Inhibition of CETP, a key protein involved in reverse cholesterol transport, can consequently lead to increases in HDL-c levels and thus, is under evaluation as an anti-atherogenic strategy. To date, anacetrapib demonstrates the greatest HDL-c raising and LDL-c lowering potential [73]. There are three CETP inhibitors that have been used in clinical trials. Torcetrapib was the first to go into human trials but was discontinued in Phase III because of excessive rates of mortality in the ILLUMINATE (investigation of lipid level

**Table 3 – Pharmacotherapy of hyperlipidemia [28,58–62].**

Drugs	Mechanism of action	Use	Effect on lipoproteins	Adverse effects
Statins Lovastatin (20–80 mg) Pravastatin (20–40 mg) Simvastatin (20–80 mg) Atorvastatin (10–80 mg) Fluvastatin (20–80 mg)	By inhibiting conversion of 3-hydroxy-3-methylglutaryl-coenzyme A-CoA to mevalonate.	Type IIa	LDL decreases 18–55% HDL increases 5–15% TG decreases 7–30%	SGOT, SGPT, Myositis, Lens opacity, Myopathy, Headache GI complaints, Increase liver enzymes Rhabdomyolysis Impaired cognitive function
Bile acid sequestrants Cholestyramine (4–16 g) Colestipol (5–20 g) Colesevelam (2.6–3.8 g)	By interrupting enterohepatic recycling of bile acids. FXR mediated CYP7A repression	Type IIa	LDL decreases 15–30% HDL increases 3–5% TG no change or increases	Constipation and bloating, Hemorrhoidal bleeding Dry flaking skin Gallstone Myopathy Flatulence
Fibric acid derivatives Gemfibrosil (600 mg) Fenofibrate (200 mg) Clofibrate (1000 mg)	Increase lipolysis of triglycerides via lipoprotein lipase. Act as agonist for PPAR- $\alpha$ , resulting in increased expression of lipoprotein lipase and inhibition of apolipoprotein-C-III gene transcription	Types III and IV	LDL decreases 5–20% HDL increases 10–20% TG decreases 20–50%	SGOT, SGPT, Myositis Gallstone Arrhythmias
Nicotinic acid Immediate release (1.5–3 g) Extended release (1–2 g) Sustained release (1–2 g)	By decreasing flux of FFA to the liver. Through Gi coupled receptor (GPR109A, PUMA-G, HM74) By noncompetitive blocking of DGAT2	Types IIa and IV	LDL decreases 5–25% HDL increases 15–35% TG decreases 20–50%	Flushing SGOT, SGPT Tachycardia Pruritus Glucose intolerance Hyperuricemia Nausea Diarrhea Hepatotoxicity

DGAT = diacylglycerol acyltransferase; FFA = free fatty acid; HDL = high-density lipoprotein; LDL = low-density lipoprotein; PPAR = peroxisome proliferator-activated receptor; SGOT = serum glutamic oxaloacetic transaminase; SGPT = serum glutamic pyruvate transaminase; TG = triglyceride; VLDL = very low-density lipoprotein.

management to understand its impact in atherosclerotic events) trial. Anacetrapib, which has a similar structure to torcetrapib but does not share its properties when it comes to the effects on aldosterone production, is presently in Phase III research. Dalcetrapib, which is structurally different than torcetrapib, is currently undergoing cardiovascular outcomes trials [74]. 2-Arylbenzoxazole, [75], tetrahydroquinoline (BAY 38-1335) [76], chromanol derivatives, and 2-(4-carbomylphenyl) benzoxazole are under development as CEPT inhibitors [77,78].

### 5.3. Cholesterol absorption inhibitors

Ezetamibe is the only drug currently available from this class whose mechanism of action involves inhibition of dietary cholesterol absorption without affecting the absorption of fat-soluble vitamins, triglycerides, and bile acids [59,62,79]. Ezetamibe binds to cholesterol transporter NPL1L1

(Niemann-pick C1-like1) protein in the brush border of intestine as well as in hepatocytes [59,80]. Decrease in cholesterol absorption leads to compensatory up-regulation of LDL receptors on the cell surface and increased LDL cholesterol uptake into cells and decreases blood LDL cholesterol content [59,62]. Ezetamibe also exerts anti-inflammatory effect and also appears to improve renal function [81]. Some side effects of ezetamibe are diarrhea, abdominal pain, arthralgia, backache, myalgia, headache, sinusitis, hepatitis, aphylaxis, myopathy, and rhabdomyolysis [62]. This drug is contraindicated in active liver diseases [62]. Ezetamibe is primarily metabolized in the small intestine and liver via glucuronide conjugation with subsequent biliary and renal excretion [82]. After oral administration, ezetamibe is absorbed and extensively conjugated to a pharmacologically active phenolic glucuronide (ezetamibe-glucuronide), the drug and its metabolite have a half-life of approximately 22 hours [83].



#### 5.4. Cholesterol O-acyltransferase inhibitors

Acyl-CoA: cholesterol O-acyltransferase (ACAT) is an important enzyme involved in re-esterification of absorbed cholesterol within enterocytes [84]. It is involved in the cholesterol metabolism in macrophages, liver, intestine and adrenal cortex and is believed to be involved in secretion of VLDL from liver and development of atherosclerotic lesion [85]. Two ACAT enzymes have been identified, ACAT1 and ACAT2. ACAT1 is found in the ER throughout the body, while ACAT2 is found in the ER of liver and intestinal tissues and may be responsible for the formation of cholesteryl esters [85]. In theory, inhibition of ACAT1 could prevent the transformation of macrophages into foam cells in the vessel wall and, thereby, slow the progression of atherosclerosis and prevent the development of vulnerable plaque and inhibition of ACAT-2 could decrease serum lipid levels by reducing the synthesis of lipoproteins [70]. HL-004 has been found preclinically to be an effective ACAT inhibitor [86]. Presently, specific ACAT 2 inhibitor, such as derivatives of fungal pyripyroneA, are under scrutiny [87].

#### 5.5. Diacylglycerolacyltransferases inhibitors

Diacylglycerolacyltransferases (DGATs) are enzymes involved in adipocyte lipid accumulation and catalyzes the final step reaction of triacylglycerol formation from diacylglycerol [88]. DGAT1 belongs to the same family of proteins as the ACATs [89]. In mammals, DGAT1 is expressed in skeletal muscle, skin, intestine (ileum, colon), and testis, with lower levels of expression in liver and adipose tissue, while DGAT2 is ubiquitous with high expression levels in hepatocytes and adipocytes [90].

#### 5.6. Microsomal TG transfer protein inhibitors

Microsomal TG transfer protein (MTTP) is a heterodimeric lipid transfer protein that catalyzes the transport of TG, cholesteryl ester, and phosphatidylcholine between membranes [91]. MTTP is a protein located in intestine and liver tissues where it plays a role in lipid assembly, transport, and secretion of lipoproteins, triglyceride rich chylomicrons (in enterocytes), and VLDL (in hepatocytes) [92,93]. *In vitro* studies show that MTTP catalyzes the transport of molecules between phospholipid membranes and is also involved in the synthesis of nascent lipoprotein particles within the lumen and ER [94]. The inhibition of MTTP by small molecules should lead to the reduction in plasma TGs and cholesterol levels [95].

Some clinical candidates, such as CP-346086 and BMS-201038, have been shown to inhibit MTTP in both the enterocytes and in the liver [96]. Dirlotapide is an enterocyte-specific MTTP inhibitor that has recently been approved by the FDA as an anti-obesity agent [97]. The most significant side effects involve elevation of hepatic transaminases, nausea, diarrhea, gassiness, and gastrointestinal cramping [98]. Of several MTTP inhibitors, only BMS-201038, now renamed AEGR-733, is still in development [98]. Microsomal triglyceride transfer protein inhibition with lomitapide may offer a treatment option for patients who cannot tolerate statin therapy or who experience insufficient LDL-c reduction with available therapies [95].

#### 5.7. Squalene synthase inhibitors

Squalene synthase, a key enzyme in the cholesterol biosynthetic pathway, occupies the first and solely committed step towards the biosynthesis of the sterol nucleus of cholesterol; hence it is an attractive target for inhibition and the development of novel and improved antihypercholesterolemic agents [99]. Squalene synthase catalyzes one of the subsequent reactions in the cholesterol biosynthetic pathway (i.e. it reductively dimerizes two farnesyl pyrophosphate molecules to form squalene) which is the first intermediate committed to cholesterol [100]. Squalene synthase inhibitors are emerging new stars in the hypolipidemic drug sky and represent a novel class of antihyperlipidemics [59]. Squalene synthase is implicated in the late step in cholesterol biosynthesis and, the squalene synthase inhibitors exerts same effect as that of 3-hydroxy-3-methylglutaryl-coenzyme A-CoA reductase inhibitors, with decreased cholesterol production and up-regulation of LDL receptors [79]. Early inhibitors such as the zaragozic acids showed significant toxicity (acidosis), but a recent compound, lapaquistat, reached Phase III clinical trials [101,102]. EP2306 and EP2302 have been shown to possess antioxidant properties both *in vitro* and *in vivo* [103] as well as to inhibit squalene synthase activity and lipid biosynthesis *in vitro* [104].

#### 5.8. Thyroid hormone analogues

Thyroid hormone has been known to lower total serum cholesterol for many years in hyperthyroidism and during thyroid hormone replacement therapy for hypothyroidism [105]. This action is the result of an accelerated LDL-c clearance rate [106]. T3 increases levels of both the hepatic LDL receptor and its mRNA [107,108]. Additional thyroid hormone actions on lipid metabolism include increasing the activity of lipoprotein lipase [106,109]. More recent understanding of thyroid hormone receptors has led to the development of thyroid hormone mimetics that have selective functions and are potential therapeutic agents to lower cholesterol [110]. Several thyroid hormone analogues have been developed, but the only one with published human data is eprotirome [98], a thyroid hormone analogue containing two bromines that only interacts with the  $\beta$ -receptors found primarily in the liver. It does not seem to have adverse effects on heart and bone [108,111].

#### 5.9. Lanosterol synthase inhibitors

Oxido-squalene-cyclase (lanosterol synthase, LSS) is the second enzyme below the farnesyl pyrophosphate branch point that has been identified as a target for novel antihypercholesterolemic drugs that could complement statins [112]. LSS is located in the ER and converts 2,3-oxidosqualene to lanosterol, the initial four-ringed sterol intermediate in the cholesterol synthesis pathway. The 24(S),25-epoxycholesterol is a ligand of liver X receptor [113]. It also sets the template for the design of inhibitors with improved pharmacological properties for cholesterol lowering and treatment of atherosclerosis. Through the dual mechanism of LSS action (formation of lanosterol; formation of ligands for liver X

receptor), LSS inhibitors have a potential to decrease plasma levels LDL-c and to prevent cholesterol deposition within macrophages [59].

### 5.10. Cholesterol metabolizing cytochrome P450: implication for cholesterol lowering

From the family of P450s, the 7A1, 27A1 and 46A1 are the most important enzymes involved in the control of cholesterol levels in the periphery and brain [114]. CYP7A1 is an important determinant of plasma cholesterol levels and is considered as target for cholesterol lowering [115].

CYP27A1 converts cholesterol to 27-hydroxycholesterol by oxygenation reaction and this is suggested to be important reaction for cholesterol elimination from human lung macrophages and cells in arterial endothelium [116].

### 5.11. AMP-activated protein kinase activator

AMP-activated protein kinase (AMPK), a heterotrimeric energy sensing protein, which restores cellular energy balance by promoting ATP-generating pathways (e.g. FA oxidation) and inhibiting ATP-utilizing pathways (e.g. FA synthesis) [117]. AMPK system plays a major role in regulating glucose and lipid metabolism by effect on energy metabolism and long-term effect on gene expression in the liver [118]. In liver, activation of AMPK results in decreased production of plasma TG and cholesterol and enhanced FA oxidation [119,120]. WS070117 is synthetic lipid lowering agent that is approved preclinically as an effective activator of AMPK with potential capability of inhibition of *de novo* hepatic lipogenesis [121].

### 5.12. Omega-3 FAs

Omega-3 belongs to the polyunsaturated FA family (n-3 PUFA), which includes the 20-carbon eicosapentanoic acid and 22-carbon docosahexaenoic acid, which lowers the TG levels and atherogenic remnant lipoproteins [81]. These FAs are derived from marine sources, especially salmon, mackerel, sardines, and tuna [28]. Omega-3 FAs at 4 g/day usually have favorable effect in lowering TG concentration particularly in the postprandial state and their addition to statins significantly decreases TG, VLDL, and non-HDL-c levels compared with simvastatin alone [122]. Omega-3 FA inhibits expression of SREBP-1, which is involved in the synthesis of FAs [80]. Another broad variety of biological actions shown by omega-3 FAs are hypotriglyceridemia, antiaggregatory, anti-inflammatory, and antiarrhythmic responses [4]. The most common adverse events shown by omega 3 fatty acids in clinical trials are eructation, infection, dyspepsia, and flu syndrome [8].

## 6. Conclusion

Hyperlipidemia is a metabolic disorder characterized by HC and HTG. FHC is one of the types of hyperlipidemia with a genetic basis. In the present review we mainly focused on the new therapeutic drug targets in the treatment of hyperlipidemia. PPARs are regulators of numerous metabolic pathways;

hence there is huge increase in the development and use of agonists of these receptors as therapeutics of dyslipidemia. Inhibition of CETP should lead to elevation of plasma HDL cholesterol and lowering of plasma LDL cholesterol. Ezetimibe is the only drug available today that acts by inhibition of dietary cholesterol absorption without affecting the absorption of fat-soluble vitamins, TGs and bile acids. The inhibitors of ACAT, DGAT, and MTTP, along with thyroid hormone analogue, cholesterol-metabolizing cytochrome P450, AMPK activators, and omega-3 FAs, will be the new therapeutic drug targets in treatment of hyperlipidemia. The inhibitors of certain enzymes such as squalene synthase and lanosterol synthase contribute to the reduction of hyperlipidemia.

## REFERENCES

- [1] Kolovou GD, Anagnostopoulou KK, Cokkinos DV. Pathophysiology of dyslipidemia in the metabolic syndrome. *Postgrad Med J* 2005;81:358–66.
- [2] Bernard J. Free fatty acid receptor family: novel targets for the treatment of diabetes and dyslipidemia. *Curr Opin Investig Drugs* 2008;9:1078–83.
- [3] Funatsu T, Suzuki K, Goto M, Arai Y, Kakuta H, Tanaka H, et al. Prolonged inhibition of cholesterol synthesis by atorvastatin inhibits apo B-100 and triglyceride secretion from HepG2 cells. *Atherosclerosis* 2001;157:107–15.
- [4] Micallef MA, Garg ML. Beyond blood lipids: phytosterols, statins and omega-3 polyunsaturated fatty acid therapy for hyperlipidemia. *J Nutr Biochem* 2009;20:927–39.
- [5] Gordon T, Kannel WB. Premature mortality from coronary heart disease. The Framingham study. *JAMA* 1971;215:1617–25.
- [6] Holvoet P, Jenny NS, Schreiner PJ, Tracy RP, Jacobs DR. The relationship between oxidized LDL and other cardiovascular risk factors and subclinical CVD in different ethnic groups: the Multi-Ethnic Study of Atherosclerosis (MESA). *Atherosclerosis* 2007;194:245–52.
- [7] Foley KA, Vasey J, Alexander CM, Markson LE. Development and validation of the Hyperlipidemia Attitudes and Beliefs In Treatment (HABIT) survey for physicians. *J Gen Intern Med* 2003;18:984–90.
- [8] NHFA. Lipid management guideline. National Heart Foundation of Australia, The Cardiac Society of Australia and New Zealand. *Med J Aust* 2001;175(Suppl):S57–85.
- [9] Fonseca V. The metabolic syndrome, hyperlipidemia and insulin resistance. *Clin Cornerstone* 2005;7:61–72.
- [10] Ginsberg HN, Huang LS. The insulin resistance syndrome: impact on lipoprotein metabolism and atherothrombosis. *J Cardiovasc Risk* 2000;7:325–31.
- [11] Genest JG. Dyslipidemia and coronary artery disease. *Can J Cardiol* 2000;16(Suppl. A): 3A–4A.
- [12] Ninomiya JK, L'Italien G, Criqui MH, Whyte JL, Gamst A, Chen RS. Association of the metabolic syndrome with history of myocardial infarction and stroke in the Third National Health and Nutrition Examination Survey. *Circulation* 2004;109:42–6.
- [13] Anderson JL, Horne BD, Jones HU, Reyna SP, Carlquist JF, Bair TL, et al. For the intermountain heart collaborative (IHC) study. Which features of the metabolic syndrome predict the prevalence and clinical outcomes of angiographic coronary artery disease? *Cardiology* 2004;101:185–93.
- [14] Halpern A, Mancini M, Magalhães ME, Fisberg M, Radominski R, Bertolami MC, et al. Metabolic syndrome,

- dyslipidemia, hypertension and type 2 diabetes mellitus in youth, from diagnosis to treatment. *Diabetol Metab Syndr* 2010;55:1–20.
- [15] Steinberger J, Daniels SR, Eckel RH, Hayman L, Lustig RH, McCrindle B. Progress and challenges in metabolic syndrome in children and adolescents: a scientific statement from the American Heart Association Atherosclerosis, Hypertension, and Obesity in the Young Committee of the Council on Cardiovascular Disease in the Young; Council on Cardiovascular Nursing; and Council on Nutrition, Physical Activity, and Metabolism. *Circulation* 2009;119:628–47.
  - [16] Executive summary of the third report of National Cholesterol Education Program (NCEP) expert panel on detection, evaluation, and treatment of high blood cholesterol in adults (Adult Treatment Panel III). *JAMA* 2001;285:2486–97.
  - [17] McCrindle BW, Ose L, Marais AD. Efficacy and safety of atorvastatin in children and adolescents with familial hypercholesterolemia or severe hyperlipidemia: a multicenter, randomized, placebo-controlled trial. *J Pediatr* 2003;143:74–80.
  - [18] Iqbal J, Hussain MM. Intestinal lipid absorption. *Am J Physiol Endocrinol Metab* 2009;296:E1183–94.
  - [19] Lien EL. The role of fatty acid composition and positional distribution in fat absorption in infants. *J Pediatr* 1994;125: S62–8.
  - [20] Carlier H, Bernard A, Caselli C. Digestion and absorption of polyunsaturated fatty acids. *Reprod Nutr Dev* 1991;31: 475–500.
  - [21] Ramírez M, Amate L, Gil A. Absorption and distribution of dietary fatty acids from different sources. *Early Human Development* 2001;65(Suppl.):S95–101.
  - [22] Hernell O, Bläckberg L. Digestion of human milk lipids: physiological significance of sn-2 monoacylglycerol hydrolysis by salt-stimulated lipase. *Pediatr Res* 1982;16: 882–5.
  - [23] Jain KS, Kathiravan MK, Somania RS, Shishoo CJ. The biology and chemistry of hyperlipidemia. *Bioorg Med Chem* 2007;15:4674–99.
  - [24] Capell WH, Spiegelman KP, Eckel RH. Therapeutic targets in severe hypertriglyceridemia. *Drug Disc Today: Dis Mech* 2004;1:171–7.
  - [25] Athyros VG, Giouleme OI, Nikolaidis NL, Vasiliadis TV, Bouloukos VI, Kontopoulos AG, et al. Long-term follow-up of patients with acute hypertriglyceridemia-induced pancreatitis. *J Clin Gastroenterol* 2002;34:472–5.
  - [26] Pejic RN, Lee DT. Hypertriglyceridemia. *J Am Board Fam Med* 2006;19:310–6.
  - [27] Hopkins PN, Heiss G, Ellison RC, Province MA, Pankow JS, Eckfeldt JH, et al. Coronary artery disease risk in familial combined hyperlipidemia and familial hypertriglyceridemia: a case-control comparison from the National Heart, Lung, and Blood Institute Family Heart Study. *Circulation* 2003;108:519–23.
  - [28] Bersot T, Haffner S, Harris WS, Kellick KA, Morris CM. Hypertriglyceridemia: management of atherogenic dyslipidemia. *J Fam Pract* 2006;55:S1–8.
  - [29] Feoli-Fonseca JC, Lévy E, Godard M, Lambert M. Familial lipoprotein lipase deficiency in infancy: clinical, biochemical, and molecular study. *J Pediatr* 1998;133:417–23.
  - [30] Francis A, Levy Y. Chylomicronemia syndrome. *Harefuah* 2002;14:201–3.
  - [31] Dunbar RL, Rader DJ. Demystifying triglycerides: a practical approach for the clinician. *Cleve Clin J Med* 2005;72:661–80.
  - [32] Bhatnagar D, Soran H, Durrington PN. Hypercholesterolemia and its management. *BMJ* 2008;337: 503–8.
  - [33] Jia L, Fu M, Tian Y, Xu Y, Gou L, Tian H, et al. Alterations of high-density lipoprotein subclasses in hypercholesterolemia and combined hyperlipidemia. *Int J Cardiol* 2007;120:331–7.
  - [34] Department of Health. National service framework for coronary heart disease. London: DoH; 2000.
  - [35] Orsó E, Ahrens N, Kilalić D, Schmitz G. Familial hypercholesterolemia and lipoprotein(a) hyperlipidemia as independent and combined cardiovascular risk factors. *Atherosclerosis Suppl* 2009;10:74–8.
  - [36] Bhatnagar D. Diagnosis and screening for familial hypercholesterolemia: finding the patient, finding the genes. *Ann Clin Biochem* 2006;43:441–56.
  - [37] Lambert G, Charlton F, Rye KA, Piper DE. Molecular basis of PCSK9 function. *Atherosclerosis* 2009;203:1–7.
  - [38] Hopkins PN. Familial hypercholesterolemia—improving treatment and meeting guidelines. *Int J Cardiol* 2003;89:13–23.
  - [39] de Graaf J, van der Vleuten GM, ter Avest E, Dallinga-Thie GM, Stalenhoef AF. High plasma level of remnant-like particles cholesterol in familial combined hyperlipidemia. *J Clin Endocrinol Metab* 2007;92:1269–75.
  - [40] Brunzell JD, Schrott HG, Motulsky AG, Bierman EL. Myocardial infarction in the familial forms of hypertriglyceridemia. *Metabolism* 1976;25:313–20.
  - [41] de Graaf J, Veerkamp MJ, Stalenhoef AF. Metabolic pathogenesis of familial combined hyperlipidaemia with emphasis on insulin resistance, adipose tissue metabolism and free fatty acids. *J R Soc Med* 2002;95:46–53.
  - [42] Kieffer TJ, Habener JF. The adipoinular axis: effects of leptin on pancreatic beta-cells. *Am J Physiol Endocrinol Metab* 2000;278:E1–14.
  - [43] Considine RV, Sinha MK, Heiman ML, Kriauciunas A, Stephens TW, Nyce MR, et al. Serum immunoreactive leptin concentrations in normal-weight and obese humans. *N Engl J Med* 1996;334:292–5.
  - [44] Melloul D, Marshak S, Cerasi E. Regulation of insulin gene transcription. *Diabetologia* 2002;45:309–26.
  - [45] van der Vleuten GM, Veerkamp MJ, van Tits LJ, Toenhake H, den Heijer M, Stalenhoef AF, et al. Elevated leptin levels in subjects with familial combined hyperlipidemia are associated with the increased risk for CVD. *Atherosclerosis* 2005;183:355–60.
  - [46] Grundy SM. Low-density lipoprotein, non-high-density lipoprotein, and apolipoprotein B as targets of lipid-lowering therapy. *Circulation* 2002;106:2526–9.
  - [47] Frost PH, Havel RJ. Rationale for use of non-high-density lipoprotein cholesterol rather than low-density lipoprotein cholesterol as a tool for lipoprotein cholesterol screening and assessment of risk and therapy. *Am J Cardiol* 1998;81: 26B–31B.
  - [48] Karpe F, Hamsten A. Postprandial lipoprotein metabolism and atherosclerosis. *Curr Opin Lipidol* 1995;6:123–9.
  - [49] Karpe F, Taskinen MR, Nieminen MS, Frick MH, Kesäniemi YA, Pasternack A, et al. Remnant-like lipoprotein particle cholesterol concentration and progression of coronary and vein-graft atherosclerosis in response to gemfibrozil treatment. *Atherosclerosis* 2001;157:181–7.
  - [50] ter Avest E, Holeywijn S, Bredie SJ, Stalenhoef AF, de Graaf J. Remnant particles are the major determinant of an increased intima media thickness in patients with familial combined hyperlipidemia (FCH). *Atherosclerosis* 2007;191: 220–6.
  - [51] Austin MA, Brunzell JD, Fitch WL, Krauss RM. Inheritance of low density lipoprotein subclass patterns in familial combined hyperlipidemia. *Arteriosclerosis* 1990;10:520–30.
  - [52] Cortner JA, Coates PM, Bennet MJ, Cryer DR, Le NA. Familial combined hyperlipidaemia: use of stable isotopes to demonstrate overproduction of very low-density



- lipoprotein apolipoprotein B by the liver. *J Inherit Metab Dis* 1991;14:915–22.
- [53] Venkatesan S, Cullen P, Pacy PJ, Halliday D, Scott J. Stable isotopes show a direct relation between VLDL apoB overproduction and serum triglyceride levels and indicate a metabolically and biochemically coherent basis for familial combined hyperlipidemia. *Arterioscler Thromb* 1993;13:1110–8.
- [54] Austin MA, Krauss RM. Genetic control of low-density-lipoprotein subclasses. *Lancet* 1986;2:592–5.
- [55] Calabresi L, Donati D, Pazzucconi F, Sirtori CR, Franceschini G. Omacor in familial combined hyperlipidemia: effects on lipids and low density lipoprotein subclasses. *Atherosclerosis* 2000;148:387–96.
- [56] van der Vleuten GM, Kluijtmans LA, Hijmans A, Blom HJ, Stalenhoef AFH, de Graaf J. The Gln223Arg polymorphism in the leptin receptor is associated with familial combined hyperlipidemia. *Int J Obes* 2006;30:892–8.
- [57] van der Vleuten GM, Isaacs A, Hijmans A, van Duijn CM, Stalenhoef AFH, de Graaf J. The involvement of upstream stimulatory factor 1 in Dutch patients with familial combined hyperlipidemia. *J Lipid Res* 2007;48:193–200.
- [58] Ranjan N. Management of hyperlipidemias: an update. *Indian J Dermatol Venereol Leprol* 2009;75:452–62.
- [59] Rozman D, Monostory K. Perspectives of the non-statin hypolipidemic agents. *Pharmacol Ther* 2010;127:19–40.
- [60] Kontush A, Chapman MJ. Functionally defective high density lipoprotein: A new therapeutic target at the crossroads of dyslipidemia, inflammation, and atherosclerosis. *Pharmacol Rev* 2006;58:342–74.
- [61] Jacobson TA, Millar M, Schaefer EJ. Hypertriglyceridemia and cardiovascular risk reduction. *Clin Therap* 2007;29:763–77.
- [62] Lin Y, Mousa SS, Elshourbagy N, Mousa SA. Current status and future directions in lipid management: emphasizing low density lipoproteins, high density lipoproteins, and triglycerides as targets for therapy. *Vasc Health Risk Manag* 2010;6:73–85.
- [63] Willson TM, Brown PJ, Sternbach DD, Henke BR. The PPARs: from orphan receptors to drug discovery. *J Med Chem* 2000;43:527–50.
- [64] Pakala R, Kuchulakanti K, Rha SW, Cheneau E, Baffour R, Waksman R. Peroxisome proliferator-activated receptor gamma: its role in metabolic syndrome. *Cardiovasc Radiat Med* 2004;5:97–103.
- [65] Gross BS, Fruchart JC, Staels B. Peroxisome Proliferator-Activated Receptor  $\beta/\delta$ : A novel target for the reduction of atherosclerosis. *Drug Disc Today: Therap Strat* 2005;2:237–43.
- [66] Gervois P, Torra IP, Fruchart JC, Staels B. Regulation of lipid and lipoprotein metabolism by PPAR activators. *Clin Chem Lab Med* 2000;38:3–11.
- [67] Chinetti G, Lestavel S, Bocher V, Remaley AT, Neve B, Torra IP, et al. PPAR-alpha and PPAR-gamma activators induce cholesterol removal from human macrophage foam cells through stimulation of the ABCA1 pathway. *Nat Med* 2001;7:53–8.
- [68] Kasuga J, Yamasaki D, Araya Y, Nakagawa A, Makishima M, Doi T, et al. Design, synthesis, and evaluation of a novel series of  $\alpha$ -substituted phenylpropanoic acid derivatives as human peroxisome proliferator-activated receptor (PPAR)  $\alpha/\delta$  dual agonists for the treatment of metabolic syndrome. *Bioorg Med Chem* 2006;14:8405–14.
- [69] Jeong HW, Lee JW, Kim WS, Choe SS, Shin HJ, Lee GY, et al. A nonthiazolidinedione peroxisome proliferator-activated receptor  $\alpha/\gamma$  dual agonist CG301360 alleviates insulin resistance and lipid dysregulation in db/db mice. *Mol Pharmacol* 2010;78:877–85.
- [70] Tall AR. Plasma cholesteryl ester transfer protein. *J Lipid Res* 1993;34:1255–74.
- [71] Rano TA, Sieber-McMaster E, Pelton PD, Yang M, Demarest KT, Kuo G. Design and synthesis of potent inhibitors of cholesteryl ester transfer protein (CETP) exploiting a 1,2,3,4-tetrahydroquinoline platform. *Bioorg Med Chem Lett* 2009;19:2456–60.
- [72] Champman JM, Guerin M. CETP, a key player in atherogenic dyslipidemia of Type ii diabetes. *Int Congr Ser* 2004;1262:503–6.
- [73] Gurfinkel R, Joy TR. Anacetrapib: hope for CETP inhibitors? *Cardiovasc Therap* 2011;29:327–39.
- [74] Davidson MH. Update on CETP inhibition. *J Clin Lipidol* 2010;4:394–8.
- [75] Smith CJ, Ali A, Chen L, Hammond ML, Anderson MS, Chen Y, et al. 2-Arylbenzoxazoles as CETP inhibitors: substitution of the benzoxazole moiety. *Bioorg Med Chem Lett* 2010;20:346–9.
- [76] Schmeck C, Gielen-Haertwig H, Vakalopoulos A, Bischoff H, Li V, Wirtz G, et al. Novel tetrahydroquinoline derived CETP inhibitors. *Bioorg Med Chem Lett* 2010;20:1740–3.
- [77] Vakalopoulos A, Schmeck C, Thutewohl M, Li V, Bischoff H, Lustig K, et al. Chromanol derivatives-A novel class of CETP inhibitors. *Bioorg Med Chem Lett* 2011;21:488–91.
- [78] Sweis RF, Hunt JA, Kallashi F, Hammond ML, Chen Y, Eveland SS, et al. 2-(4-Carbonylphenyl) benzoxazole inhibitors of CETP: scaffold design and advancement in HDLc-raising efficacy. *Bioorg Med Chem Lett* 2011;21:1890–5.
- [79] Nutescu EA, Shapiro NL. Ezetimibe: a selective cholesterol absorption inhibitor. *Pharmacotherapy* 2003;23:1463–74.
- [80] Duntas L, Kolovou G. Options for the treatment of hyperlipidemia in type 2 diabetes mellitus and hypothyroidism: lowering the cardiovascular risk. *Future Cardiol* 2011;7:137–44.
- [81] Athyros VG, Tziomalos K, Karagiannis A, Mikhailidis DP. Dyslipidaemia of obesity, metabolic syndrome and type 2 diabetes mellitus: the case for residual risk reduction after statin treatment. *Open Cardiovasc Med J* 2011;5:24–34.
- [82] Jeu L, Cheng JW. Pharmacology and therapeutics of ezetimibe (SCH 58235), a cholesterol-absorption inhibitor. *Clin Ther* 2003;25:2352–87.
- [83] Al-Shaer MH, Choueiri NE, Suleiman ES. The pivotal role of cholesterol absorption inhibitors in the management of dyslipidemia. *Lipids Health Dis* 2004;3:22.
- [84] Norum KA, Lolljeqvist A, Helgerud P, Normann ER, Selbekk AM, Selbekk B. Esterification of cholesterol in humans small intestine: the importance of acyl-CoA:cholesterolacyltransferase. *Eur J Clin Invest* 1979;9:55–62.
- [85] Insull W, Koren M, Davignon J, Sprecher D, Schrott H, Keilson LM, et al. Efficacy and short-term safety of a new ACAT inhibitor, avasimibe, on lipids, lipoproteins, and apolipoproteins, in patients with combined hyperlipidemia. *Atherosclerosis* 2001;157:137–44.
- [86] Asami Y, Kondo Y, Murakami S, Araki H, Tsuchida K, Higuchi S. The ACAT inhibitor HL-004 inhibits cholesterol absorption and lowers serum cholesterol in rats. *Gen Pharmac* 1998;31:593–6.
- [87] Costet P. Molecular pathways and agents for lowering LDL-cholesterol in addition to statins. *Pharmacol Ther* 2010;126:263–78.
- [88] Meuwese MC, de Groot E, Duivenvoorden R, Trip MD, Ose L, Maritz FJ, et al. ACAT Inhibition and progression of carotid atherosclerosis in patients with familial hypercholesterolemia. *JAMA* 2009;301:1131–9.
- [89] Lardizabal KD, Mai JT, Wagner NW, Wyrick A, Voelker T, Hawkins DJ. DGAT2 is a new diacylglycerol acyltransferase

- gene family: purification, cloning, and expression in insect cells of two polypeptides from *Mortierella ramanniana* with diacylglycerol acyltransferase activity. *J Biol Chem* 2001;276:38862–9.
- [90] Zammit VA, Buckett LK, Turnbull AV, Wure H, Proven A. Diacylglycerol acyltransferases: potential roles as pharmacological targets. *Pharmacol Ther* 2008;118:295–302.
- [91] Jamil H, Gordon DA, Eustice DC, Brooks CM, Dickson JK, Chen Y, et al. An inhibitor of the microsomal triglyceride transfer protein inhibits apoB secretion from HepG2 cells. *Proc Natl Acad Sci U S A* 1996;93:11991–5.
- [92] Sulsky R, Robl JA, Biller SA, Harrity TW, Wetterau J, Connolly F, et al. 5-Carboxamido-1,3,2-dioxaphosphorinanes, potent inhibitors of MTP. *Bioorg Med Chem Lett* 2004;14:5067–70.
- [93] Li J, Bertinato P, Cheng H, Cole BM, Bronk BS, Jaynes BH, et al. Discovery of potent and orally active MTP inhibitors as potential anti-obesity agents. *Bioorg Med Chem Lett* 2006;16:3039–42.
- [94] Matsuda D, Tomoda H. DGAT inhibitors for obesity. *Curr Opin Investig Drugs* 2007;10:836–41.
- [95] Rizzo M. Lomitapide, a microsomal triglyceride transfer protein inhibitor for the treatment of hypercholesterolemia. *IDrugs* 2010;13:103–11.
- [96] Wetterau JR, Gregg RE, Harrity TW, Arbeeney C, Cap M, Connolly F, et al. An MTP inhibitor that normalizes atherogenic lipoprotein levels in WHHL rabbits. *Science* 1998;282:751–4.
- [97] Vu CB, Milne JC, Carney DP, Song J, Choy W, Lambert PD, et al. Discovery of benzothiazole derivatives as efficacious and enterocyte-specific MTP inhibitors. *Bioorg Med Chem Lett* 2009;19:1416–20.
- [98] Goldberg AC. Novel therapies and new targets of treatment for familial hypercholesterolemia. *J Clin Lipidol* 2010;4:350–6.
- [99] Kourounakis AP, Matralis AN, Nikitakis A. Design of more potent squalene synthase inhibitors with multiple activities. *Bioorg Med Chem* 2010;18:7402–12.
- [100] Hiyoshi H, Yanagimachi M, Ito M, Saeki T, Yoshida I, Okada T, et al. Squalene synthase inhibitors reduce plasma triglyceride through a low-density lipoprotein receptor-independent mechanism. *Eur J Pharmacol* 2001;431:345–52.
- [101] Stein EA. Other therapies for reducing low-density lipoprotein cholesterol: medications in development. *Endocrinol Metab Clin North Am* 2009;38:99–119.
- [102] Davidson MH. Novel nonstatin strategies to lower low-density lipoprotein cholesterol. *Curr Atheroscler Rep* 2009;11:67–70.
- [103] Tavidou A, Manolopoulos VG. Antioxidant properties of two novel 2-biphenylmorpholine compounds (EP2306 and EP2302) *in vitro* and *in vivo*. *Eur J Pharmacol* 2004;505:213–21.
- [104] Tavidou A, Kaklamanis L, Megaritis G, Kourounakis AP, Papalois A, Roukounas D, et al. Pharmacological characterization *in vitro* of EP2306 and EP2302, potent inhibitors of squalene synthase and lipid biosynthesis. *Eur J Pharmacol* 2006;535:34–42.
- [105] Mason RL, Hunt HM, Hurxthal LM. Blood cholesterol values in hyperthyroidism and hypothyroidism: their significance. *N Engl J Med* 1930;203:1273–8.
- [106] Morkin E, Ladenson P, Goldman S, Adamson C. Thyroid hormone analogs for treatment of hypercholesterolemia and heart failure: past, present and future prospects. *J Mol Cell Cardiol* 2004;37:1137–46.
- [107] Staels B, van Tol A, Chan L, Will HM, Verhoeven GA, Auwerx J. Alterations in thyroid status modulate apolipoprotein, hepatic triglyceride lipase, and low-density lipoprotein receptor in rats. *Endocrinology* 1990;127:1144–52.
- [108] Salter AM, Hayashi R, Al-Seeni M, Brown NF, Bruce J, Sorensen O, et al. Effects of hypothyroidism and high-fat feeding on mRNA concentrations for the low-density lipoprotein receptor and on acyl-CoA:cholesterol acyltransferase activities in rat liver. *Biochem J* 1991;276:825–32.
- [109] Packard CJ, Shepard J, Lindsay GM, Gaw A, Taskinen MR. Thyroid replacement therapy and its influence on postheparin plasma lipases and apolipoprotein-b metabolism in hypothyroidism. *J Clin Endocrinol Metab* 1993;76:1209–16.
- [110] Baxter DJ, Webb P. Thyroid hormone mimetics: potential applications in atherosclerosis, obesity and type 2 diabetes. *Nat Rev Drug Disc* 2009;8:308–20.
- [111] Ladenson PW, Kristensen JD, Ridgway EC, Olsson AG, Carlsson B, Klein I, et al. Use of the thyroid hormone analogue eprotirome in statin-treated dyslipidemia. *N Engl J Med* 2010;362:906–16.
- [112] Korosec T, Acimovic J, Seliskar M, Kocjan D, Tacer KF, Rozmanb D, Urleba U. Novel cholesterol biosynthesis inhibitors targeting human lanosterol 14a-demethylase (CYP51). *Bioorg Med Chem* 2008;16:209–21.
- [113] Rowe AH, Argmann CA, Edwards JY, Sawyez CG, Morand OH, Hegele RA, et al. Enhanced synthesis of the oxysterol 24(S), 25-epoxycholesterol in macrophages by inhibitors of 2, 3-oxidosqualene:lanosterol cyclase: A novel mechanism for the attenuation of foam cell formation. *Circ Res* 2003;93:717–25.
- [114] Pikuleva IA. Cholesterol-metabolizing cytochromes P450: implication for cholesterol lowering. *Expert Opin Drug Metab Toxicol* 2008;4:1403–14.
- [115] Bjorkhem I, Reihner E, Angelin B, Ewerth S, Akerlund JE, Einarsson K. On the possible use of the serum level of 7 alpha-hydroxycholesterol as a marker for increased activity of the cholesterol 7 alpha-hydroxylase in humans. *J Lipid Res* 1987;28:889–94.
- [116] Babiker A, Anderson O, Lund E, Xiu RJ, Deeb S, Reshef A, et al. Elimination of cholesterol in macrophages and endothelial cells by the sterol 27-hydroxylase mechanism. Comparison with high density lipoprotein-mediated reverse cholesterol transport. *J Biol Chem* 1997;272:26253–61.
- [117] Hardie DG. Role of AMP-activated protein kinase in the metabolic syndrome and in heart disease. *FEBS Lett* 2008;582:81–9.
- [118] Viollet B, Guigas B, Leclerc J, Hébrard S, Lantier L, Mounier R, et al. AMP-activated protein kinase in the regulation of hepatic energy metabolism: from physiology to therapeutic perspectives. *Acta Physiol* 2009;196:81–98.
- [119] Henin N, Vincent MF, Gruber HE, van den Berghe G. Inhibition of fatty acid and cholesterol synthesis by stimulation of AMP-activated protein kinase. *FASEB J* 1995;9:541–6.
- [120] Muoio DM, Seefeld K, Witters LA, Coleman RA. AMP-activated kinase reciprocally regulates triacylglycerol synthesis and fatty acid oxidation in liver and muscle: evidence that sn-glycerol-3-phosphate acyltransferase is a novel target. *Biochem J* 1999;338:783–91.
- [121] Lian Z, Li Y, Gao J, Qu K, Li J, Hao L, et al. A novel AMPK activator, WS070117, improves lipid metabolism disorders in hamsters and HepG2 cells. *Lipids Health Dis* 2011;10:1–8.
- [122] Durrington PN, Bhatnager D, Mackness MI, Morgan J, Julier K, Khan MA, et al. An omega-3-polyunsaturated fatty acid concentrate administered for one year decreased triglycerides in simvastatin treated patients with coronary heart disease and persisting hypertriglyceridemia. *Heart* 2001;85:544–8.



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## Review article

# Negatively charged L5 as a naturally occurring atherogenic low-density lipoprotein

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## ABSTRACT

Deranged metabolism of low-density lipoprotein (LDL) is considered the preeminent modifiable risk factor for atherosclerotic disease, and it is widely viewed as a chronic inflammatory disorder. Yet, the search for a circulating atherogenic LDL species continues, as the risk of coronary artery disease (CAD) cannot be measured by absolute LDL cholesterol concentrations in the plasma. Oxidized LDL (oxLDL) and small, dense LDL are associated with CAD, but neither has been retrieved from human plasma for mechanistic scrutiny. Electronegative LDL, a subclass of human plasma LDL, exhibits atherogenic properties in cultured vascular cells. L5, the most negatively charged subfraction of LDL, is an extreme form of electronegative LDL that we isolated through anion-exchange chromatography from the plasma of patients with increased cardiac risk (active smoking, hypercholesterolemia, type 2 diabetes mellitus, and metabolic syndrome). L5, which is scant in healthy normal patients, is as potent as artificially prepared oxLDL in inducing endothelial cell (EC) apoptosis by disrupting fibroblast growth factor 2 autoregulation that involves protein kinase B. Unlike oxLDL, however, L5 is not oxidized. Among subfractions L1–L5, which were separated by our chromatographic method, L1 is the most abundant and least negatively charged. It represents harmless normal LDL. Compared with L1, L5 has a greater content of total protein and triglycerides but a lesser amount of cholesteryl esters. Size exclusion chromatography and equilibrium density gradient ultracentrifugation indicated that L5 is neither smaller nor denser than L1. Negative charge on the particle surface has made L5 unrecognizable by the normal LDL receptor. Instead, L5 signals through and is internalized by lectin-like oxidized LDL receptor-1 (LOX-1), which has high affinity for negatively charged ligands. LOX-1 is also inducible by L5 but not L1. Through LOX-1, L5 disturbs homeostasis between the prosurvival and proapoptotic members of the Bcl-2 family, leading to mitochondrial destabilization. Additionally, it induces overexpression of various adhesion molecules and chemokines, thus promoting monocyte-EC adhesion, an early event during atherosclerosis development. Endothelial progenitor cells (EPCs) are important construction units for vascular repair and endothelial regeneration. Adding to the damage, L5 impairs EPC differentiation from

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mononuclear cells by inhibiting the induction of needed growth factor receptors. It also accelerates EPC senescence by suppressing the enzymatic activity of telomerase, which is essential for chromosome preservation. Thus, L5 is a naturally occurring, negatively charged but not oxidized LDL entity that is neither smaller nor denser than normal LDL but possesses a capacity for inducing a spectrum of atherogenic responses in vascular cells. Further investigation aimed at establishing its clinical relevance is warranted to confirm its atherogenic role. Subsequent efforts in L5 research will be directed toward the development of new diagnostic and treatment methods for CAD and other ischemic vascular diseases.

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## 1. Introduction

There is no further doubt that low-density lipoprotein (LDL) abnormality is a primary etiology of atherosclerosis and associated complications, especially coronary artery disease (CAD). Under this notion and in view of the heterogeneity of LDL, investigators have searched extensively for the LDL species responsible for atherogenesis. Oxidized LDL (oxLDL) and small, dense LDL receive the greatest attention. Although it is supported by accumulating experimental evidence and the localization of oxidized lipids in lesions [1–3], it remains uncertain whether LDL oxidation is a cause of atherosclerosis [4]. One major reason for this doubt is that most experimental data have been derived from oxLDL that has been artificially prepared *in vitro*. Circulating small, dense LDL has been statistically related to atherosclerosis and CAD events [5,6]. Despite a possible correlation with CAD, neither form of LDL has been isolated from human plasma, to allow investigation of their effects in vascular cells. By contrast, interest is growing in another possible candidate: negatively charged LDL, a subclass of circulating LDL that is defined as either unoxidized or minimally oxidized yet potentially atherogenic.

## 2. Electronegative LDL

Hoff, Gotto, and associates [7–9] first used the term “electronegative LDL” when they noted that LDL isolated from human atherosclerotic lesions exhibited greater mobility toward the anode end in agarose gel electrophoresis than LDL isolated from normal plasma. Using fast protein liquid chromatography (FPLC) equipped with an ion-exchange column, Avogaro and colleagues [10] first divided human plasma LDL dichotomously into electropositive LDL(+) and electronegative LDL(–) in 1988. They were also the first to report that LDL(–) particles were “stickier” to one another and more toxic to vascular cells than their LDL(+) counterparts [10]. Since then, several groups have described the chemical composition and functional traits of LDL(–), which they isolated by using a similar protocol [11–26]. Among these investigators, Sánchez-Quesada and associates [19,20] have reported on a wide range of chemical and biologic features of LDL(–). In a 2004 review, they summarized that in cultured vascular endothelial cells (ECs), LDL(–) induces the production of chemokines, such as interleukin 8 (IL-8) and monocyte chemoattractant protein 1, and increases tumor necrosis factor alpha (TNF- $\alpha$ )-induced

production of vascular cell adhesion molecule 1 (VCAM-1). Most recently, this group reported that phospholipase C-like activity may play a role in higher aggregability of LDL(–) [25], the phenomenon originally noted by Avogaro [10].

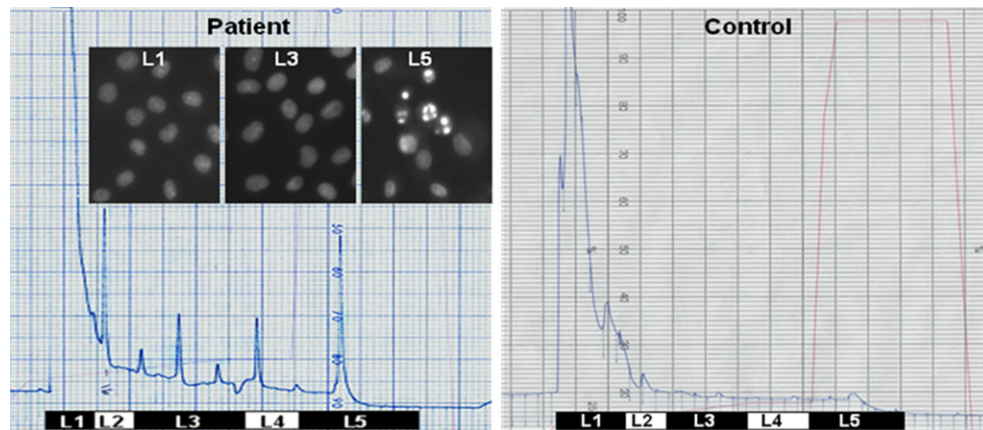
## 3. L5 and negatively charged LDL

Using a different protocol, we used anion-exchange chromatography to divide plasma LDL from patients with familial hypercholesterolemia heterogeneity and those with moderately increased LDL cholesterol (LDL-C) into increasingly negatively charged subfractions, L1–L5 [27,28]. Strictly speaking, in chemistry, the term “electronegativity” is defined as the tendency of an atom to attract a bonding pair of electrons. On the Pauling scale, fluorine (the most electronegative element) is assigned a value of 4.0; values range down to 0.7, in the case of cesium and francium (the least electronegative elements) [29]. L5 is only relatively more negative in surface charge than other subfractions, and the complexity of an LDL particle prevents accurate calculation of its electronegativity; we decided to define L5 as the most negatively charged LDL subfraction. This allows us to avoid inaccurately implying that L4–L1 are positively charged LDL particles. Among all subfractions, L5 is the only subfraction capable of inducing apoptosis in cultured vascular ECs (Fig. 1).

## 4. Chemical basis of L5 negative charge and its biologic implications

We previously reported that L5 accounts for approximately 2% of total plasma LDL in asymptomatic patients with type 2 diabetes mellitus (DM) [30]. L1, the least negatively charged subfraction, represents a majority (> 85%) of total LDL. The content of both protein and triglyceride (TG) increased progressively in the direction of L1 to L5, whereas that of cholesteryl esters decreased. The content of phospholipids and free cholesterol was largely the same among all subfractions [30]. These findings concur with those reported for LDL(–) [12]. Sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE) analysis revealed higher total protein content in L5 particles, which is partially explained by the inclusion of apolipoprotein (apo)-AI, apoE, and apoCIII, as well as apoB-100, which is the sole apolipoprotein of L1 [30]. Additional experiments with L1–L5 isolated from hypercholesterolemic human plasma yielded





**Fig. 1 – Isolation of L5 and other LDL subfractions by FPLC. L5 is present in patients with elevated LDL-C (> 160 mg/dL) but not in nondiabetic, normolipidemic healthy subjects. L5 is the only subfraction that induces marked EC apoptosis, as demonstrated by nuclear condensation and fragmentation. C = cholesterol; EC = endothelial cell; FPLC = fast protein liquid chromatography; LDL = low-density lipoprotein.**

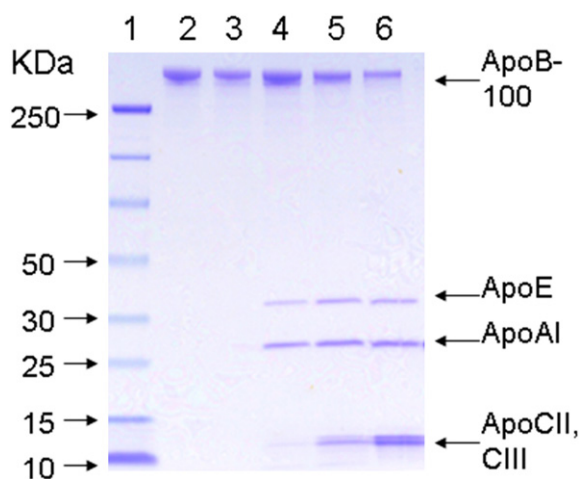
similar results; Fig. 2 shows a representative SDS-PAGE gel. Fig. 3 illustrates similarities and differences between L5 and L1. Because L5 contains twice the amount of TG than L1, it can be regarded as a TG-rich LDL by comparison. Compatible with our findings, Bancells and colleagues [26] demonstrated recently by proteomics analysis that the amounts of apoE, apoAI, apoC-III, apoAII, apoD, apoF, and apoJ are higher in LDL(–) than in LDL(+). In our latest experiments with liquid chromatography/mass spectrometry (LC/MS<sup>E</sup>), we quantitated in detail all apolipoproteins and other low molecular weight proteins associated with L5. Selective association of these molecules with low isoelectric points in L5, but not L1, in part explains the relatively high negative charge on the surface of L5 (unpublished data), which may contribute to the altered affinity of L5 for the normal LDL receptor (LDLR). The chemically oriented

shift of L5's affinity from LDLR to other receptors constitutes the basis of its unique pathologic functionality.

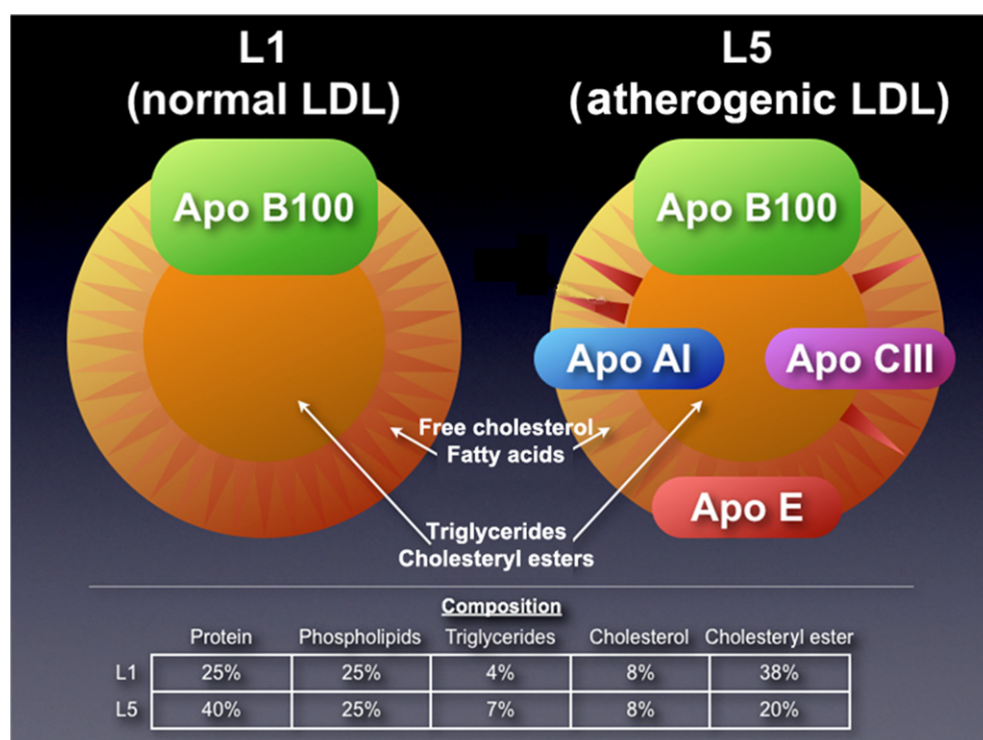
## 5. L5 vs. oxLDL and small, dense LDL

Although L5 and copper-oxidized LDL are equally potent in suppressing the transcription of fibroblast growth factor 2 (FGF2) and inducing apoptosis in vascular ECs [28,31], they differ widely in their chemical and physical characteristics. The production of thiobarbituric acid-reactive substances (TBARS) is often used as a measure of oxidative lipid modification. After copper and oxygen exposure for 24 hours, the TBARS value of oxLDL often reaches the high concentration of 18–22 nmol/mg protein. In contrast, the TBARS value of L5 is mostly less than 1 nmol/mg protein, which is no different than that of L1–L4 [28]; this is in accordance with observations made in LDL(–) [11,20,22]. Various artificial oxidation methods increase the negative charge of normal LDL and turn it into electronegative LDL [17,32]. In preliminary experiments, copper oxidation of L1 yielded ox-L1, which exhibited increased electrophoretic mobility on agarose gel, as well as proapoptotic effects, as seen with non oxidized L5 (data not shown). However, neither oxLDL nor ox-L1 has the same chemical composition as naturally occurring L5. Therefore, L5 is a nonoxidized, naturally occurring atherogenic LDL.

L5 is a subfraction of plasma LDL, which is defined by density. The high TG content and the association with apoCIII, apoE, and apoAI suggest L5's close resemblance to remnant-like particle cholesterol [33,34]. Size exclusion chromatography and equilibrium density gradient ultracentrifugation showed that L5 is no different from L1–L4 in either particle size or density [35]. This suggests that L5 is neither smaller nor denser than other particles in the same lipoprotein class defined by density. Small, dense LDL is defined by particle size on gradient gel electrophoresis or nuclear magnetic resonance (NMR) [36]. Small, dense LDL is considered atherogenic chiefly because it has a greater propensity for oxidation, and it may be concluded that it is an etiologic agent in atherosclerosis. The association of



**Fig. 2 – Representative 4–20% SDS-PAGE gel that shows the apolipoprotein distribution in L1–L5 from hypercholesterolemic human plasma. Protein profiles are shown for L1–L5 (from left to right: Lanes 2–6); apoB-100 is the sole protein in L1, whereas L5 also contains apoE, apoAI, and apoCII/CIII. apo = apolipoprotein; SDS-PAGE = sodium dodecyl sulfate polyacrylamide gel electrophoresis.**



**Fig. 3 – Schematic comparison between L1 and L5 isolated from hypercholesterolemic human plasma. L5 and L1 differ in their lipid and protein compositions. LDL = low-density lipoprotein; apo = apolipoprotein.**

LDL particle size with cardiovascular diseases has been tested for magnitude and independence in many studies, e.g., clinical intervention trials, and cross-sectional and prospective epidemiologic studies. Nearly all show a significant univariate association of small, dense LDL with increased CAD risk, but LDL size is seldom a significant and independent predictor of CAD risk after multivariate adjustment for confounding variables, in particular, plasma TG and high-density lipoprotein cholesterol (HDL-C) concentration. Hence, it may be that the increased risk associated with smaller LDL size in univariate analyses arises from broader pathophysiologic causes, of which small, dense LDL is a part, rather than a reflection, of intrinsic increased atherogenic potential. Thus, a clear causal relationship between small, dense LDL and increased cardiovascular risk has not been proven [37]. This further accentuates the importance of L5, which is not a small, dense LDL, in atherogenesis.

## 6. Receptors for and active components of L5

Our data suggest that at least two receptors, platelet activating factor (PAF) receptor (PAFR) and lectin-like oxidized LDL receptor-1 (LOX-1), are involved in transducing L5-elicited signaling of ECs and endothelial progenitor cells (EPCs) [31,38,39]. Protein kinase B (Akt) exerts multiple prosurvival and vasomotor effects by activating downstream targets, such as endothelial nitric oxide synthase (eNOS), after its own activation by phosphatidylinositol 3-kinase (PI3K) [40,41]. L5 inhibits EC proliferation and induces EC apoptosis, disrupting FGF2 autoregulation via an FGF2-PI3K-Akt loop [31]. These effects are significantly attenuated by pharmacological

blockage of PAFR or by inhibiting G protein incorporation into PAFR, a G protein-coupled receptor [28,31,42].

LOX-1, though originally cloned against copper-oxLDL, has a strong affinity for negatively charged particles through an electrostatic interaction [43,44]. We reported that, unlike L1, L5 is not recognized by LDLR on the plasma membrane of ECs and EPCs, but it is internalized by LOX-1 in a competitive manner against artificially prepared oxLDL [31,38]. Preliminary experiments suggest that the low pI values of non-apoB-100 protein molecules contribute to the switch in affinity from normal LDLR to LOX-1 (unpublished data), which is also inducible by L5 but not L1 [31,38].

The active components of L5 are not yet fully identified, but our current data suggest that they reside on lipids that accumulate in L5 particles. The total lipid extract derived from L5 is as efficient as L5 in inducing intracellular calcium transient in polymorphonuclear neutrophils (PMNs) [28]. Pretreatment of L5 with PAF acetylhydrolase to degrade PAF and PAF-like lipids via hydrolysis of sn-2 residues removes L5's capacity to downregulate FGF2 and induce EC apoptosis. Also, the lipid extract of degraded L5 fails to elicit calcium transient in PMNs, which is readily restored by adding exogenous PAF [28]. Findings suggest that protein configuration is important for receptor recognition but that lipid components, especially certain phospholipids, are responsible for signaling.

## 7. Evidence of L5's atherogenicity in vitro

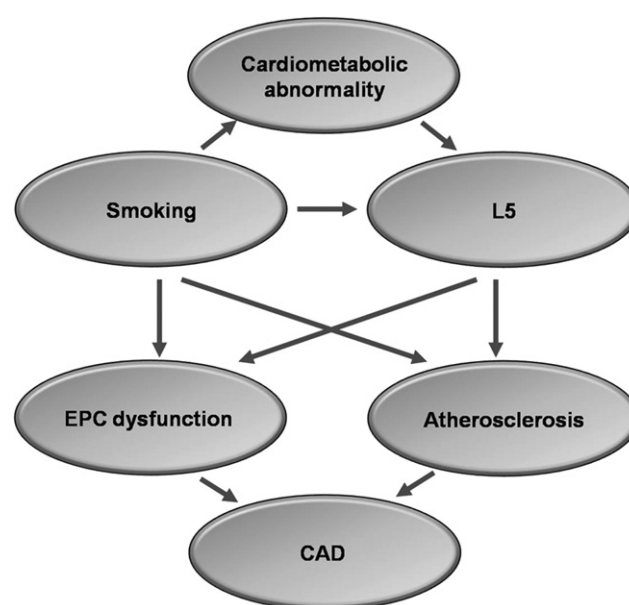
Apoptosis of the vascular endothelium contributes to increased transendothelial permeability [45,46]; released microparticles

enhance tissue factor expression and thus provoke coagulation [47,48]. Evidence that L5 induces EC apoptosis in a concentration- and time-dependent manner indicates L5's involvement in both early and late stages of atherothrombosis [28,30,31,35,39]. In addition to EC apoptosis, L5 induces monocyte-EC adhesion, another early event in atherosclerosis development [27,28]. Cell-to-cell adhesion is promoted by EC secretion of adhesion molecules, including VCAM-1, IL-8, and CXC chemokines [49], which is in agreement with what has been described for LDL(–) [20,50,51].

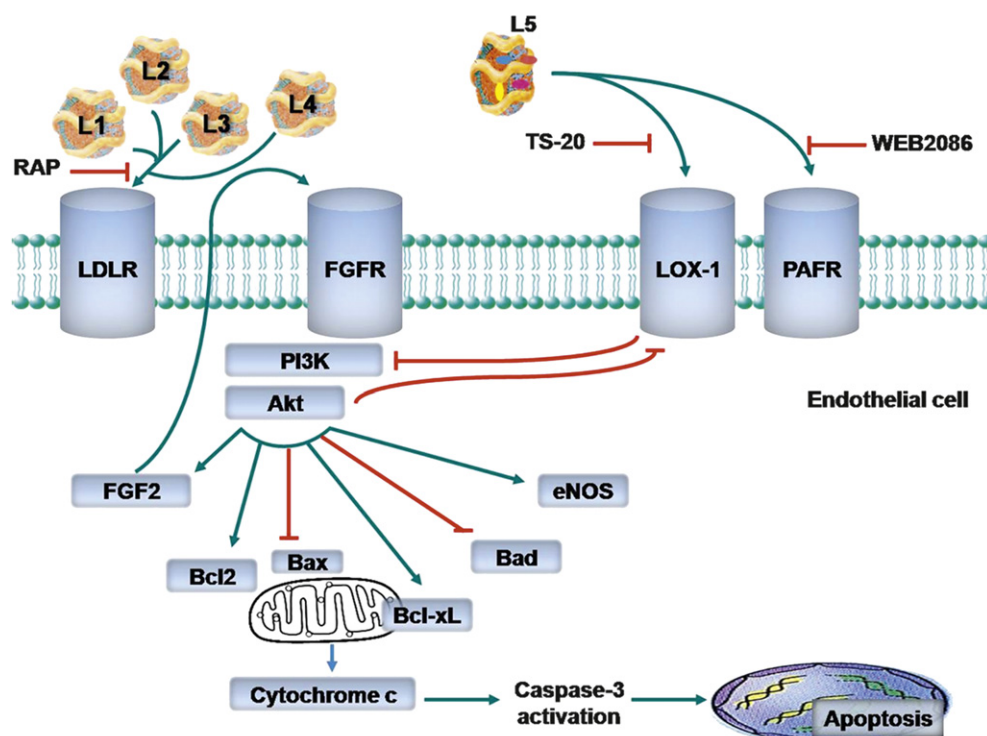
FGF2 functions by activating downstream kinases and effectors, including Akt, Bcl-2, Bad, Bax, Bcl-xL, and eNOS [52]. These effectors are major regulators of mitochondrial function and structural integrity [53]. We recently demonstrated that L5 suppresses the expression of the mitochondria-stabilizing and prosurvival effectors Bcl-2, Bcl-xL, and eNOS, as well as eNOS phosphorylation, and upregulates the proapoptotic effectors Bax, Bad, and TNF- $\alpha$  in vascular ECs through LOX-1 [39]. The L5-initiated signaling that leads to endothelial dysfunction and atherogenesis is summarized in the schematic illustration seen in Fig. 4.

## 8. L5 and EPC physiology

Evidence suggests that bone marrow-derived EPCs play a key role in endothelial regeneration, as well as in vasculogenesis



**Fig. 5 – Risk factors for L5 formation and the consequent effects on cardiovascular health.** Cardiometabolic abnormalities include hypercholesterolemia, type 2 diabetes mellitus, and metabolic syndrome. CAD = coronary artery disease; EPC = endothelial progenitor cell.



**Fig. 4 – Schematic summary of L5 signaling through LOX-1 in vascular ECs.** L5 signals and is internalized through LOX-1, whereas L1–L4 are endocytosed via LDLR. Green arrows indicate stimulation; blue arrows indicate release from mitochondria; red lines with end bars indicate inhibition. RAP = the LDLR inhibitor receptor-associated protein; TS-20 = LOX-1 neutralizing antibody; EC = endothelial cell; LDL = low-density lipoprotein; LDLR = LDL receptor; LOX-1 = lectin-like oxidized LDL receptor-1; FGFR = fibroblast growth factor receptor; PAFR = platelet-activating factor receptor; WEB2086 = thieno-triazolodiazepine, an antagonist of PAF; PI3K = phosphatidylinositol 3-kinase; Akt = protein kinase B; FGF2 = fibroblast growth factor 2; Bcl-2 = B-cell lymphoma 2; Bax = Bcl-2-associated X protein; Bcl-xL = B-cell lymphoma extra large; Bad = Bcl-2-associated death promoter; eNOS = endothelial nitric oxide synthase.



[54–56]. EPCs are reduced in number and/or functional activity in the presence of traditional and emerging major risk factors, whether separately or as clusters [57]. Reported correlated risk factors include aging [58], subclinical and clinical atherothrombotic disease [59,60], types 1 and 2 DM [61,62], hypercholesterolemia [63], smoking [57], and metabolic syndrome [64]. One common trait in people with these risk factors is the percentage increase of L5 in their plasma LDL [27,28,30,35,38,65,66]. At subapoptotic concentrations, L5 inhibited vascular endothelial growth factor (VEGF)-induced differentiation of human circulating monocytes into EPCs; this is achieved by suppression of Akt-mediated induction of VEGF kinase insert domain-containing receptor and other endothelial cell markers [38]. The impairment of mitogenic activity in early EPCs by abnormal LDL can accelerate EPC senescence [67]. Cellular senescence is critically influenced by telomerase, which elongates telomeres and thereby counteracts the telomere length reduction induced by each cell division [68,69]. In our setting, L5 accelerates EPC senescence by inhibiting telomerase activity [38], thus severely compromising the regenerative capacity of progenitor cells.

## 9. Conclusions and perspectives

Based on our findings and a review of the literature, we conclude that L5 is a non oxidized, naturally occurring atherogenic LDL that is not smaller or denser than other LDL subfractions. In preliminary experiments, repeated injection of human L5 into apoE knockout mice induced atherosclerotic changes in the aorta, and addition of L5 into organ chambers attenuated the endothelium-dependent relaxation of aortas that were removed from rats (data not shown). These *in vivo* and *ex vivo* observations support a role for L5 in endothelial dysfunction and atherosclerosis formation. Our recent data also suggest that (a) L5 is increased in patients undergoing acute coronary events and that (b) intracoronary thrombi contain tissue that exhibits strong LOX-1 expression. Further study is needed to confirm the role of L5 in atherothrombotic development in animals and in humans. Fig. 5 summarizes the links between risk factors that favor L5 formation and the consequences of L5 accumulation. Our ongoing studies aim to develop both diagnostic and therapeutic methods for the early detection and effective treatment of L5-mediated vascular disease.

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## REFERENCES

- [1] Witztum JL, Steinberg D. The oxidative modification hypothesis of atherosclerosis: does it hold for humans? *Trends Cardiovasc Med* 2001;11:93–102.
- [2] Binder CJ, Shaw PX, Chang MK, Boullier A, Hartvigsen K, Horkko S, et al. The role of natural antibodies in atherogenesis. *J Lipid Res* 2005;46:1353–63.
- [3] Chatterjee S, Berliner JA, Subbanagounder GG, Bhunia AK, Koh S. Identification of a biologically active component in minimally oxidized low density lipoprotein (MM-LDL) responsible for aortic smooth muscle cell proliferation. *Glycoconj J* 2004;20:331–8.
- [4] Libby P. Inflammation in atherosclerosis. *Nature* 2002;420:868–74.
- [5] Carmena R, Duriez P, Fruchart JC. Atherogenic lipoprotein particles in atherosclerosis. *Circulation* 2004;109:III2–7.
- [6] Koba S, Hirano T, Ito Y, Tsunoda F, Yokota Y, Ban Y, et al. Significance of small dense low-density lipoprotein-cholesterol concentrations in relation to the severity of coronary heart diseases. *Atherosclerosis* 2006;189:206–14.
- [7] Hoff HF, Karagas M, Heideman CL, Gaubatz JW, Gotto Jr AM. Correlation in the human aorta of APO B fractions with tissue cholesterol and collagen content. *Atherosclerosis* 1979;32:259–68.
- [8] Hoff HF, Gaubatz JW. Isolation, purification, and characterization of a lipoprotein containing Apo B from the human aorta. *Atherosclerosis* 1982;42:273–97.
- [9] Clevidence BA, Morton RE, West G, Dusek DM, Hoff HF. Cholesterol esterification in macrophages. Stimulation by lipoproteins containing apo B isolated from human aortas. *Arteriosclerosis* 1984;4:196–207.
- [10] Avogaro P, Bon GB, Cazzolato G. Presence of a modified low density lipoprotein in humans. *Arteriosclerosis* 1988;8:79–87.
- [11] Chappéy B, Myara I, Benoit MO, Maziere C, Maziere JC, Moatti N. Characteristics of ten charge-differing subfractions isolated from human native low-density lipoproteins (LDL). No evidence of peroxidative modifications. *Biochim Biophys Acta* 1995;1259:261–70.
- [12] Demuth K, Myara I, Chappéy B, Védie B, Pech-Amsellem MA, Haberland ME, et al. A cytotoxic electronegative LDL subfraction is present in human plasma. *Arterioscler Thromb Vasc Biol* 1996;16:773–83.
- [13] Sanchez-Quesada JL, Perez A, Caixas A, Ordonmez-Llanos J, Carreras G, Payes A, et al. Electronegative low density lipoprotein subform is increased in patients with short-duration IDDM and is closely related to glycaemic control. *Diabetologia* 1996;39:1469–76.
- [14] Cordoba-Porras A, Sanchez-Quesada JL, Gonzalez-Sastre F, Ordonez-Llanos J, Blanco-Vaca F. Susceptibility of plasma low- and high-density lipoproteins to oxidation in patients with severe hyperhomocysteinemia. *J Mol Med* 1996;74:771–6.
- [15] Védie B, Jeunemaitre X, Megnien JL, Myara I, Trebeden H, Simon A, Moatti N. Charge heterogeneity of LDL in asymptomatic hypercholesterolemic men is related to lipid parameters and variations in the ApoB and CIII genes. *Arterioscler Thromb Vasc Biol* 1998;18:1780–9.
- [16] Sanchez-Quesada JL, Otal-Entraigas C, Franco M, Jorba O, Gonzalez-Sastre F, Blanco-Vaca F, et al. Effect of simvastatin treatment on the electronegative low-density lipoprotein present in patients with heterozygous familial hypercholesterolemia. *Am J Cardiol* 1999;84:655–9.
- [17] Fabjan JS, Abuja PM, Schaur RJ, Sevanian A. Hypochlorite induces the formation of LDL(-), a potentially atherogenic low density lipoprotein subspecies. *FEBS Lett* 2001;499:69–72.

- [18] Benitez S, Sanchez-Quesada JL, Ribas V, Jorba O, Blanco-Vaca F, Gonzalez-Sastre F, et al. Platelet-activating factor acetylhydrolase is mainly associated with electronegative low-density lipoprotein subfraction. *Circulation* 2003;108:92–6.
- [19] Ziouzenkova O, Asatryan L, Sahady D, Orasanu G, Perrey S, Cutak B, et al. Dual roles for lipolysis and oxidation in peroxisome proliferation-activator receptor responses to electronegative low density lipoprotein. *J Biol Chem* 2003; 278:39874–81.
- [20] Sanchez-Quesada JL, Benitez S, Ordonez-Llanos J. Electronegative low-density lipoprotein. *Curr Opin Lipidol* 2004;15:329–35.
- [21] Barros MR, Bertolami MC, Abdalla DS, Ferreira WP. Identification of mildly oxidized low-density lipoprotein (electronegative LDL) and its auto-antibodies IgG in children and adolescents hypercholesterolemic offsprings. *Atherosclerosis* 2006;184:103–7.
- [22] Oliveira JA, Sevanian A, Rodrigues RJ, Apolinario E, Abdalla DS. Minimally modified electronegative LDL and its autoantibodies in acute and chronic coronary syndromes. *Clin Biochem* 2006;39:708–14.
- [23] Bancells C, Sanchez-Quesada JL, Birkelund R, Ordonez-Llanos J, Benitez S. HDL and electronegative LDL exchange anti- and pro-inflammatory properties. *J Lipid Res* 2010;51:2947–56.
- [24] Brunelli R, Balogh G, Costa G, De Spirito M, Greco G, Mei G, et al. Estradiol binding prevents ApoB-100 misfolding in electronegative LDL(-). *Biochemistry* 2010;49:7297–302.
- [25] Bancells C, Villegas S, Blanco FJ, Benitez S, Gallego I, Beloki L, et al. Aggregated electronegative low density lipoprotein in human plasma shows a high tendency toward phospholipolysis and particle fusion. *J Biol Chem* 2010;285:32425–35.
- [26] Bancells C, Canals F, Benitez S, Colome N, Julve J, Ordonez-Llanos J, et al. Proteomic analysis of electronegative low-density lipoprotein. *J Lipid Res* 2010;51:3508–15.
- [27] Yang CY, Raya JL, Chen HH, Chen CH, Abe Y, Pownall HJ, et al. Isolation, characterization, and functional assessment of oxidatively modified subfractions of circulating low-density lipoproteins. *Arterioscler Thromb Vasc Biol* 2003;23:1083–90.
- [28] Chen CH, Jiang T, Yang JH, Jiang W, Lu J, Marathe GK, et al. Low-density lipoprotein in hypercholesterolemic human plasma induces vascular endothelial cell apoptosis by inhibiting fibroblast growth factor 2 transcription. *Circulation* 2003;107:2102–8.
- [29] Bessell EM, Thomas P. The effect of substitution at C-2 of D-glucose 6-phosphate on the rate of dehydrogenation by glucose 6-phosphate dehydrogenase (from yeast and from rat liver). *Biochem J* 1973;131:83–9.
- [30] Yang CY, Chen HH, Huang MT, Raya JL, Yang JH, Chen CH, et al. Pro-apoptotic low-density lipoprotein subfractions in type II diabetes. *Atherosclerosis* 2007;193:283–91.
- [31] Lu J, Jiang W, Yang JH, Chang PY, Walterscheid JP, Chen HH, et al. Electronegative LDL impairs vascular endothelial cell integrity in diabetes by disrupting fibroblast growth factor 2 (FGF2) autoregulation. *Diabetes* 2008;57:158–66.
- [32] Chang YH, Abdalla DS, Sevanian A. Characterization of cholesterol oxidation products formed by oxidative modification of low density lipoprotein. *Free Radic Biol Med* 1997;23:202–14.
- [33] Campos E, Nakajima K, Tanaka A, Havel RJ. Properties of an apolipoprotein E-enriched fraction of triglyceride-rich lipoproteins isolated from human blood plasma with a monoclonal antibody to apolipoprotein B-100. *J Lipid Res* 1992;33:369–80.
- [34] Kawakami A, Yoshida M. Remnant lipoproteins and atherogenesis. *J Atheroscler Thromb* 2005;12:73–6.
- [35] Chen HH, Hosken BD, Huang M, Gaubatz JW, Myers CL, Macfarlane RD, et al. Electronegative LDLs from familial hypercholesterolemic patients are physicochemically heterogeneous but uniformly proapoptotic. *J Lipid Res* 2007; 48:177–84.
- [36] Cromwell WC, Otvos JD. Low-density lipoprotein particle number and risk for cardiovascular disease. *Curr Atheroscler Rep* 2004;6:381–7.
- [37] Rizzo M, Berneis K. Low-density lipoprotein size and cardiovascular risk assessment. *QIM* 2006;99:1–14.
- [38] Tang D, Lu J, Walterscheid JP, Chen HH, Engler DA, Sawamura T, et al. Electronegative LDL circulating in smokers impairs endothelial progenitor cell differentiation by inhibiting Akt phosphorylation via LOX-1. *J Lipid Res* 2008; 49:33–47.
- [39] Lu J, Yang JH, Burns AR, Chen HH, Tang D, Walterscheid JP, et al. Mediation of electronegative low-density lipoprotein signaling by LOX-1: a possible mechanism of endothelial apoptosis. *Circ Res* 2009;104:619–27.
- [40] Rommel C, Clarke BA, Zimmermann S, Nunez L, Rossman R, Reid K, et al. Differentiation stage-specific inhibition of the Raf-MEK-ERK pathway by Akt. *Science* 1999;286:1738–41.
- [41] Chavakis E, Dernbach E, Hermann C, Mondorf UF, Zeiher AM, Dimmeler S. Oxidized LDL inhibits vascular endothelial growth factor-induced endothelial cell migration by an inhibitory effect on the Akt/endothelial nitric oxide synthase pathway. *Circulation* 2001;103:2102–7.
- [42] Chang PY, Luo S, Jiang T, Lee YT, Lu SC, Henry PD, et al. Oxidized low-density lipoprotein downregulates endothelial basic fibroblast growth factor through a pertussis toxin-sensitive G-protein pathway: mediator role of platelet-activating factor-like phospholipids. *Circulation* 2001;104: 588–93.
- [43] Sawamura T, Kume N, Aoyama T, Moriwaki H, Hoshikawa H, Aiba Y, et al. An endothelial receptor for oxidized low-density lipoprotein. *Nature* 1997;386:73–7.
- [44] Chen M, Inoue K, Narumiya S, Masaki T, Sawamura T. Requirements of basic amino acid residues within the lectin-like domain of LOX-1 for the binding of oxidized low-density lipoprotein. *FEBS Lett* 2001;499:215–9.
- [45] Chen YL, Jan KM, Lin HS, Chien S. Relationship between endothelial cell turnover and permeability to horseradish peroxidase. *Atherosclerosis* 1997;133:7–14.
- [46] von Eckardstein A, Rohrer L. Transendothelial lipoprotein transport and regulation of endothelial permeability and integrity by lipoproteins. *Curr Opin Lipidol* 2009;20:197–205.
- [47] Agouni A, Lagrue-Lak-Hal AH, Ducluzeau PH, Mostefai HA, Draunet-Busson C, Leftheriotis GH, et al. Endothelial dysfunction caused by circulating microparticles from patients with metabolic syndrome. *Am J Pathol* 2008;173:1210–9.
- [48] Tedgui A, Mallat Z. Apoptosis as a determinant of atherothrombosis. *Thromb Haemost* 2001;86:420–6.
- [49] Abe Y, Fornage M, Yang CY, Bui-Thanh NA, Wise V, Chen HH, et al. LS, the most electronegative subfraction of plasma LDL, induces endothelial vascular cell adhesion molecule 1 and CXC chemokines, which mediate mononuclear leukocyte adhesion. *Atherosclerosis* 2007;192:56–66.
- [50] Sanchez-Quesada JL, Camacho M, Anton R, Benitez S, Vila L, Ordonez-Llanos J. Electronegative LDL of FH subjects: chemical characterization and induction of chemokine release from human endothelial cells. *Atherosclerosis* 2003; 166:261–70.
- [51] Sanchez-Quesada JL, Benitez S, Perez A, Wagner AM, Rigla M, Carreras G, et al. The inflammatory properties of electronegative low-density lipoprotein from type 1 diabetic patients are related to increased platelet-activating factor acetylhydrolase activity. *Diabetologia* 2005;48:2162–9.
- [52] Chen CH, Poucher SM, Lu J, Henry PD. Fibroblast growth factor 2: from laboratory evidence to clinical application. *Curr Vasc Pharmacol* 2004;2:33–43.

- [53] Holinger EP, Chittenden T, Lutz RJ. Bak BH3 peptides antagonize Bcl-xL function and induce apoptosis through cytochrome c-independent activation of caspases. *J Biol Chem* 1999;274:13298–304.
- [54] Dimmeler S, Aicher A, Vasa M, Mildner-Rihm C, Adler K, Tiemann M, et al. HMG-CoA reductase inhibitors (statins) increase endothelial progenitor cells via the PI 3-kinase/Akt pathway. *J Clin Invest* 2001;108:391–7.
- [55] Crosby JR, Kaminski WE, Schatteman G, Martin PJ, Raines EW, Seifert RA, et al. Endothelial cells of hematopoietic origin make a significant contribution to adult blood vessel formation. *Circ Res* 2000;87:728–30.
- [56] Asahara T, Murohara T, Sullivan A, Silver M, van der Zee R, Li T, et al. Isolation of putative progenitor endothelial cells for angiogenesis. *Science* 1997;275:964–7.
- [57] Vasa M, Fichtlscherer S, Aicher A, Adler K, Urbich C, Martin H, et al. Number and migratory activity of circulating endothelial progenitor cells inversely correlate with risk factors for coronary artery disease. *Circ Res* 2001;89:E1–7.
- [58] Hoetzer GL, Van Guilder GP, Irmiger HM, Keith RS, Stauffer BL, DeSouza CA. Aging, exercise, and endothelial progenitor cell clonogenic and migratory capacity in men. *J Appl Physiol* 2007;102:847–52.
- [59] Guven H, Shepherd RM, Bach RG, Capoccia BJ, Link DC. The number of endothelial progenitor cell colonies in the blood is increased in patients with angiographically significant coronary artery disease. *J Am Coll Cardiol* 2006;48:1579–87.
- [60] Schwartzberg S, Deutsch V, Maysel-Auslender S, Kissil S, Keren G, George J. Circulating apoptotic progenitor cells: a novel biomarker in patients with acute coronary syndromes. *Arterioscler Thromb Vasc Biol* 2007;27:e27–31.
- [61] Loomans CJ, de Koning EJ, Staal FJ, Rookmaaker MB, Verseyden C, de Boer HC, et al. Endothelial progenitor cell dysfunction: a novel concept in the pathogenesis of vascular complications of type 1 diabetes. *Diabetes* 2004;2004(53):195–9.
- [62] Tepper OM, Galiano RD, Capla JM, Kalka C, Gagne PJ, Jacobowitz GR, et al. Human endothelial progenitor cells from type II diabetics exhibit impaired proliferation, adhesion, and incorporation into vascular structures. *Circulation* 2002;106:2781–6.
- [63] Chen JZ, Zhang FR, Tao QM, Wang XX, Zhu JH. Number and activity of endothelial progenitor cells from peripheral blood in patients with hypercholesterolaemia. *Clin Sci (Lond)* 2004;107:273–80.
- [64] Hill JM, Zalos G, Halcox JP, Schenke WH, Waclawiw MA, Quyyumi AA, et al. Circulating endothelial progenitor cells, vascular function, and cardiovascular risk. *N Engl J Med* 2003;348:593–600.
- [65] Tai MH, Kuo SM, Liang HT, Chiou KR, Lam HC, Hsu CM, et al. Modulation of angiogenic processes in cultured endothelial cells by low density lipoproteins subfractions from patients with familial hypercholesterolemia. *Atherosclerosis* 2006;186:448–57.
- [66] Lu J, Jiang W, Yang JH, Chang PY, Walterscheid JP, Chen HH, et al. Electronegative LDL impairs vascular endothelial cell integrity in diabetes by disrupting FGF2 autoregulation. *Diabetes* 2008;57:158–66.
- [67] Imanishi T, Hano T, Sawamura T, Nishio I. Oxidized low-density lipoprotein induces endothelial progenitor cell senescence, leading to cellular dysfunction. *Clin Exp Pharmacol Physiol* 2004;31:407–13.
- [68] Greider CW, Blackburn EH. Identification of a specific telomere terminal transferase activity in *Tetrahymena* extracts. *Cell* 1985;43:405–13.
- [69] Greider CW, Blackburn EH. A telomeric sequence in the RNA of *Tetrahymena* telomerase required for telomere repeat synthesis. *Nature* 1989;337:331–7.



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## Review article

# Androgen and androgen receptor signals jamming monocyte/macrophage functions in premalignant phase of livers

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## ABSTRACT

There is a widely discussed concept that chronic inflammation and repeated damage-repair cycles result in malignant transformation of the liver. Kupffer cells are the major host defending macrophages that reside in the liver. They are also considered as critical in episodes of cancer immune surveillance that wipe out liver malignancies, yet, turn into cancer assisting cells in certain conditions. Monocyte/macrophage population and cytokine profile hierarchization in hepatocellular carcinoma (HCC) has increased the curiosity to search for macrophage modulators. Androgenic signals [androgen/androgen receptors (A/AR)] play an important role in liver function and disease progression. Basic and clinical studies have revealed that A/AR might play a biphasic function in HCC progression. Whether A/AR work with TAM to further manipulate HCC progression is of great interest. In this review article, we will focus on the interaction of hepatocytes and monocytes/macrophages in preneoplastic/ inflammatory liver diseases, underlining A/AR actions.

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## 1. Introduction

### 1.1. Androgen/androgen receptor (A/AR) roles in hepatic host immunity

The liver is the largest visceral organ and is responsible for systemic homeostasis such as blood glucose, lipid and protein metabolism and the clearance of xenobiotics [1]. There are several different cell types found in a liver unit: hepatocytes, Ito

cells (lipocyte), Kupffer cells (monocytes), and oval/stellate cells (fibroblast) [1]. The hepatic sinusoids are unique structures that are lined with a thin discontinuous endothelium. Kupffer cells are part of this thin line and are responsible for scavenging hepatic debris for the hepatic host immune response. Liver is under a constant immunological challenge, and the immunological response is dominated by innate immunological components including macrophages, dendritic cells, natural killer cells, natural killer T cells, complement

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components, acute phase proteins, and chemokines [2]. The immune surveillance function (tolerant to daily food absorption to the liver) is largely executed within hepatic reticuloendothelial cells, which are largely composed of Kupffer cells. The cell surface receptors, e.g., TLR, complement receptor and Fc-receptors on Kupffer cells, are known to be largely responsible for sensing innate immunological responses [2,3]. However, little is known in diet-induced liver damage.

During acute illnesses, acute innate immunity could be activated by the activation of macrophages, and by producing certain acute phase proteins. Macrophages and lymphocytes could secrete inflammatory cytokines such as tumor necrosis factor- $\alpha$  (TNF $\alpha$ ), interleukin-1 (IL-1), and interleukin-6 (IL-6) when antigens are present in the organs. Interacting with complement pathways, the complex could bring microbes to phagocytes, cause macrophage cytolysis, and chemotactically attract phagocytes to the infected area. Inflammatory cytokines travel through the blood and stimulate hepatocytes in the liver to synthesize and secrete acute phase proteins. This response provides an early defense and enables the body to recognize foreign substances early on in the infection process, prior to the full activation and implementation of the immune responses [2,3].

Using an AR knockout animal model, Lin et al explored the potential roles of A/AR signals in food-induced hepato steatosis and the relative metabolic syndrome in the mouse [4]. In this report, Lin et al pointed out that AR could act as protein-tyrosine phosphatase 1B (PTP1B) suppressors, which suppress insulin signaling and inhibit  $\beta$ -oxidation of lipids in hepatocytes. However, as previously described, Kupffer cells are host immune confounders against metabolic and chronic steatosis of the liver. Introducing the 5 $\alpha$ -reductase inhibitor, 4-hydroxyandrostenedione (4-OHA), suppresses testosterone conversion to 5 $\alpha$ -dihydrotestosterone (DHT) in mice and could reduce Kupffer cells cytokine production, but not splenic macrophages [5]. Their studies provide some evidence to suggest that A/AR might also play a role in the overindulgent stress in which gate-keeping by host immune defense in the liver.

## 1.2. A/AR roles in the hepatic damage/regenerating state

Liver regeneration is the homeostatic machinery to recover the liver structure and function after physical or chemical hazards [6]. Distinguishable from other regenerative organs (skin, bone marrow), the liver regenerates from large populations of mature cells, and not from small amounts of stem/progenitor cells [6,7]. Several distinct cells, such as hepatocytes (the main functional cells of the liver), cholangiocytes (biliary epithelial cells), fenestrated endothelial cells (gates between blood fluid and hepatocytes), Kupffer cells (liver macrophages), and Ito cells (stellate cells; extracellular matrix secretion, growth factors production, and vitamin A storage) [6] are involved in the healing process. Among them, hepatocytes are the primary and most important initiators during the whole liver regeneration process.

There are several animal models using rodents for studying liver regeneration. The most common is the removal of two-thirds of the liver (partial hepatectomy; PHx) to induce

liver regeneration. Another is the injection of the hepatic toxin, CCl<sub>4</sub>, to facilitate hepatic proliferation. Results from these animal studies suggested that proliferation within the hepatic parenchyma starts from the portal triad to the pericentral area within 48 hours. Several immediate early response genes, such as NF $\kappa$ B and STAT3, could be activated rapidly (within 60 minutes) after PHx [8,9]. Other factors involved in liver regeneration after PHx are the growth factors and cytokines, such as hepatocyte growth factor (HGF), tumor necrosis factor  $\alpha$  (TNF $\alpha$ ), interleukine-6 (IL-6), epidermal growth factor and transforming growth factor  $\alpha$  [10]. Nuclear receptor signals, such as thyroid hormone [11], retinoid acids [12], and glucocorticoid [13] as well as estrogens and androgens, also play important roles in liver regeneration.

In general, the liver regenerative response is better in females than in males [14], suggesting that A/AR signals may play a negative role in regenerating the liver. The decreased serum androgen levels, accompanied by down-regulation of AR protein in the liver during liver regeneration, also implicates the negative role of A/AR signals [15,16]. In addition, Kan et al, found that using flutamide (an androgen action antagonist) can protect trauma-hemorrhage-induced liver injury through reducing the systemic inflammatory response [17]. Published by same group, Schneider et al found that trauma and hemorrhage-shock induced TNF $\alpha$  and IL-6 production is by Kupffer cells and splenic macrophages [5]. In other words, in the normal liver, A/AR might balance the other stimulating signals, such as growth factors or prolactin [18–21]. Once the liver has been damaged, the homeostasis machinery tends to suppress negative factors, i.e., A/AR signals, to facilitate liver regeneration.

However, other reports also suggested that A/AR signals might play a small role in the liver regeneration. For example, the antiandrogen, cimetidine, showed little effects on PHx rat liver [22], and administration of tamoxifen in male hepatectomized rats resulted in increased AR activity, yet had little effect on liver regeneration [18]. Similarly, the addition of the antiandrogen, flutamide, resulted in initial overexpression of AR, yet had little effect on liver regeneration in PHx rats [23].

Kahn et al [22] also demonstrated that in the rat liver, hepatic injury with a portacaval shunt results in minor effects on AR activity, and partial clamp of the portal vein or clamp of the hepatic artery yields little effect on AR activity. However, using CCl<sub>4</sub> toxication in the liver, Smirnova et al [19,20] and Shchelkunova et al [25] found that A/AR signals might indirectly facilitate liver regeneration through modulation of a specific protein, unusual estrogen-binding protein (UEBP), in the hepatocytes that can increase the uptake of estrogen.

The above contradictory effects (suppression vs. promotion vs. little influence) of A/AR signals using either androgen administration or surgical castration to test the liver regeneration ability in different animal models (PHx or CCl<sub>4</sub> toxication) implied the complexity of A/AR signals during the liver regeneration process. There is no perfect explanation for the contradictory effects of A/AR signals between liver regeneration models. Whether the differential secretion of growth factors or cytokines that were induced by surgical hepatectomy, or chemical toxication in different animal models, results in such diverse A/AR responses, is an interesting question for further study.

### 1.3. A/AR signals and hepatitis B virus (HBV) virus antigen expression and replication

The evidence that A/AR signals influence HBV replication was based on HBV transgenic mice studies. Although HBV virions, HBsAg and HBeAg, can be detected in HBV transgenic mice, the mice did not exhibit pathological changes because of the immune tolerance. The high HBV DNA amount is associated with the occurrence of human HCC, so it is important to know how A/AR signals affect HBV antigen expression and replication in mice. It has been found that serum HBsAg concentration is higher in male than in female HBV transgenic mice. Castration of male mice could eliminate this sexual dimorphism, while supplementation with testosterone could restore the difference [26–28]. Furthermore, in order to determine the effects of AR on the HBV virus, Breidbart et al [26], using testicular feminization mutation (Tfm) mice, in which 90% of AR was reduced, found that the serum HBsAg concentration is higher in wild type mice compared with XYTfm mice. Except for direct A/AR signals on regulating HBV antigen expression [29], more evidence is required for the effect of A/AR signals in HBV-related HCC. A previous bottleneck of HBV-HCC studies was due to the lack of proper animal models in which HCC spontaneously develops from hepatitis B. Woodchuck hepatitis virus (WHV) is similar to the human HBV, both in structure and replicative viral life cycle. WHV infection can cause acute and chronic hepatitis. Chronic WHV infection in woodchuck will develop into HCC within the first 2 to 4 years [30]. Therefore, the WHV model might allow us to know more about the effects of A/AR in the progression of HBV infection and the related liver disease. Recent studies showed that androgenic signals promote HBV viral replication through direct regulation of HBV replication [31–34]. Furthermore, Wu et al, found that HBV transgenic mice, supplemented with a subminimal dose of carcinogen, can induce spontaneous HCC development, while knockout hepatic AR could reduce HBV-related hepatocarcinogenesis through direct regulation of HBV virus replication [31].

### 1.4. A/AR signals in cirrhotic livers

Cirrhosis is the pathological feature that can be observed in diverse liver diseases, generally arising from chronic liver injury. During the injury healing process, the liver could develop fibrotic lesions that may lead to the loss of normal hepatic function, impaired liver regeneration, aberrant polarity for cells to proliferate, and obstruction of the portal system's ability to secrete bile acid. Based on the population epidemiology, liver cirrhosis can be classified into three categories: (1) alcoholic-related cirrhosis (ARC); (2) virus-induced hepatic cirrhosis; and (3) non-alcoholic cirrhosis.

The linkage of male hypogonadism and hypotestosteronemia with ARC victims [35] suggests that A/AR signals might play some role in the ARC. Reduced testicular size and the clinical features of inadequate testicular function are manifested as clinical hypogonadism in cirrhotic men. Between 50% and 75% of cirrhotic men have both macroscopic and histological testicular atrophy. Associated with this, 80%–90% of cirrhotic men are impotent, and seminal fluid characteristics are grossly abnormal in the small

minority of patients who are able to produce an ejaculation [35]. Furthermore, cirrhotic men have a decreased incidence of benign prostatic hypertrophy, and gynecomastia could also be found in about 40% of cirrhotic men [24].

Testosterone administration can improve both hypogonadism and gynecomastia syndrome effectively, however, it may have little effect in improving cirrhosis [36]. This suggests that the decrease of testosterone, due to irreversible damage of the liver, may be a consequence of cirrhosis. Interestingly, Kley [37] found that administration of testosterone to male ARC might yield some improvement. In agreement with this, Thole et al [38] also reported that administration of steroidal or non-steroidal antiandrogens, such as cimetidine or flutamide, might lead to cirrhosis [24], although the potential toxicity of these antiandrogens might also contribute to such cirrhosis.

In contrast, Gluud C et al [39,40] found that oral testosterone treatment yielded little change in the liver pathogenesis of ARC men, even when such treatment could significantly reduce the prevalence of gynecomastia. Similarly, a large population in a randomized clinical trial also found androgens, such as testosterone and oxandrolone administration could not improve ARC patients.

## 2. Concluding remark: Illustrating the relations within Kupffer cells, A/AR and hepatocarcinogenesis

Recent reports on A/AR signals in hepatocarcinogenesis have gained advances regarding carcinogen- and HBV-related liver malignancy [31,41,42]. However, the bimodal function of A/AR has also been described in HCC progression [43]. Although there is little information regarding A/AR signals in the immune surveillance of hepatocarcinogenesis, A/AR signals did have documented in prostatic malignancy in terms of innate and adaptive immunities. Published by Lai et al, in the immune cell-specific AR knockout mice described, A/AR is pivotal for both innate and adaptive immune regulations [50,51].

It has been reported that innate immunity plays a pivotal role in hepatocarcinogenesis. By removing NF- $\kappa$ B signals in the hepatocytes, the mouse became resistant to carcinogen-induced HCC [44]. This result clearly demonstrated the linkage of innate immunity to HCC. The Kupffer cells population is not increased in the cancer lesion. The Kupffer cell number may even be decreased [45]. These conflicting findings could lead to the conclusion that innate immunity might be an important factor, however, the innate immunity does not necessarily require invasive macrophages into the tumor lesion. Previous studies, in both humans and rodents, have shown that tumor necrosis factor alpha (TNF $\alpha$ ) is an important mediator of liver injury [46–48]. Although many types of cells in the liver are capable of producing TNF $\alpha$ , Kupffer cells are thought to be the main hepatic source of TNF $\alpha$ . They also produce factors such as IL-12 and IL-18 and interferon gamma, which enhance TNF activity, as well as those that inhibit TNF $\alpha$ , such as IL-10 [49]. The role of A/AR in the whole process of HCC development and progression is still a mystery. Whether A/AR and TAM participate in the hepatocarcinogenesis process is yet to be explored.

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## REFERENCES

- [1] Boyer TD, TLW, Manns MP. Zakim and Boyer's hepatology. A textbook of liver disease. Canada: Elsevier; 2006.
- [2] Gershwin ME, Vierling JM, Manns MP, O'Farrelly C, Doherty DG. Liver immunology: principles and practice. Humana Press; 2007.
- [3] Sompayrac LM. How the immune system works. Wiley-Blackwell; 2008.
- [4] Lin HY, Yu IC, Wang RS, Chen YT, Liu NC, Altuwaijri S, et al. Increased hepatic steatosis and insulin resistance in mice lacking hepatic androgen receptor. *Hepatology* 2008;47: 1924–35.
- [5] Schneider CP, Schwacha MG, Samy TS, Bland KI, Chaudry IH. Androgen-mediated modulation of macrophage function after trauma-hemorrhage: central role of 5 $\alpha$ -dihydrotestosterone. *J Appl Physiol* 2003;95:104–12.
- [6] Schiff ER, MFS, Maddrey WC. Schiff's disease of the liver. Lippincott Williams & Wilkins; 2003.
- [7] Otu HH, Naxerova K, Ho K, Can H, Nesbitt N, Libermann TA, et al. Restoration of liver mass after injury requires proliferative and not embryonic transcriptional patterns. *J Biol Chem* 2007;282:11197–204.
- [8] Cressman DE, Greenbaum LE, Haber BA, Taub R. Rapid activation of post-hepatectomy factor/nuclear factor kappa B in hepatocytes, a primary response in the regenerating liver. *J Biol Chem* 1994;269:30429–35.
- [9] Cressman DE, Diamond RH, Taub R. Rapid activation of the Stat3 transcription complex in liver regeneration. *Hepatology* 1995;21:1443–9.
- [10] Webber EM, Wu JC, Wang L, Merlino G, Fausto N. Overexpression of transforming growth factor- $\alpha$  causes liver enlargement and increased hepatocyte proliferation in transgenic mice. *Am J Pathol* 1994;145:398–408.
- [11] Nakamura RM, Miyada DS, Moyer DL. Effect of liver regeneration following partial hepatectomy on the uptake of tritiated thymidine in the pituitary gland of the rat. *Nature* 1963;199:707–8.
- [12] Ozeki A, Tsukamoto I. Retinoic acid repressed the expression of c-fos and c-jun and induced apoptosis in regenerating rat liver after partial hepatectomy. *Biochim Biophys Acta* 1990; 1450:308–19.
- [13] Richman RA, Claus TH, Pilgis SJ, Friedman DL. Hormonal stimulation of DNA synthesis in primary cultures of adult rat hepatocytes. *Proc Natl Acad Sci U S A* 1976;73:3589–93.
- [14] Yokoyama Y, Nagino M, Nimura Y. Which gender is better positioned in the process of liver surgery? Male or female? *Surg Today* 2007;37:823–30.
- [15] Yamaguchi M, Yu L, Nazmy El-Assal O, Satoh T, Kumar Dhar D, Yamanoi A, et al. Androgen metabolism in regenerating liver of male rats: evidence for active uptake and utilization of testosterone. *Hepatol Res* 2001;20:114–27.
- [16] Tsukamoto I, Kojo S. The sex difference in the regulation of liver regeneration after partial hepatectomy in the rat. *Biochim Biophys Acta* 1990;1033:287–90.
- [17] Kan WH, Hsieh CH, Schwacha MG, Choudhry MA, Raju R, Bland KI, et al. Flutamide protects against trauma-hemorrhage-induced liver injury via attenuation of the inflammatory response, oxidative stress, and apoptosis. *J Appl Physiol* 2008;105:595–602.
- [18] Kahn D, Eagon PK, Porter LE, Elm MS, Makowka L, Podesta L, et al. Effect of tamoxifen on hepatic regeneration in male rats. *Dig Dis Sci* 1989;34:27–32.
- [19] Smirnova OV, Vishnyakova TG, Bocharov AV, Kovtun IV, Rozen VB. Evidence for direct action of testosterone on rat liver cells: *in vivo* and *in vitro* induction of unusual estrogen-binding protein. *J Steroid Biochem Mol Biol* 1992;42:243–9.
- [20] Smirnova OV, Kovtun IV, Smirnov AN, Shchelkunova TA, Faktor VM, Rozen VB. Inheritance of androgen program of male-specific expression of unusual estrogen-binding protein by daughter hepatocytes at rat liver regeneration. *J Steroid Biochem Mol Biol* 1993;44:155–62.
- [21] Kahn D, Gavalier JS, Makowka L, Chapchap P, Mazzaferro V, Casavilla A. Does hyperprolactinemia affect hepatic regeneration independent of sex steroids? *J Lab Clin Med* 1988;112:644–51.
- [22] Kahn D, Svanas GW, Eagon PK, Makowka L, Podesta L, Chapchap P, et al. Effect of an antiandrogenic H2 receptor antagonist on hepatic regeneration in rats. *J Lab Clin Med* 1988;112:232–9.
- [23] Svanas GW, Eagon PK, Elm M, Makowka L, Podesta L, Chapchap P, et al. Effect of antiandrogen flutamide on measures of hepatic regeneration in rats. *Dig Dis Sci* 1989;34: 1916–23.
- [24] Yoshitsugu M, Ihori M. Endocrine disturbances in liver cirrhosis—focused on sex hormones. *Nippon Rinsho* 1997;55: 3002–6.
- [25] Shchelkunova TA, Smirnov AN, Faktor VM, Brodskii V, Rozen VB. An androgenic program of expression of the particular estrogen-binding protein is present in male rats in all hepatocytes. *Ontogene* 1993;24:70–5.
- [26] Breidbart S, Burk RD, Saenger P. Hormonal regulation of hepatitis B virus gene expression: influence of androgen receptor. *Pediatr Res* 1993;34:300–2.
- [27] DeLoia JA, Burk RD, Gearhart JD. Developmental regulation of hepatitis B surface antigen expression in two lines of hepatitis B virus transgenic mice. *J Virol* 1989;63:4069–73.
- [28] Farza H, Salmon AM, Hadchouel M, Moreau JL, Babinet C, Tiollais P, et al. Hepatitis B surface antigen gene expression is regulated by sex steroids and glucocorticoids in transgenic mice. *Proc Natl Acad Sci U S A* 1987;84:1187–91.
- [29] Tian Y, Kuo CF, Chen WL, Ou JH. Enhancement of hepatitis B virus replication by androgen and its receptor in mice. *J Virol* 2010;86:1904–10.
- [30] Popper H, Roth L, Purcell RH, Tennant BC, Gerin JL. Hepatocarcinogenicity of the woodchuck hepatitis virus. *Proc Natl Acad Sci U S A* 1987;84:866–70.
- [31] Wu MH, Hsu CL, Chen YL, Ou JH, Ryan CK, Hung YC, et al. Androgen receptor promotes hepatitis B virus-induced hepatocarcinogenesis through modulation of hepatitis B virus RNA transcription. *Sci Transl Med* 2010;2:32ra35.
- [32] Yang WJ, Chang CJ, Yeh SH, Lin WH, Wang SH, Tsai TF, et al. Hepatitis B virus X protein enhances the transcriptional activity of the androgen receptor through c-Src and glycogen synthase kinase-3 $\beta$  kinase pathways. *Hepatology* 2009;49:1515–24.
- [33] Yeh SH, Chen PJ. Gender disparity of hepatocellular carcinoma: the roles of sex hormones. *Oncology* 2010; 78(Suppl. 1):172–9.
- [34] Zhu R, Zhang JS, Zhu YZ, Fan J, Mao Y, Chen Q, et al. HBx-induced androgen receptor expression in HBV-



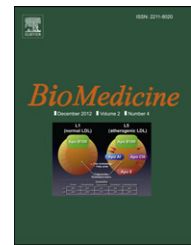
- associated hepatocarcinoma is independent of the methylation status of its promoter. *Histol Histopathol* 2011;26:23–35.
- [35] Green GR. Mechanism of hypogonadism in cirrhotic males. *Gut* 1977;18:843–53.
- [36] Nieschlag E, Cuppers HJ, Wickings EJ. Influence of sex, testicular development and liver function on the bioavailability of oral testosterone. *Eur J Clin Invest* 1977;7:145–7.
- [37] Kley HK. Plasma-estrogens and liver cirrhosis. *Z Gastroenterol* 1979;17:406–12.
- [38] Thole Z, Manso G, Salgueiro E, Revuelta P, Hidalgo A. Hepatotoxicity induced by antiandrogens: a review of the literature. *Urol Int* 2044;3:289–295.
- [39] Glud C, Christoffersen P, Eriksen J, Wantzin P, Knudsen BB. No effect of long-term oral testosterone treatment on liver morphology in men with alcoholic cirrhosis. *Am J Gastroenterol* 1987;82:660–4.
- [40] Glud C. Testosterone and alcoholic cirrhosis. Epidemiologic, pathophysiologic and therapeutic studies in men. *Dan Med Bull* 1988;35:564–75.
- [41] Ma WL, Hsu CL, Wu MH, Wu CT, Wu CC, Lai JJ, et al. Androgen receptor is a new potential therapeutic target for the treatment of hepatocellular carcinoma. *Gastroenterology* 2008;135. 947–55.
- [42] Chiu CM, Yeh SH, Chen PJ, Kuo TJ, Chang CJ, Chen PJ, et al. Hepatitis B virus X protein enhances androgen receptor-responsive gene expression depending on androgen level. *Proc Natl Acad Sci U S A* 2007;104:2571–8.
- [43] Ma WL, Hsu CL, Yeh CC, Wu MH, Huang CK, Jeng LB, et al. Hepatic androgen receptor suppresses hepatocellular carcinoma metastasis through modulation of cell migration and anoikis. *Hepatology* 2012;56:176–85.
- [44] Pikarsky E, Porat RM, Stein I, Abramovitch R, Amit S, Kasem S, et al. NF-kappaB functions as a tumour promoter in inflammation-associated cancer. *Nature* 2004;431:461–6.
- [45] Tanaka M, Nakashima O, Wada Y, Kage M, Kojiro M. Pathomorphological study of Kupffer cells in hepatocellular carcinoma and hyperplastic nodular lesions in the liver. *Hepatology* 1996;24:807–12.
- [46] Seki S, Habu Y, Kawamura T, Takeda K, Dobashi H, Ohkawa T, et al. The liver as a crucial organ in the first line of host defense: the roles of Kupffer cells, natural killer (NK) cells and NK1.1 Ag+ T cells in T helper 1 immune responses. *Immunol Rev* 2000;174:35–46.
- [47] Bird GL, Sheron N, Goka AK, Alexander GJ, Williams RS. Increased plasma tumor necrosis factor in severe alcoholic hepatitis. *Ann Intern Med* 1990;112:917–20.
- [48] McClain CJ, Cohen DA. Increased tumor necrosis factor production by monocytes in alcoholic hepatitis. *Hepatology* 1989;9:349–51.
- [49] Mayeux PR. Pathobiology of lipopolysaccharide. *J Toxicol Environ Health* 1997;51:415–35.
- [50] Lu T, Lin WJ, Izumi K, Wang X, Xu D, Fang LY, et al. Targeting androgen receptor to suppress macrophage-induced EMT and benign prostatic hyperplasia (BPH) development. *Mol Endocrinol* 2012;26:1707–15.
- [51] Lai JJ, Lai KP, Zeng W, Chuang KH, Altuwaijri S, Chang C, et al. How does the androgen receptor in the innate and adaptive immune system defend the body?: lessons from conditional AR knockout mice. *Am J Pathol* 2012;181(5):1504–12.



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## Review article

# Incorporating behavioral research to examine the relationship between betel quid chewing and oral cancer in Taiwan

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## ABSTRACT

Cancer of the oral cavity is one of the most commonly diagnosed cancers and one of the leading causes of death among men in Taiwan. Extensive research findings have linked betel quid chewing with oral cancer and precancerous conditions. To date, no pharmacological or behavioral treatments exist for betel quid cessation. This paper discusses the potential benefits of applying behavioral research to better understand why betel quid chewers consume betel quid. Specifically, it discusses using behavioral research methods to examine betel quid chewing initiation, dependence, motivation, and withdrawal. Better understanding of these different aspects of betel quid chewing is likely to aid researchers in developing treatment programs.

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## 1. Introduction

Cancer of the oral cavity is one of the fastest growing cancers among men in Taiwan. Between 1979 and 2006, the age-adjusted incidence for oral cancer among men increased seven times, from 5.04 per 100,000 to 35.88 per 100,000, an increase that is greater than that of the other nine most common cancers among Taiwanese men [1]. In 2006, cancer of the oral cavity was the fourth most commonly diagnosed cancer and the fourth leading cause of cancer death among men in Taiwan.

The International Agency for Research on Cancer has identified tobacco smoking and betel quid (BQ) chewing, with or without tobacco, as Group 1 carcinogens to humans [2]. Both habits have been found to be significantly related to oral cancer among men in Taiwan. In this paper, we

review current research on BQ use in Taiwan and the evidence regarding the link between BQ chewing and smoking and oral cancer among Taiwanese men. We also discuss the need for cessation of BQ chewing and smoking as a form of primary cancer prevention and as reduction of risk for additional cancer following cancer diagnosis and treatment.

## 2. BQ chewing in Taiwan

It is estimated that some 600 million people use BQ worldwide [2]. BQ chewing is popular in many Asian countries. Three types of quid, *lao-hwa* quid, BQ, and stem quid, are typically found in Taiwan [2]. *Lao-hwa* quid and stem quid are prepared by adding, respectively, a piece of inflorescence of *Piper betle* L.

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with red lime paste or a piece of *Piper betle* L. stem to an unripe areca nut. A BQ is made by wrapping a split unripe areca nut with slaked lime paste with a piece of betel leaf. In contrast, the practice in many southeast Asian countries, tobacco is never added to BQ in Taiwan.

The 2005 Taiwan National Health Interview Survey [3] found that, among the 16,542 adult respondents (age 18–64 years), 9.2% reported that they were current BQ chewers, 4.5% indicated that they had quit in the last 6 months, 7.2% indicated that they had tried BQ but were not regular users, and 79.1% responded that they had never used BQ. Of the current chewers, 42% chewed BQ daily, 16% chewed BQ on 3–5 days/week, and 22% used BQ on 1–2 days/week, and 20% chewed <4 days/month. Most chewers were men. Although 17% of all men living in Taiwan reported chewing BQ, <1% of all women reported regular use of BQ. Several other studies have confirmed this gender disparity in BQ chewing among adults [4–6] and adolescents [7,8]. The National Health Interview Survey also found that the majority of chewers were aged 35–44 years, lived in rural and less urbanized areas, and reported a family income of lower than NT\$30,000. Furthermore, individuals who completed junior high school or less in education and those who worked in nonprofessional, blue-collar occupations were more likely to chew BQ [9].

Due to historical and cultural factors, BQ chewing is especially widespread among ethnic aborigines, who make up approximately 2% of the population in Taiwan [10]. A stratified random survey that examined 7144 individuals in over 50 aboriginal communities found that 53% of male and 38% of female aborigines reported current or former use of BQ [10]. Three other large-scale surveys that sampled both aborigines and nonaborigines have also confirmed a higher rate of BQ chewing among aborigines [5,9,11]. The prevalence was estimated to be as high as 61% for men and 79% for women in a study that surveyed BQ chewing in a single aboriginal community in southern Taiwan [12]. Thus, the gender disparity that is found in the Taiwanese general population does not exist and may even be reversed among the ethnic aboriginal population.

Cigarette smoking and alcohol drinking have been found to be significantly associated with BQ use [5,8–10]. For example, Wen et al [13] analyzed data collected in the 2001 National Health Interview Survey in Taiwan and found that 93% of BQ chewers were current cigarettes smokers. It was also estimated that 50–67% of betel quit chewers consumed alcohol regularly [9,10].

We identified three surveys that examined unaided BQ cessation [9,10,14]. One study, which classified chewers as successful quitters if they had not chewed any BQ in the last 12 months, reported quit rates of 8.2% in male aborigines and 6.7% in female aborigines [10]. Using a less stringent criterion in which successful quitters were defined as chewers who had not chewed any BQ in the last 6 months, Lai et al [14] found that 49% of the Chinese (i.e., nonaborigine) chewers in their study had quit successfully. Similarly, Yap et al [9] reported that 28.6% of male nonaborigine chewers and 15.7% of female nonaborigine chewers had quit chewing in the last 6 months. Aborigine chewers had less success in quitting, with 7.8% of male and 9.8% of female chewers stopping chewing for at least 6 months. No information on relapse was discussed in any of the three studies.

Several factors, including gender, alcohol use, and level of education, have been found to be predictive of BQ cessation. Regarding gender, while one study found that male chewers were significantly more likely than female chewers to quit [odds ratio (OR) = 4.22, 95% confidence interval (CI) = 3.74–4.77] [9], another study found that although men quit at a higher rate than women, the odds ratio only approached significance (OR = 1.25, 95% CI = 0.95–1.64) [10]. These studies also found that those who drank alcohol and chewers who completed less education were significantly less likely to quit [9,10,14]. Younger chewers were more successful than older chewers in quitting [9,10]. The findings on the association between smoking and BQ cessation were mixed. One study found that chewers who did not smoke were more successful in quitting than those who did [9], whereas another failed to find significant difference in quitting success between smoking and non-smoking chewers [10].

### 3. BQ chewing, cigarette smoking, and cancer

A high incidence of oral cancer has been observed in countries where tobacco was routinely added to BQ [2]. In Taiwan, where tobacco is never added to BQ, numerous studies have found that BQ chewing, by itself or in combination with smoking, is significantly associated with precancerous oral lesions, cancer, and cancer death.

#### 3.1. Oral submucous fibrosis and oral leukoplakia

Oral mucosal lesions such as oral submucous fibrosis (OSF) and oral leukoplakia (OL) are risk factors for and have been found to transform malignantly to oral cancer [15,16]. Two case-control studies have investigated the effects of BQ chewing and cigarette smoking on OSF and OL [17,18]. One study that examined 62 histologically diagnosed OSF patients and 62 matched controls found that BQ chewers were 4.51 times (95% CI = 1.20–16.94) more likely than nonchewers to have OSF [18]. Individuals with both smoking and chewing habits had even higher risk of OSF (OR = 8.68, 95% CI = 1.87–40.23). A significant dose response effect was found, with chewers who used between 10 and 29 quids/day having a 4.55-fold (95% CI = 1.16–17.84) increase in risk and chewers who used 30 or more quids/day having a 10.34-fold (95% CI = 2.39–44.73) increase in risk for developing OSF compared to matched controls.

In another study that included 219 patients with histologically confirmed OL or OSF and 876 community controls, the risk for developing OL was 22.3 (95% CI = 11.3–43.8) times higher for current chewers than never chewers and the risk for developing OSF was 40.7 times (95% CI = 16.0–103.7) higher for current chewers than never chewers [17]. Significant dose response effects were also found for the development of both OL and OSF. A synergistic effect of BQ chewing and cigarette smoking on OL and OSF was also reported. Compared to individuals who neither chew BQ nor smoke cigarettes, those with both habits had a 40-fold (95% CI = 16.3–99.2) increase in risk of OL and a 57.9-fold (95% CI = 16.0–209.6) increase in risk for OSF.

In addition to case-control studies, three community surveys employed dentists to conduct oral examinations to diagnose OSF and OL in order to investigate the effects of BQ chewing on precancerous oral lesions. Yang et al [19] screened over 2000 participants and found that BQ chewing increased men's risk for OL and OSF, respectively, by 6.57-fold (95% CI = 3.51–12.28) and 22.86-fold (95% CI = 7.28–71.73), and increased women's risk for OL and OSF, respectively, by 15.63 fold (95% CI = 8.31–29.39) and 13.03-fold (95% CI = 5.21–32.62). The results also showed a significant positive association between chewing frequency and risks of OL and OSF, as well as between years of chewing and risks of OL and OSF. That is, heavier chewers with longer durations had higher risks of OL and OSF than lighter chewers with shorter durations of chewing.

A second large-scale study surveyed 1075 adults in southern Taiwan. It found that quid chewers' risk of oral precancerous lesions (OL, OSF, and verrucous lesions) were 8.4 times (95% CI = 5.13–13.75) higher than nonchewers [20]. This study also found a significant synergistic effect of BQ chewing, cigarette smoking, and alcohol drinking on OL. Compared to individuals who did not use BQ, cigarettes, or alcohol, those with reported regular use of all three substances had a 15.12-fold (95% CI = 6.34–36.05) increase in risk of OL.

Given the high prevalence of BQ chewing among aborigines, their risks of oral lesions were specifically examined in a study that screened 312 individuals from an aboriginal community in southern Taiwan [12]. Results showed that the odds of developing either OSF or OL were 8.21-fold (95% CI = 1.80–37.46) higher among chewers than among nonchewers.

So far, five studies have found significant results confirming the etiological role of BQ chewing in the development of oral precancerous conditions. These oral lesions have often been found to be precursors of oral cancer [15,16]. Two case-control studies that examined the effect of BQ chewing on the malignant transformation from oral lesions to oral cancer yielded mixed results. In one study that examined a cohort of 435 individuals diagnosed with OL, BQ chewers were 4.59 times (95% CI = 1.25–16.85) more likely than nonchewers to experience a malignant transformation to oral cancer [21]. However, another study that examined 104 histologically confirmed OSF patients failed to find a significant effect of BQ chewing on the development of oral cancer among OSF patients [22].

### 3.2. Oral, esophageal, and pharyngeal cancer

Four case-control studies investigated the association between BQ chewing and oral, esophageal, and pharyngeal cancer. In a study that enrolled 107 cancer patients and 200 matched noncarcinoma patients, researchers found that chewers had a significantly higher risk than nonchewers of developing oral cancer (OR = 6.9, 95% CI = 3.1–15.2) [23]. Among BQ chewers, those who also smoked cigarettes saw their risk of oral cancer increased from 6.9 times to 89 times (95% CI = 10.0–790.7) higher than those who did not use either BQ or cigarettes.

Two studies reported a positive association between BQ chewing and squamous-cell carcinoma of the esophagus.

Both case-control investigations paired patients with histologically confirmed esophageal cancer with hospital-matched noncancer (control) patients. Wu et al [24] recruited 104 cases and 277 controls and found a significant dose-response effect of BQ chewing on esophageal cancer. Compared to non-chewers, patients who consumed  $\leq 495$  betel-years (about 20 quids/day for 20 years) had a 3.6-fold (95% CI = 1.3–10.1) increase in risk of esophageal cancer; those who consumed  $> 495$  betel-years had a 9.2-fold (95% CI = 1.8–46.7) increase in risk of esophageal cancer.

Lee et al [25] found, in a multi-site case-control study that included 513 esophageal cancer and 818 hospital-matched control patients, that BQ chewers had a 2.3-fold (95% CI = 1.4–3.7) increase in esophageal cancer than non-chewers. They also found a significant association between risk of esophageal cancer and initiation age of BQ chewing, years of chewing, and average amount of quid consumed/day. Furthermore, a synergistic effect of BQ chewing and cigarette smoking was found. Compared to individuals who neither smoked nor chewed, adding cigarette smoking to BQ chewing increased patients' risk of esophageal cancer from 2.3-fold to 8.8-fold (95% CI = 5.2–14.8).

In a study that examined 148 patients with histologically confirmed pharyngeal cancer, 128 patients with histologically confirmed laryngeal cancer, and 255 matched control patients, the results showed that BQ chewing was significantly associated with pharyngeal (OR = 6.9, 95% CI = 3.4–14.3) but not laryngeal cancer [26]. Compared to non-chewers, patients who chewed more than 20 quids/day (OR = 7.2, 95% CI = 3.6–14.8) were also found to incur a significantly higher risk of pharyngeal cancer than those who chewed less than 20 quids/day (OR = 2.5, 95% CI = 1.0–3.8). Adding cigarette smoking to BQ chewing significantly increased patients' risk of pharyngeal cancer from 6.9-fold to 19.0-fold (95% CI = 5.7–70.6). Chewers who swallowed the BQ juice had higher odds (OR = 8.7) of developing pharyngeal cancer than nonswallowers (OR = 6.2). The pharynx, which is located immediately posterior to the mouth, may be more likely than the larynx, which is located in the upper air way and is inferior to the pharynx, to come into direct contact with BQ. As such, the differential anatomical position of the two structures may explain why BQ chewing is significantly associated with pharyngeal but not laryngeal cancer.

In addition to the case-control studies, two cohort studies also found evidence that supported a significant association between BQ chewing and oral cancer. The first study screened 8356 male patients (191 patients were diagnosed with oral cancer) over 2 years at a Taichung hospital and found a significant effect of BQ chewing on the development of oral cancer (OR = 9.03, 95% CI = 3.22–37.34) [27]. The OR of oral cancer increased to 21.79 (95% CI = 11.08–42.85) for patients who chewed BQ and smoked cigarettes.

Finally, information from 177,271 men who participated in a medical screening program was included in a large cohort study that examined the relationship between BQ chewing and cancer mortality [28]. In this study, the majority (90%) of those who chewed BQ also reported cigarette smoking. Compared to men who did not chew BQ or smoke cigarettes, BQ chewers were found to have a significantly higher risk of oral cancer



death [hazard ratio (HR) = 12.52, 95% CI = 5.45–28.77]. BQ chewers were also found to have elevated risk for esophageal cancer death (HR = 5.64, 95% CI = 2.25–14.12), liver cancer death (HR = 2.27, 95% CI = 1.12–4.60), pancreatic cancer death (HR = 2.67, 95% CI = 1.23–5.78), larynx cancer death (HR = 6.24, 95% CI = 1.03–37.44), and lung cancer death (HR = 2.43, 95% CI = 1.73–3.41). Since 90% of the BQ chewers also smoked cigarettes, efforts were made to tease apart the unique contribution of BQ chewing to cancer mortality. Compared to smokers who did not chew BQ, smokers who chewed had a significantly higher hazard of oral cancer death (HR = 4.84, 95% CI = 2.68–8.72), esophageal cancer death (HR = 2.20, 95% CI = 1.12–4.02), liver cancer death (HR = 1.73, 95% CI = 1.35–2.23), and larynx cancer death (HR = 4.09, 95% CI = 1.33–12.55).

In summary, findings from case-control, survey, and cohort studies have revealed a significant effect of BQ chewing on the development of precancerous oral lesions, oral cancer, esophageal cancer, and pharyngeal cancer. These risks were found to be considerably compounded if BQ chewers also smoked cigarettes. Both chewing BQ and smoking have been found to be associated with oral, esophageal, liver, pancreatic, larynx, and lung cancer mortality.

#### 4. Behavioral research program on BQ chewing

Although extensive research findings have linked BQ chewing with oral cancer and precancerous conditions, to date, no pharmacological or behavioral treatments exist for BQ cessation. Previous studies that examined Taiwanese BQ chewers have found that male gender [3–6], membership in an aboriginal tribe [5,9,11], low socio-economic status [3,9], cigarette smoking [13], and alcohol drinking [9,10] are significantly associated with BQ use [3]. While these findings provide valuable information about who uses BQ (i.e., the characteristics of BQ users) in Taiwan, they offer relatively few insights about why these users chew BQ. Better understanding of the reasons why chewers consume BQ is likely to aid researchers in developing treatment programs.

In tobacco research, findings from behavioral studies have contributed significantly to the understanding of nicotine addiction as well as to the development of smoking cessation interventions. For example, several behavioral models of drug addiction have been proposed, which have enriched our understanding regarding smokers' motivation to smoke [29,30] as well as the importance of smoking-related cues in eliciting smoking behaviors [31,32]. Behavioral studies have also uncovered a positive association between nicotine withdrawal symptoms and cessation failure [33–36]. Furthermore, behavioral researchers have found evidence that many behavioral and cognitive constructs (e.g., smoking outcome expectancies, self-efficacy) predict smoking relapse [37,38]. An enormous trove of studies exists on each of these topics and a comprehensive review is beyond the scope of the present article. Thus, we briefly review several areas of behavioral research on smoking and discuss their potential application to BQ chewing.

Considering the contributions that behavioral research has added to the understanding of smoking and smoking

cessation, we propose that similar efforts should be initiated to define the behavioral aspects of BQ chewing. To better understand the different facets of BQ chewing, we suggest that behavioral researchers: (1) examine the factors relating BQ chewing initiation; (2) develop a self-report instrument that measures BQ dependence; (3) investigate chewers' motivation to chew BQ; and (4) document withdrawal symptoms experienced by chewers upon stopping BQ.

##### 4.1. Examination of factors for BQ chewing initiation

Earlier smoking research focused on individual differences in sensitivity to nicotine, personality, and psychopathology to explain differences in smoking initiation. For example, individuals who showed high sensitivity to their initial exposure to nicotine [39], those with neurotic personality [40], those who suffered from major depression or schizophrenia [41], and those who met criteria for conduct or oppositional defiant disorder [42–44] were at higher risk of becoming smokers. Other studies showed that differences in life experiences such as poor or abusive childhood [45,46], major negative life events [47], and acute or chronic life stressors [48,49] increased a person's risk of taking up smoking. Furthermore, energy-balance factors, such as weight control, have been found to influence adolescents, especially young women, in starting smoking [50,51].

While there are many cross-sectional studies that reported demographic differences between BQ chewers and non-chewers [3,5,10], relatively few studies (cross-sectional or longitudinal) have looked at the role that innate constitutional (e.g., sensitivity to arecoline), psychological, or life experiential factors play in BQ initiation. Understanding not only who but also how a person takes up BQ chewing, and how one progresses from experimental to dependent chewers is likely to prove useful in the future development of BQ cessation programs.

##### 4.2. A self-report instrument to measure BQ dependence

Previous studies have shown that nicotine dependence is a significant inverse predictor of success in long-term smoking cessation [52–54]. That is, smokers with a high level of nicotine dependence are significantly less likely than those with a low level of dependence to achieve and maintain smoking abstinence. Furthermore, nicotine dependence has also been found to be associated with psychiatric and other drug dependence disorders [41,55–57].

Nicotine dependence is typically measured using the Fagerström Test for Nicotine Dependence [58], a six-item self-report instrument that assesses various components of smoking behavior such as daily intake, difficulty in refraining from smoking, and time to first cigarette of the day. The items were derived based on smokers' dose (e.g., number of cigarettes smoked/day) and on behaviors often observed in dependent smokers (e.g., earlier smoking in the morning, difficulty refraining from smoking in places where smoking is prohibited). The Fagerström Test for Nicotine Dependence has been shown to be reliable and smokers' scores on the measure are significantly correlated with biochemical markers of nicotine use [58].

BQ chewing is an addictive behavior and chewers are likely to differ in their dependence on the BQ. A self-report instrument to assess BQ dependence will provide researchers with an easy to administer and valid mean to quantify chewers' dependence. With such an instrument, researchers can begin to examine questions regarding dependence (e.g., are all BQ chewers dependent?), consumption (e.g., do more dependent chewers chew more BQs/day?), and abstinence (e.g., do more dependent chewers have more difficulty quitting than their less dependent counterparts?). A self-report dependence scale will also allow researchers to examine the role that active quid ingredients (e.g., arecoline, arecaidine) plays in BQ use, and the complex ways in which these ingredients interact with behavioral and sensory factors in determining BQ consumption.

Two studies have shown that arecoline can be reliably detected, using high-performance liquid chromatography, in BQ chewers' saliva [59] and blood [60]. Wu et al [60] further demonstrated that the levels of plasma arecoline and plasma arecaidine found in BQ chewers' blood were positively correlated with the amount of BQ consumed, making them ideal biochemical markers of BQ use. Future behavioral research should consider employing plasma arecoline or arecaidine levels to validate self-report dependence measure or to verify self-report BQ use and cessation.

#### 4.3. Chewers' motivation to chew BQ

Smoking behaviors are often influenced by smoking outcome expectancies. It has been hypothesized that while smokers with high expectancies for positive outcome of smoking (e.g., positive mood enhancement, alleviation of negative mood) are likely to increase their cigarette consumption, those with high expectancies for negative smoking outcome (e.g., smoking is detrimental to one's health) are likely to reduce their cigarette intake. Several studies have found that positive smoking outcome expectancies significantly predict the occurrence of smoking lapse and relapse [61–63].

Many BQ chewers expect that chewing can improve their mood, heighten their alertness, quench their thirst, warm their body, curb their appetite, and increase their stamina [64]. They also know that chewing increases their risk of oral cancer [3]. Nevertheless, it is unclear whether or not these positive and negative outcome expectancies of chewing influence chewers' behaviors. To better understand chewers' motivation to use or to quit BQ, it is imperative for behavioral researchers to begin to examine these factors and their effects on chewing.

#### 4.4. BQ withdrawal symptoms

When smokers stop smoking, they experience nicotine withdrawal symptoms that may include a negative affect (e.g., depression), urge to smoke, irritability, anxiety, cognitive and attention deficits, sleep disturbance, and increased appetite. The nature of postcessation withdrawal symptoms may have important implications for smoking relapse. If postcessation withdrawal symptoms resolve quickly, abstinent smokers may be less motivated to smoke and may have better success in maintaining their abstinence. However, if postcessation

withdrawal symptoms are unrelenting, abstinent smokers may be more driven to smoke and may eventually resort to smoking for withdrawal relief. Numerous studies have found evidence that supports a strong link between postcessation withdrawal symptoms and smoking lapse or relapse. For example, Piasecki et al [33–36] identified several different trajectories of nicotine withdrawal and found that abstinent smokers who had prototypical withdrawal profile (transient course with quick resolution of symptoms) were significantly less likely to lapse than those who had the atypical profiles (extended course of unrelieved symptoms).

Specific nicotine withdrawal symptoms have also been found to be directly and indirectly associated with smoking and smoking relapse. At least one model of addiction has examined the effects of negative affect on smoking [29] and postcessation negative affect has been found to be one of the most potent predictor of relapse [65–69]. In addition to negative affect, smoking urge has also received a lot of attention in smoking cessation research. Several studies have found a positive link between urge and smoking relapse [70,71]. Finally, smokers often use smoking as a mean to control their body weight. Several studies have found evidence suggesting that smokers, especially women, who quit smoking resumed their cigarette consumption to curb increased appetite, a symptom of nicotine withdrawal [72–75].

Given this evidence regarding the effect of withdrawal symptoms on smoking relapse, we argue that it is important to study BQ withdrawal symptoms and to track changes in them across the postcessation period. Anecdotal evidence suggests that BQ chewers may experience withdrawal symptoms when they stop chewing. However, to date, no systematic efforts have been made to investigate what constitutes BQ withdrawal. Thus, we suggest that behavioral researchers to begin to identify the nature and frequency of all symptoms of withdrawal that BQ chewers experience when they abstain from chewing, and to track these overtime.

## 5. Summary

As many as 10% of people in Taiwan chew BQ. A typical chewer is male, middle-aged, living in a rural area, has a low level of education, and works in a blue-collar occupation. Among chewers, those who do not use alcohol and have a higher level of education are more successful in quitting. Despite a strong link between BQ chewing and oral cancer, relatively few efforts have been made to examine why chewers use BQ and the behavioral sequelae that are related to BQ addiction. We suggest several areas (dependence, drug use motivation, and withdrawal symptoms) in which researchers can begin to study the behavioral aspects of BQ chewing.

## REFERENCES

- [1] Taiwan Cancer Registry. Cancer incidence and mortality rates in Taiwan. Taipei: Taiwan Cancer Registry; 2006.
- [2] IARC. Betel-quid and areca-nut chewing and some areca-nut-derived nitrosamines. 2004. Lyon, France, World Health

- Organization. IARC monographs on the evaluation of carcinogenic risks to humans. Ref Type: Serial (Book, Monograph).
- [3] Department of Health. National health interview survey. Taipei: Department of Health; 2005.
- [4] Chen JW, Shaw JH. A study on betel quid chewing behavior among Kaohsiung residents aged 15 years and above. *J Oral Pathol Med* 1996;25:140–3.
- [5] Chen KT, Chen CJ, Fagot-Campagna A, Narayan KM. Tobacco, betel quid, alcohol, and illicit drug use among 13- to 35-year-olds in I-Lan, rural Taiwan: prevalence and risk factors. *Am J Public Health* 2001;91:1130–4.
- [6] Ko YC, Chiang TA, Chang SJ, Hsieh SF. Prevalence of betel quid chewing habit in Taiwan and related sociodemographic factors. *J Oral Pathol Med* 1992;21:261–4.
- [7] Wang SC, Tsai CC, Huang ST, Hong YJ. Betel nut chewing: the prevalence and the intergenerational effect of parental behavior on adolescent students. *J Adolescent Health* 2004;34:244–9.
- [8] Yang MS, Su IH, Wen JK, Ko YC. Prevalence and related risk factors of betel quid chewing by adolescent students in southern Taiwan. *J Oral Pathol Med* 1996;25:69–71.
- [9] Yap SF, Ho PS, Kuo HC, Yang YH. Comparing factors affecting commencement and cessation of betel quid chewing behavior in Taiwanese adults. *BMC Public Health* 2008;8:199.
- [10] Lin CF, Wang JD, Chen PH, Chang SJ, Yang YH, Ko YC. Predictors of betel quid chewing behavior and cessation patterns in Taiwan aborigines. *BMC Public Health* 2006;6:271.
- [11] Ho CS, Gee MJ, Tsai CC, Lo CI, Hwang MN. Factors related to betel chewing among junior high school students in Taiwan. *Community Dent Oral Epidemiol* 2000;28:150–4.
- [12] Yang YH, Lee HY, Tung S, Shieh TY. Epidemiological survey of oral submucous fibrosis and leukoplakia in aborigines of Taiwan. *J Oral Pathol Med* 2001;30:213–9.
- [13] Wen CP, Tsai SP, Cheng TY, Chen CJ, Levy DT, Yang HJ, et al. Uncovering the relation between betel quid chewing and cigarette smoking in Taiwan. *Tob Control* 2005;14:i16–22.
- [14] Lai CS, Shieh TY, Yang YH, Chong MY, Hung HC, Tsai CC. Factors associated with quitting areca (betel) quid chewing. *Community Dent Oral Epidemiol* 2006;34:467–74.
- [15] Gupta PC, Mehta FS, Daftary DK, Pindborg JJ, Bhonsle RB, Jalnawalla PN, et al. Incidence rates of oral cancer and natural history of oral precancerous lesions in a 10-year follow-up study of Indian villagers. *Community Dent Oral Epidemiol* 1980;8:283–333.
- [16] Murti PR, Bhonsle RB, Pindborg JJ, Daftary DK, Gupta PC, Mehta FS. Malignant transformation rate in oral submucous fibrosis over a 17-year period. *Community Dent Oral Epidemiol* 1985;13:340–1.
- [17] Lee CH, Ko YC, Huang HL, Chao YY, Tsai CC, Shieh TY, et al. The precancer risk of betel quid chewing, tobacco use and alcohol consumption in oral leukoplakia and oral submucous fibrosis in southern Taiwan. *Br J Cancer* 2003;88:366–72.
- [18] Yang YH, Lien YC, Ho PS, Chen CH, Chang JSF, Cheng TC, et al. The effects of chewing areca/betel quid with and without cigarette smoking on oral submucous fibrosis and oral mucosal lesions. *Oral Diseases* 2005;11:88–94.
- [19] Yang YH, Ho PS, Lu HM, Huang IY, Chen CH. Comparing dose–response measurements of oral habits on oral leukoplakia and oral submucous fibrosis from a community screening program. *J Oral Pathol Med* 2010;39:306–12.
- [20] Chung CH, Yang YH, Wang TY, Shieh TY, Warnakulasuriya S. Oral precancerous disorders associated with areca quid chewing, smoking, and alcohol drinking in southern Taiwan. *J Oral Pathol Med* 2005;34:460–6.
- [21] Shiu MN, Chen TH, Chang SH, Hahn LJ. Risk factors for leukoplakia and malignant transformation to oral carcinoma: a leukoplakia cohort in Taiwan. *Br J Cancer* 2000;82:1871–4.
- [22] Ho PS, Yang YH, Shieh TY, Huang IY, Chen YK, Lin KN, et al. Consumption of areca quid, cigarettes, and alcohol related to the comorbidity of oral submucous fibrosis and oral cancer. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod* 2007;104:647–52.
- [23] Ko YC, Huang YL, Lee CH, Chen MJ, Lin LM, Tsai CC. Betel quid chewing, cigarette smoking and alcohol consumption related to oral cancer in Taiwan. *J Oral Pathol Med* 1995;24:450–3.
- [24] Wu MT, Lee YC, Chen CJ, Yang PW, Lee CJ, Wu DC, et al. Risk of betel chewing for oesophageal cancer in Taiwan. *Br J Cancer* 2001;85:658–60.
- [25] Lee CH, Lee JM, Wu DC, Wang WM, Fang FM, Chiang FY, et al. Independent and combined effects of alcohol intake, tobacco smoking and betel quid chewing on the risk of esophageal cancer in Taiwan. *Int J Cancer* 2005;113:475–82.
- [26] Lee KW, Kuo WR, Tsai SM, Wu DC, Wang WM, Fang FM, et al. Different impact from betel quid, alcohol and cigarette: risk factors for pharyngeal and laryngeal cancer. *Int J Cancer* 2005;117:831–6.
- [27] Yen TT, Lin WD, Wang CP, Wang CC, Liu SA. The association of smoking, alcoholic consumption, betel quid chewing and oral cavity cancer: a cohort study. *Eur Arch Oto-Rhino-L* 2008;265:1403–7.
- [28] Wen CP, Tsai MK, Chung WS, Hsu HL, Chang YC, Chan HT, et al. Cancer risks from betel quid chewing beyond oral cancer: a multiple-site carcinogen when acting with smoking. *Cancer Causes Control* 2010;21:1427–35.
- [29] Baker TB, Piper ME, McCarthy DE, Majeskie MR, Fiore MC. Addiction motivation reformulated: an affective processing model of negative reinforcement. *Psychol Rev* 2004;111:33–51.
- [30] Stewart J, de Wit H, Eikelboom R. Role of unconditioned and conditioned drug effects in the self-administration of opiates and stimulants. *Psychol Rev* 1984;91:251–68.
- [31] Carter BL, Tiffany ST. Meta-analysis of cue-reactivity in addiction research. *Addiction* 1999;94:327–40.
- [32] Robinson TE, Berridge KC. The neural basis of drug craving: an incentive-sensitization theory of addiction. *Brain Res Rev* 2009;18:247–91.
- [33] Piasecki TM, Niaura R, Shadel WG, Abrams D, Goldstein M, Fiore MC, et al. Smoking withdrawal dynamics in unaided quitters. *J Abnorm Psychol* 2000;109:74–86.
- [34] Piasecki TM, Jorenby DE, Smith SS, Fiore MC, Baker TB. Smoking withdrawal dynamics: III. Correlates of withdrawal heterogeneity. *Exp Clin Psychopharmacol* 2003;11:276–85.
- [35] Piasecki TM, Jorenby DE, Smith SS, Fiore MC, Baker TB. Smoking withdrawal dynamics: I. Abstinence distress in lapsers and abstainers. *J Abnorm Psychol* 2003;112:3–13.
- [36] Piasecki TM, Jorenby DE, Smith SS, Fiore MC, Baker TB. Smoking withdrawal dynamics: II. Improved tests of withdrawal–relapse relations. *J Abnorm Psychol* 2003;112:14–27.
- [37] Marlatt G. Relapse prevention: theoretical rationale and overview of the model. In: Marlatt G, Gordon JR, editors. *Relapse prevention: maintenance strategies in the treatment of addictive behaviors*. New York: Guilford Press; 1985. p. 3–70.
- [38] Marlatt GA, Donovan DM. *Relapse prevention: maintenance strategies in the treatment of addictive behaviors*. 2nd ed. New York: Guilford Press; 2005.
- [39] Pomerleau OF, Collins AC, Shiffman S, Pomerleau CS. Why some people smoke and others do not: new perspectives. *J Consul Clin Psychol* 1993;61:723–31.
- [40] McCrae RR, Costa Jr PT, Bosse R. Anxiety, extraversion and smoking. *Br J Soc Clin Psychol* 1978;17:269–73.
- [41] Breslau N. Psychiatric comorbidity of smoking and nicotine dependence. *Behav Genet* 1995;25:95–101.

- [42] Ferdinand RF, Blüm M, Verhulst FC. Psychopathology in adolescence predicts substance use in young adulthood. *Addiction* 2001;96:861–70.
- [43] Kandel D, Chen K, Warner LA, Kessler RC, Grant B. Prevalence and demographic correlates of symptoms of last year dependence on alcohol, nicotine, marijuana and cocaine in the U.S. population. *Drug Alcohol Depen* 1997;44: 11–29.
- [44] Whalen CK, Jamner LD, Henker B, Delfino RJ. Smoking and moods in adolescents with depressive and aggressive dispositions: evidence from surveys and electronic diaries. *Health Psychol* 2001;20:99–111.
- [45] Felitti VJ, Anda RF, Nordenberg D, Williamson DF, Spitz AM, Edwards V, et al. Relationship of childhood abuse and household dysfunction to many of the leading causes of death in adults. The Adverse Childhood Experiences [ACE]. Study. *Am J Prev Med* 1998;14:245–58.
- [46] Anda RF, Croft JB, Felitti VJ, Nordenberg D, Giles WH, Williamson DF, et al. Adverse childhood experiences and smoking during adolescence and adulthood. *JAMA* 1999;282: 1652–8.
- [47] Siqueira L, Diab M, Bodian C, Rolnitzky L. Adolescents becoming smokers: the roles of stress and coping methods. *J Adolesc Health* 2000;27:399–408.
- [48] Koval JJ, Pederson LL, Mills CA, McGrady GA, Carvajal SC. Models of the relationship of stress, depression, and other psychosocial factors to smoking behavior: a comparison of a cohort of students in grades 6 and 8. *Prev Med* 2000;30: 463–77.
- [49] Koval JJ, Pederson LL. Stress-coping and other psychosocial risk factors: a model for smoking in grade 6 students. *Addict Behav* 1999;24:207–18.
- [50] French SA, Perry CL, Leon GR, Fulkerson JA. Weight concerns, dieting behavior, and smoking initiation among adolescents: a prospective study. *Am J Public Health* 1994;84:1818–20.
- [51] Tomeo CA, Field AE, Berkey CS, Colditz GA, Frazier AL. Weight concerns, weight control behaviors, and smoking initiation. *Pediatrics* 1999;104:918–24.
- [52] Augustson EM, Wanke KL, Rogers S, Bergen AW, Chatterjee N, Synder K, et al. Predictors of sustained smoking cessation: a prospective analysis of chronic smokers from the alpha-tocopherol beta-carotene cancer prevention study. *Am J Public Health* 2008;98:549–55.
- [53] Hyland A, Li Q, Bauer JE, Giovino GA, Steger C, Cummings KM. Predictors of cessation in a cohort of current and former smokers followed over 13 years. *Nicotine Tob Res* 2004;6:S363–9.
- [54] Hymowitz N, Cummings KM, Hyland A, Lynn WR, Pechacek TF, Hartwell TD. Predictors of smoking cessation in a cohort of adult smokers followed for five years. *Tob Control* 1997;6:S57–62.
- [55] Breslau N, Kilbey MM, Andreski P. Vulnerability to psychopathology in nicotine dependent smokers: an epidemiologic study of young adults. *Am J Psychiatry* 1993; 150:941–6.
- [56] Breslau N, Peterson E, Schultz L, Andreski P, Chilcoat H. Are smokers with alcohol disorders less likely to quit? *Am J Public Health* 1996;86:985–90.
- [57] Breslau N, Johnson EO. Predicting smoking cessation and major depression in nicotine-dependent smokers. *Am J Public Health* 2000;90:1122–7.
- [58] Heatherton TF, Kozlowski LT, Frecker RC, Fagerström KO. The Fagerström test for nicotine dependence: a revision of the Fagerström tolerance questionnaire. *Br J Addict* 1991;86: 1119–27.
- [59] Cox S, Piatkov I, Vickers ER, Ma G. High-performance liquid chromatographic determination of arecoline in human saliva. *J Chromatogr* 2004;1032:93–5.
- [60] Wu IC, Chen PH, Wang CJ, Wu DC, Tsai SM, Chao MR, et al. Quantification of blood betel quid alkaloids and urinary 8-hydroxydeoxyguanosine in humans and their association with betel chewing habits. *J Anal Toxicol* 2010; 34:325–31.
- [61] Copeland AL, Brandon TH, Quinn EP. The smoking consequences questionnaire-adult: measurement of smoking outcome expectancies of experienced smokers. *Psychol Assessment* 1995;7:484–94.
- [62] Gwaltney CJ, Shiffman S, Balabanis MH, Paty JA. Dynamic self-efficacy and outcome expectancies: prediction of smoking lapse and relapse. *J Abnorm Psych* 2005;114:661–75.
- [63] Wetter DW, Smith SS, Kenford SL, Jorenby DE, Fiore MC, Hurt RD, et al. Smoking outcome expectancies: factor structure, predictive validity, and discriminant validity. *J Abnorm Psych* 1994;103:801–11.
- [64] Chu NS. Neurological aspects of areca and betel chewing. *Addict Biol* 2002;7:111–4.
- [65] Borland R. Slip-ups and relapse in attempts to quit smoking. *Addict Behav* 1990;15:235–45.
- [66] Brandon TH, Tiffany ST, Obremski KM, Baker TB. Postcessation cigarette use: the process of relapse. *Addict Behav* 1990;15:105–14.
- [67] Shiffman S. Relapse following smoking cessation: a situational analysis. *J Consul Clin Psychol* 1982;50:71–86.
- [68] Shiffman S, Paty J, Gnys M, Kassel J, Hickcox M. First lapses to smoking: within-subjects analysis of real-time reports. *J Consul Clin Psychol* 1996;64:366–79.
- [69] Shiffman S, Hickcox M, Paty JA, Gnys M, Kassel JD, Richards TJ. Progression from a smoking lapse to relapse: prediction from abstinence violation effects, nicotine dependence, and lapse characteristics. *J Consul Clin Psychol* 1996;64:993–1002.
- [70] Killen JD, Fortmann SP. Craving is associated with smoking relapse: findings from three prospective studies. *Exp Clin Psychopharmacol* 1997;5:137–42.
- [71] Shiffman S, Perz WG, Gnys M, Kassel JD, Hickcox M. A day at a time: predicting smoking lapse from daily urge. *J Abnorm Psychol* 1997;106:104–16.
- [72] Gritz ER, Nielsen IR, Brooks LA. Smoking cessation and gender: the influence of physiological, psychological, and behavioral factors. *J Am Med Womens Assoc* 1996;51: 35–42.
- [73] Klesges RC, Brown K, Pascale RW, Murphy M, Williams E, Cigrang JA. Factors associated with participation, attrition and outcome in a smoking cessation program at the workplace. *Health Psychol* 1988;7:575–89.
- [74] Klesges RC, Meyers AW, Klesges LM, LaVasque ME. Smoking, body weight, and their effects on smoking behavior: a comprehensive review of the literature. *Psychol Bull* 1989; 106:204–30.
- [75] Klesges RC, Klesges LM. Cigarette smoking as a dieting strategy in a university population. *Int J Eat Disord* 1988;7: 413–9.



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## Review article

## Very long non-coding RNA and human disease

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## ABSTRACT

A role for non-coding RNAs (ncRNAs) in the development of disease has been well documented in the case of miRNAs. Recent studies have shown that long non-coding RNAs (lncRNAs), greater than 200 nt in length, are also implicated in various diseases. In this review, we focus on these lncRNAs, the very long non-coding RNAs (vlncRNAs), which are more than 5 kb long and for which detailed information is available. These studies have demonstrated that vlncRNAs have important biological functions, and that their aberrant expression may result in various cancers. Future investigations in this exciting field are needed to explore the role of vlncRNAs in pathogenesis and, in particular, to further understand their functional mechanisms.

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## 1. Introduction

Classically, proteins are recognized as having the main responsibility for biological function, with RNA merely a messenger that transfers protein-coding information from DNA [1,2]. This concept has changed in recent years, however—whereas only 2% of the genome encodes protein, more than 80% of the genome produces non-protein coding RNA transcripts [1–5], and these non-coding RNAs (ncRNAs) have important biological functions including gene regulation [6,7], imprinting [8–12], epigenetic

regulation [13,14], cell cycle control [15], regulation of transcription, translation, and splicing [7,16–20].

There are two major classes of ncRNAs, grouped according to size: small RNA, which includes microRNA (miRNA), PIWI-interacting RNA (piRNA), endogenous short interfering RNA (endo-siRNA), and other ncRNAs less than 200 nt; and long non-coding RNA (lncRNA), which is larger than 200 nt and is transcribed from intergenic, intragenic, or around protein-coding regions. miRNAs are involved in post-transcriptional regulation of mRNA through the RNA-induced silencing complex [16],

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whereas piRNAs and siRNAs maintain genomic integrity by suppressing transposable elements [17] or other unknown factors in cell nucleus [18]. The lncRNAs are involved in various levels of genome regulation and related fundamental epigenetic processes [19–25].

The importance of lncRNAs in gene regulation has become apparent in recent years [6,20–30], but the key sequences of lncRNA that determine regulatory function remain unknown. Thus, although the rules of translation via the genetic code are well understood and a mutation in a protein-coding gene that contributes to a given disease can be attributed to the resultant change in amino acid sequence [6,20–30], there is no equivalent code for lncRNA function. Genetic studies on lncRNA could help us identify their regulatory sequences and understand their mechanism of action more clearly. In this review, we focus on a specific group of lncRNAs, those more than 5000 nt long, which we call very long non-coding RNAs (vlncRNAs), and explore their roles in the development of disease.

## 2. vlncRNA annotation and relevant databases

High-throughput technologies such as Tiling Chip or Deep Sequencing data, combined with computational approaches, have identified long, abundantly expressed non-coding transcripts associated with various cancers [13,28,29,31–37]. Currently, ncRNAs are curated by a variety of public databases, such as lncRNadb (<http://lncrnadb.com/>), a database of eukaryotic lncRNAs validated by experimental data [38]; RNadb (<http://research.imb.uq.edu.au/rnadb/>), a lncRNA set conserved between human and mouse that was used in a high-throughput functional screen [39]; fRNadb (<http://www.ncrna.org/frnadb/index.html>), a database hosting a large collection of ncRNA sequence data from public non-coding databases [40]; NON-CODE (<http://www.noncode.org/NONCODERV3/>), a database of non-coding RNAs [41], together with annotation of potential

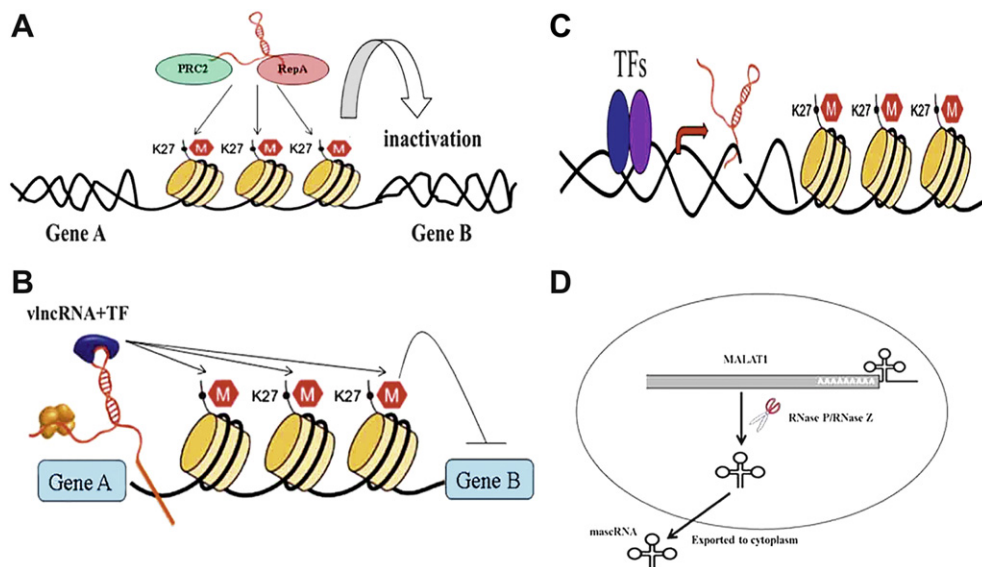
function, based on a coding–non-coding coexpression network [34]; and NRED (<http://jsm-research.imb.uq.edu.au/nred/>), a database of expression data of human and mouse lncRNAs with various gene expression profiles [31]. Moreover, the Encyclopedia of DNA Elements consortium is annotating lncRNAs using a combination of RNA-seq data, chromatin state maps, and computational approaches [42]. In this review, we survey the vlncRNAs, collected from various ncRNA databases by filtering for sequence length greater than 5000 nt.

## 3. Function of vlncRNAs

vlncRNAs originate from intronic, exonic, intergenic, intragenic, and promoter regions, and from 3'- and 5'-UTRs and enhancer sequences; they are sometimes bidirectional transcripts [30]. In particular, a large group of vlncRNAs, referred to as natural antisense transcripts, is antisense to known protein-coding genes [43,44]. In the following sections, we will discuss various vlncRNAs with respect to their functions in epigenetic regulation, transcriptional and post-transcriptional regulation, as well as in tumorigenesis (Fig. 1).

## 4. vlncRNAs control transcription by mediating changes in chromatin structure

The structure of chromatin determines the accessibility of DNA to polymerase II and transcription factors, and is integral to transcriptional control. Chromatin structure can be altered by specific post-translational modification, such as trimethylation of histone H3 lysine 4 (H3K4me3) at gene promoters, whereas H3K36me3 in transcribed regions is linked to gene activation, and H3K9me3, H3K27me3, and H4K20me3 are linked to repression [45] (Fig. 1A–C). There is substantial evidence for an important role of lncRNAs in these processes, and



**Fig. 1 – Mechanisms for regulation of epigenetics and gene expression by vlncRNAs. (A) vlncRNA regulates histone modification in cis and in trans. (B) Promoter-associated vlncRNA as an inhibitor of transcription. (C) Promoter-associated vlncRNA as an activator of transcription. (D) vlncRNA generation of sRNA regulatory transcript.**

approximately one-third of long intergenic non-coding RNA (lincRNA) is associated with chromatin-modifying complexes [13]. In addition, many intronic and intergenic ncRNAs have been found by different methods [14], suggesting that lncRNAs are key participants in controlling the chromatin structure. Similar to lncRNAs, vlncRNAs are also associated with chromatin-modifying complexes and can affect gene expression. The following examples describe vlncRNAs that mediate changes in chromatin structure and control transcription.

#### 4.1. *XIST* control of X-chromosome inactivation

On X-chromosome inactivation (XCI), one of the two X chromosomes in female mammals is inactivated to achieve comparable expression levels of X-chromosome genes in males and females. A number of vlncRNAs including *XIST* and *TSIX* participate in this process [46–49]. There are data suggesting that *XIST* recruits chromatin modeling complexes to silence the X chromosome (resulting in an inactive X chromosome, denoted Xi). Zhao et al [50] discovered a 1.6-kb ncRNA (*RepA*) transcribed from the 5' region of *XIST*, which binds polycomb repressive complex 2 (PRC2) to bring about silencing (Fig. 1A). In pre-XCI cells, *RepA* initially recruits PRC2 to the future Xi and inhibits the interaction of *TSIX* by binding PRC2. During initiation of the silencing process, *TSIX* is down-regulated on the future Xi, and *RepA* can engage PRC2 resulting in the activation of full-length *XIST* transcription (Fig. 1B and C). The up-regulated *XIST*, in turn, preferentially binds to PRC2 through its *RepA* sequence, resulting in the spread of *XIST* along Xi and the distribution of PRC2 and trimethylated histone H3K27 throughout the Xi [46,47,50]. Therefore, *RepA* and *XIST* are capable of recruiting PRC2 to establish the local chromatin modification required for the initiation and spread of XCI, resulting in the suppression of expression of genes on the Xi.

#### 4.2. vlncRNAs involved in imprinting

Just as vlncRNAs play important roles in X-inactivation, similar mechanisms have also been observed during genomic imprinting [8–10]. *AIR*, a 108-kb-long vlncRNA, is required for allele-specific silencing of the cis-linked *SLC22A3*, *SLC22A2*, and *IGF2R* genes [11]. *AIR* interacts with the *SLC22A3* promoter chromatin and the H3K9-specific histone methyltransferase G9a in placenta [10]. Depletion of G9a fails to silence *SLC22A3* and results in nonimprinted transcription. When truncated, *AIR* does not accumulate at the *SLC22A3* promoter, resulting in reduced G9a recruitment and biallelic transcription [10]. Similarly, the 90.5-kb vlncRNA *KCNQ1OT1* has been linked to the bidirectional silencing of about 10 paternally imprinted genes in the *KCNQ1OT1* domain, and the mechanism involves the interaction between *KCNQ1OT1* and both G9a and PRC2 in a lineage-specific manner [12].

#### 4.3. vlncRNAs involved in epigenetic silencing

The *INK4b/ARF/INK4a* locus in human cells contains three important tumor suppressors, whose expression is controlled in part by PRC1, PRC2, and histone methylation [51]. A vlncRNA called *ANRIL* (antisense non-coding RNA in the *INK4* locus) is transcribed in an antisense direction to the protein-coding genes in this locus, and is up-regulated in some

cancer tissues [52]. *ANRIL* plays an important role in controlling the epigenetic state of the *INK4b/ARF/INK4a* locus through interactions with subunits of PRC1 and PRC2. In brief, *ANRIL* binds to and recruits PRC1 and PRC2, resulting in trimethylation of H3K27 and repression of the *INK4b/ARF/INK4a* locus, facilitating oncogenesis [52] (Fig. 1A).

### 5. vlncRNA regulates splicing

The vlncRNA *MALAT1* (metastasis-associated in lung adenocarcinoma transcript 1) was identified in an attempt to characterize transcripts associated with early-stage non-small cell lung cancer (NSCLC) [53]. Two recent studies found that *MALAT1* regulates alternative splicing through its interaction with the serine/arginine-rich (SR) family of nuclear phosphoproteins involved in the splicing machinery [7,54], and *MALAT1* has been suggested to serve as a fine-tuning mechanism to modulate the activity of SR proteins.

*MALAT1* is an abundant vlncRNA transcribed from chromosome 11q13 and primarily localized in nuclear speckles. It modulates the distribution of pre-mRNA splicing factors to nuclear speckles and particularly affects the phosphorylation state of SR proteins [54]. In *MALAT1*-depleted cells, levels of mislocalized and unphosphorylated SR proteins increase, resulting in a higher number of exon inclusion events [7]. Therefore, *MALAT1* contributes to a broad post-transcriptional gene-regulatory mechanism by coordinating a specific mRNA patterning in distinct cell types.

### 6. vlncRNA is involved in a wide variety of other biological processes

Apart from their roles in nuclear processes discussed above, vlncRNAs have also been implicated in the regulation of a number of other biological processes. For example, some vlncRNAs can act as precursors for small RNAs, either by an RNase III-like cleavage from the sense and antisense duplexes, such as *XIST/TSIX*, or by a tRNA-like 3'-end processing of *MALAT1* [43,44] (Fig. 1D). Another example is the vlncRNA *DLEU2* (deleted in lymphocytic leukemia 2), which is frequently deleted in malignancy and functions as a critical host gene of the cell cycle inhibitory miR-15/16 [55]. Finally, *P15AS* (*P15*-antisense), which overlaps a protein-coding gene but is transcribed in the opposite direction, facilitates cancer progression by silencing its parental gene in cis and in trans through the formation of a type of heterochromatin [52,56].

### 7. vlncRNAs and diseases

The data gathered to date strongly implicate vlncRNAs in the basal regulation of protein-coding genes, including those central to normal development and oncogenesis, at both the transcriptional and post-transcriptional levels, and an increasing number have been functionally validated as affecting different cellular and developmental pathways [57,58]. It is not surprising, therefore, that the dysregulation of vlncRNAs appears to be a primary feature of many complex human diseases, including cancer.

Here, we describe some of the better characterized vlncRNAs that have been associated with cancer biology (Table 1).

### 7.1. Involvement in various cancers

XIST expression levels are correlated with outcome in some cancers [59,60], such as the therapeutic response in ovarian cancer [61], and it is frequently implicated in breast cancer [62–64]. Notably, however, because XIST is not expressed in males, such correlations are only of value in samples obtained from females [65].

### 7.2. Involvement in metastasis

The MALAT1 gene was associated with high metastatic potential and poor patient prognosis during a comparative screen of NSCLC patients with and without metastatic tumors [53]. Another report indicated that up-regulated MALAT1 contributes to bladder cancer cell migration by inducing epithelial–mesenchymal transition-associated genes [66].

### 7.3. Cancer type-specific vlncRNA expression

Many studies describe vlncRNA expression in various cancers, but a few vlncRNAs are specifically expressed in a particular

type of cancer. For example, the vlncRNAs PCA3 (prostate cancer antigen 3), PCGEM1 (prostate-specific transcript 1), and PCNCR1 (prostate cancer non-coding RNA 1) are associated with prostate cancer [67–70]. Indeed, PCA3 is a very sensitive and specific molecular marker for the diagnosis of prostate cancer [71]. Likewise, expression of PCGEM1, a prostate-specific gene with a function in the regulation of apoptosis, is associated with high-risk prostate cancer patients [70]. The vlncRNA PCNCR1 is also involved in prostate cancer progression [68].

## 8. Perspectives

As in the case of lncRNAs, vlncRNAs do not have protein-coding capacity, but nevertheless have functions relating to the programming and regulation of the mammalian genome. There have been recent rapid advancements in the understanding of these functions, but several important problems remain to be solved. For example, vlncRNAs frequently exhibit sequence divergence but have conserved functions, perhaps indicating the importance of secondary structure. Moreover, although a number of vlncRNAs are associated with PRC2 complexes, they do not interact exclusively with these proteins. Therefore, it will

**Table 1 – Identification of tumor-associated vlncRNAs.**

vlncRNA	Molecular mechanism	Tumor	Reference
P15AS/ANRIL	Antisense, transcription regulation	Prostate cancer	[52,56]
BA318C17.1	Unknown	Colon cancer	[72]
GNAS-AS1	Unknown		[73]
His-1 RNA	Unknown		[74]
KCNQ1OT1	DNMT1 interaction and transcription gene silencing	Colon cancer	[8,12]
DLEU2	Primary miRNA, other	Chronic lymphocytic leukemia	[55,75]
LOC285194	Unknown	Osteosarcoma	[76]
MALAT1	RNA splicing, small RNA Production, protein interaction	Multiple cancers	[44,53,66,77]
MEG3	Unknown	Multiple cancers	[78,79]
NCRMS	Unknown		[80]
NTT sense/antisense	Unknown		[81,82]
p53 mRNA	RNA protein binding	Multiple cancers	[83]
p53int1	unknown		[84]
PCA3/DD3	Unknown	Prostate cancer	[67]
PCGEM1	Unknown	Prostate cancer	[70]
PCNCR1	Unknown	Prostate cancer	[68]
PR Antisense	Regulation of gene expression		[85]
PRINS	Unknown		[86,87]
SRA RNA	RNA–protein binding, transcription factor coactivator	Breast cancer	[88,89]
TSIX	Antisense of Xist		[62,90]
SNHG14/UBE3A-AS1	Unknown		[91,92]
UCA1	Unknown	Bladder cancer	[93,94]
XIST	X inactivation	Multiple cancers	[59,60,62,95]
ZNFx1-AS1	Unknown	Breast cancer	[96]



be of great interest to unravel the sequences and structural motifs in vlncRNAs that determine their function.

Another challenging unanswered question is how protein partners interact with vlncRNAs to bring about their specialized functions. vlncRNAs, similar to lncRNAs, may recruit and then guide their protein partners to the correct chromosomal destinations. Specific sequences within vlncRNAs could recognize particular chromatin regions via sequence complementarity, thereby bringing the associated proteins to the targeted region. For example, XIST recruits PRC2 to establish local chromatin modification on Xi [50]. In the case of vlncRNAs recruiting proteins at a distance [34,35] or in *trans*, the higher-order, tertiary structure of chromatin might help bring distant chromosomal regions together. Alternatively, vlncRNAs might induce allosteric structural modifications of their protein partners to either enhance or suppress their normal activities.

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## REFERENCES

- [1] Birney E, Stamatoyannopoulos JA, Dutta A, Guigo R, Gingeras TR, Margulies EH, et al. Identification and analysis of functional elements in 1% of the human genome by the ENCODE pilot project. *Nature* 2007;447:799–816.
- [2] Kapranov P, Cheng J, Dike S, Nix DA, Duttagupta R, Willingham AT, et al. RNA maps reveal new RNA classes and a possible function for pervasive transcription. *Science* 2007;316:1484–8.
- [3] Kapranov P, Willingham AT, Gingeras TR. Genome-wide transcription and the implications for genomic organization. *Nat Rev Genet* 2007;8:413–23.
- [4] Alexander RP, Fang G, Rozowsky J, Snyder M, Gerstein MB. Annotating non-coding regions of the genome. *Nat Rev Genet* 2010;11:559–71.
- [5] Cooper DN, Chen JM, Ball EV, Howells K, Mort M, Phillips AD, et al. Genes, mutations, and human inherited disease at the dawn of the age of personalized genomics. *Hum Mutat* 2010;31:631–55.
- [6] Wery M, Kwapisz M, Morillon A. Noncoding RNAs in gene regulation. *Wiley Interdiscip Rev Syst Biol Med* 2011;3:728–38.
- [7] Bernard D, Prasanth KV, Tripathi V, Colasse S, Nakamura T, Xuan Z, et al. A long nuclear-retained non-coding RNA regulates synaptogenesis by modulating gene expression. *EMBO J* 2010;29:3082–93.
- [8] Pandey RR, Mondal T, Mohammad F, Enroth S, Redrup L, Komorowski J, et al. Kcnq1ot1 antisense noncoding RNA mediates lineage-specific transcriptional silencing through chromatin-level regulation. *Mol Cell* 2008;32:232–46.
- [9] Terranova R, Yokobayashi S, Stadler MB, Otte AP, van Lohuizen M, Orkin SH, et al. Polycomb group proteins Ezh2 and Rnf2 direct genomic contraction and imprinted repression in early mouse embryos. *Dev Cell* 2008;15:668–79.
- [10] Nagano T, Mitchell JA, Sanz LA, Pauler FM, Ferguson-Smith AC, Feil R, et al. The Air noncoding RNA epigenetically silences transcription by targeting G9a to chromatin. *Science* 2008;322:1717–20.
- [11] Sleutels F, Zwart R, Barlow DP. The non-coding Air RNA is required for silencing autosomal imprinted genes. *Nature* 2002;415:810–3.
- [12] Kanduri C. Kcnq1ot1: a chromatin regulatory RNA. *Semin Cell Dev Biol* 2011;22:343–50.
- [13] Khalil AM, Guttman M, Huarte M, Garber M, Raj A, Rivea Morales D, et al. Many human large intergenic noncoding RNAs associate with chromatin-modifying complexes and affect gene expression. *Proc Natl Acad Sci U S A* 2009;106:11667–72.
- [14] Mondal T, Rasmussen M, Pandey GK, Isaksson A, Kanduri C. Characterization of the RNA content of chromatin. *Genome Res* 2010;20:899–907.
- [15] Hung T, Wang Y, Lin MF, Koegel AK, Kotake Y, Grant GD, et al. Extensive and coordinated transcription of noncoding RNAs within cell-cycle promoters. *Nat Genet* 2011;43:621–9.
- [16] Diederichs S, Haber DA. Dual role for argonautes in microRNA processing and posttranscriptional regulation of microRNA expression. *Cell* 2007;131:1097–108.
- [17] Lee KJ, Conley AB, Lunyak VV, Jordan IK. Do human transposable element small RNAs serve primarily as genome defenders or genome regulators? *Mob Genet Elements* 2012;2:19–25.
- [18] Ishizu H, Nagao A, Siomi H. Gatekeepers for Piwi–piRNA complexes to enter the nucleus. *Curr Opin Genet Dev* 2011;21:484–90.
- [19] Chen LL, Carmichael GG. Decoding the function of nuclear long non-coding RNAs. *Curr Opin Cell Biol* 2010;22:357–64.
- [20] Clark MB, Mattick JS. Long noncoding RNAs in cell biology. *Semin Cell Dev Biol* 2011;22:366–76.
- [21] Guttman M, Rinn JL. Modular regulatory principles of large non-coding RNAs. *Nature* 2012;482:339–46.
- [22] Orom UA, Shiekhattar R. Noncoding RNAs and enhancers: complications of a long-distance relationship. *Trends Genet* 2011;27:433–9.
- [23] Saxena A, Carninci P. Long non-coding RNA modifies chromatin: epigenetic silencing by long non-coding RNAs. *Bioessays* 2011;33:830–9.
- [24] Wapinski O, Chang HY. Long noncoding RNAs and human disease. *Trends Cell Biol* 2011;21:354–61.
- [25] Wilusz JE, Sunwoo H, Spector DL. Long noncoding RNAs: functional surprises from the RNA world. *Genes Dev* 2009;23:1494–504.
- [26] Esteller M. Non-coding RNAs in human disease. *Nat Rev Genet* 2011;12:861–74.
- [27] Wang XQ, Crutchley JL, Dostie J. Shaping the genome with non-coding RNAs. *Curr Genomics* 2011;12:307–21.
- [28] Perez DS, Hoage TR, Pritchett JR, Ducharme-Smith AL, Halling ML, Ganapathiraju SC, et al. Long, abundantly expressed non-coding transcripts are altered in cancer. *Hum Mol Genet* 2008;17:642–55.
- [29] Chu C, Qu K, Zhong FL, Artandi SE, Chang HY. Genomic maps of long noncoding RNA occupancy reveal principles of RNA–chromatin interactions. *Mol Cell* 2011;44:667–78.
- [30] Nie L, Wu HJ, Hsu JM, Chang SS, Labaff AM, Li CW, et al. Long non-coding RNAs: versatile master regulators of gene expression and crucial players in cancer. *Am J Transl Res* 2012;4:127–50.
- [31] Dinger ME, Pang KC, Mercer TR, Crowe ML, Grimmond SM, Mattick JS. NRED: a database of long noncoding RNA expression. *Nucleic Acids Res* 2009;37:D122–6.
- [32] Pang KC, Dinger ME, Mercer TR, Malquori L, Grimmond SM, Chen W, et al. Genome-wide identification of long noncoding RNAs in CD8+ T cells. *J Immunol* 2009;182:7738–48.
- [33] Zhao J, Ohsumi TK, Kung JT, Ogawa Y, Grau DJ, Sarma K, et al. Genome-wide identification of polycomb-associated RNAs by RIP-seq. *Mol Cell* 2010;40:939–53.

- [34] Liao Q, Liu C, Yuan X, Kang S, Miao R, Xiao H, et al. Large-scale prediction of long non-coding RNA functions in a coding-non-coding gene co-expression network. *Nucleic Acids Res* 2011;39:3864–78.
- [35] Michelhaugh SK, Lipovich L, Blythe J, Jia H, Kapatos G, Bannon MJ. Mining Affymetrix microarray data for long non-coding RNAs: altered expression in the nucleus accumbens of heroin abusers. *J Neurochem* 2011;116:459–66.
- [36] Khachane AN, Harrison PM. Mining mammalian transcript data for functional long non-coding RNAs. *PLoS One* 2010;5:e10316.
- [37] Bellucci M, Agostini F, Masin M, Tartaglia GG. Predicting protein associations with long noncoding RNAs. *Nat Methods* 2011;8:444–5.
- [38] Amaral PP, Clark MB, Gascoigne DK, Dinger ME, Mattick JS. lncRNAdb: a reference database for long noncoding RNAs. *Nucleic Acids Res* 2011;39:D146–51.
- [39] Pang KC, Stephen S, Dinger ME, Engstrom PG, Lenhard B, Mattick JS. RNAdb 2.0—an expanded database of mammalian non-coding RNAs. *Nucleic Acids Res* 2007;35:D178–82.
- [40] Kin T, Yamada K, Terai G, Okida H, Yoshinari Y, Ono Y, et al. fRNAdb: a platform for mining/annotating functional RNA candidates from non-coding RNA sequences. *Nucleic Acids Res* 2007;35:D145–8.
- [41] Liu C, Bai B, Skogerbo G, Cai L, Deng W, Zhang Y, et al. NONCODE: an integrated knowledge database of non-coding RNAs. *Nucleic Acids Res* 2005;33:D112–5.
- [42] Raney BJ, Cline MS, Rosenbloom KR, Dreszer TR, Learned K, Barber GP, et al. ENCODE whole-genome data in the UCSC genome browser (2011 update). *Nucleic Acids Res* 2011;39:D871–5.
- [43] Ogawa Y, Sun BK, Lee JT. Intersection of the RNA interference and X-inactivation pathways. *Science* 2008;320:1336–41.
- [44] Wilusz JE, Freier SM, Spector DL. 3' end processing of a long nuclear-retained noncoding RNA yields a tRNA-like cytoplasmic RNA. *Cell* 2008;135:919–32.
- [45] Barski A, Cuddapah S, Cui K, Roh TY, Schones DE, Wang Z, et al. High-resolution profiling of histone methylations in the human genome. *Cell* 2007;129:823–37.
- [46] Jeon Y, Sarma K, Lee JT. New and Xisting regulatory mechanisms of X chromosome inactivation. *Curr Opin Genet Dev* 2012;22:62–71.
- [47] Gribnau J, Grootegoed JA. Origin and evolution of X chromosome inactivation. *Curr Opin Cell Biol* 2012;24:397–404.
- [48] Lee JT. Gracefully ageing at 50, X-chromosome inactivation becomes a paradigm for RNA and chromatin control. *Nat Rev Mol Cell Biol* 2011;12:815–26.
- [49] Morey C, Avner P. The demoiselle of X-inactivation: 50 years old and as trendy and mesmerising as ever. *PLoS Genet* 2011;7:e1002212.
- [50] Zhao J, Sun BK, Erwin JA, Song JJ, Lee JT. Polycomb proteins targeted by a short repeat RNA to the mouse X chromosome. *Science* 2008;322:750–6.
- [51] Bracken AP, Kleine-Kohlbrecher D, Dietrich N, Pasini D, Gargiulo G, Beekman C, et al. The Polycomb group proteins bind throughout the INK4A-ARF locus and are disassociated in senescent cells. *Genes Dev* 2007;21:525–30.
- [52] Yap KL, Li S, Munoz-Cabello AM, Raguz S, Zeng L, Mujtaba S, et al. Molecular interplay of the noncoding RNA ANRIL and methylated histone H3 lysine 27 by polycomb CBX7 in transcriptional silencing of INK4a. *Mol Cell* 2010;38:662–74.
- [53] Ji P, Diederichs S, Wang W, Boing S, Metzger R, Schneider PM, et al. MALAT-1, a novel noncoding RNA, and thymosin beta4 predict metastasis and survival in early-stage non-small cell lung cancer. *Oncogene* 2003;22:8031–41.
- [54] Tripathi V, Ellis JD, Shen Z, Song DY, Pan Q, Watt AT, et al. The nuclear-retained noncoding RNA MALAT1 regulates alternative splicing by modulating SR splicing factor phosphorylation. *Mol Cell* 2010;39:925–38.
- [55] Lerner M, Harada M, Loven J, Castro J, Davis Z, Oscier D, et al. DLEU2, frequently deleted in malignancy, functions as a critical host gene of the cell cycle inhibitory microRNAs miR-15a and miR-16-1. *Exp Cell Res* 2009;315:2941–52.
- [56] Yu W, Gius D, Onyango P, Muldoon-Jacobs K, Karp J, Feinberg AP, et al. Epigenetic silencing of tumour suppressor gene p15 by its antisense RNA. *Nature* 2008;451:202–6.
- [57] Mitra SA, Mitra AP, Triche TJ. A central role for long non-coding RNA in cancer. *Front Genet* 2012;3:17.
- [58] Spizzo R, Almeida MI, Colombatti A, Calin GA. Long non-coding RNAs and cancer: a new frontier of translational research? *Oncogene* 2012;31:4577–87.
- [59] Weakley SM, Wang H, Yao Q, Chen C. Expression and function of a large non-coding RNA gene XIST in human cancer. *World J Surg* 2011;35:1751–6.
- [60] Agrelo R, Wutz A. Context of change—X inactivation and disease. *EMBO Mol Med* 2010;2:6–15.
- [61] Huang KC, Rao PH, Lau CC, Heard E, Ng SK, Brown C, et al. Relationship of XIST expression and responses of ovarian cancer to chemotherapy. *Mol Cancer Ther* 2002;1:769–76.
- [62] Ganesan S, Silver DP, Greenberg RA, Avni D, Drapkin R, Miron A, et al. BRCA1 supports XIST RNA concentration on the inactive X chromosome. *Cell* 2002;111:393–405.
- [63] Vincent-Salomon A, Ganem-Elbaz C, Manie E, Raynal V, Sastre-Garau X, Stoppa-Lyonnet D, et al. X inactive-specific transcript RNA coating and genetic instability of the X chromosome in BRCA1 breast tumors. *Cancer Res* 2007;67:5134–40.
- [64] Sirchia SM, Ramoscelli L, Grati FR, Barbera F, Coradini D, Rossella F, et al. Loss of the inactive X chromosome and replication of the active X in BRCA1-defective and wild-type breast cancer cells. *Cancer Res* 2005;65:2139–46.
- [65] Carrel L, Willard HF. X-inactivation profile reveals extensive variability in X-linked gene expression in females. *Nature* 2005;434:400–4.
- [66] Ying L, Chen Q, Wang Y, Zhou Z, Huang Y, Qiu F. Upregulated MALAT-1 contributes to bladder cancer cell migration by inducing epithelial-to-mesenchymal transition. *Mol Biosyst* 2012;8:2289–94.
- [67] Lee GL, Dobi A, Srivastava S. Prostate cancer: diagnostic performance of the PCA3 urine test. *Nat Rev Urol* 2011;8:123–4.
- [68] Chung S, Nakagawa H, Uemura M, Piao L, Ashikawa K, Hosono N, et al. Association of a novel long non-coding RNA in 8q24 with prostate cancer susceptibility. *Cancer Sci* 2011;102:245–52.
- [69] Bussemakers MJ, van Bokhoven A, Verhaegh GW, Smit FP, Karthaus HF, Schalken JA, et al. DD3: a new prostate-specific gene, highly overexpressed in prostate cancer. *Cancer Res* 1999;59:5975–9.
- [70] Srikantan V, Zou Z, Petrovics G, Xu L, Augustus M, Davis L, et al. PCGEM1, a prostate-specific gene, is overexpressed in prostate cancer. *Proc Natl Acad Sci U S A* 2000;97:12216–21.
- [71] de Kok JB, Verhaegh GW, Roelofs RW, Hessels D, Kiemeny LA, Aalders TW, et al. DD3(PCA3), a very sensitive and specific marker to detect prostate tumors. *Cancer Res* 2002;62:2695–8.
- [72] Davison EJ, Tarpey PS, Fiegler H, Tomlinson IP, Carter NP. Deletion at chromosome band 20p12.1 in colorectal cancer revealed by high resolution array comparative genomic hybridization. *Genes Chromosomes Cancer* 2005;44:384–91.
- [73] Hayward BE, Bonthron DT. An imprinted antisense transcript at the human GNAS1 locus. *Hum Mol Genet* 2000;9:835–41.
- [74] Askew DS, Bartholomew C, Buchberg AM, Valentine MB, Jenkins NA, Copeland NG, et al. His-1 and His-2: identification and chromosomal mapping of two commonly

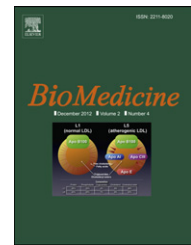
- rearranged sites of viral integration in a myeloid leukemia. *Oncogene* 1991;6:2041–7.
- [75] Migliazza A, Bosch F, Komatsu H, Cayanis E, Martinotti S, Toniato E, et al. Nucleotide sequence, transcription map, and mutation analysis of the 13q14 chromosomal region deleted in B-cell chronic lymphocytic leukemia. *Blood* 2001;97:2098–104.
- [76] Pasic I, Shlien A, Durbin AD, Stavropoulos DJ, Baskin B, Ray PN, et al. Recurrent focal copy-number changes and loss of heterozygosity implicate two noncoding RNAs and one tumor suppressor gene at chromosome 3q13.31 in osteosarcoma. *Cancer Res* 2010;70:160–71.
- [77] Li L, Feng T, Lian Y, Zhang G, Garen A, Song X. Role of human noncoding RNAs in the control of tumorigenesis. *Proc Natl Acad Sci U S A* 2009;106:12956–61.
- [78] Benetatos L, Vartholomatos G, Hatzimichael E. MEG3 imprinted gene contribution in tumorigenesis. *Int J Cancer* 2011;129:773–9.
- [79] Zhang X, Rice K, Wang Y, Chen W, Zhong Y, Nakayama Y, et al. Maternally expressed gene 3 (MEG3) noncoding ribonucleic acid: isoform structure, expression, and functions. *Endocrinology* 2010;151:939–47.
- [80] Chan AS, Thorner PS, Squire JA, Zielenska M. Identification of a novel gene NCRMS on chromosome 12q21 with differential expression between rhabdomyosarcoma subtypes. *Oncogene* 2002;21:3029–37.
- [81] Delgado Andre N, De Lucca FL. Non-coding transcript in T cells (NTT): antisense transcript activates PKR and NF-kappaB in human lymphocytes. *Blood Cells Mol Dis* 2008;40:227–32.
- [82] Liu AY, Torchia BS, Migeon BR, Siliciano RF. The human NTT gene: identification of a novel 17-kb noncoding nuclear RNA expressed in activated CD4+ T cells. *Genomics* 1997;39:171–84.
- [83] Candeias MM, Malbert-Colas L, Powell DJ, Daskalogianni C, Maslon MM, Naski N, et al. P53 mRNA controls p53 activity by managing Mdm2 functions. *Nat Cell Biol* 2008;10:1098–105.
- [84] Reisman D, Balint E, Loging WT, Rotter V, Almon E. A novel transcript encoded within the 10-kb first intron of the human p53 tumor suppressor gene (D17S2179E) is induced during differentiation of myeloid leukemia cells. *Genomics* 1996;38:364–70.
- [85] Chu Y, Yue X, Younger ST, Janowski BA, Corey DR. Involvement of argonaute proteins in gene silencing and activation by RNAs complementary to a non-coding transcript at the progesterone receptor promoter. *Nucleic Acids Res* 2010;38:7736–48.
- [86] Sonkoly E, Bata-Csorgo Z, Pivarcsi A, Polyanka H, Kenderessy-Szabo A, Molnar G, et al. Identification and characterization of a novel, psoriasis susceptibility-related noncoding RNA gene, PRINS. *J Biol Chem* 2005;280:24159–67.
- [87] Szegedi K, Sonkoly E, Nagy N, Nemeth IB, Bata-Csorgo Z, Kemeny L, et al. The anti-apoptotic protein G1P3 is overexpressed in psoriasis and regulated by the non-coding RNA, PRINS. *Exp Dermatol* 2010;19:269–78.
- [88] Colley SM, Leedman PJ. SRA and its binding partners: an expanding role for RNA-binding coregulators in nuclear receptor-mediated gene regulation. *Crit Rev Biochem Mol Biol* 2009;44:25–33.
- [89] Foulds CE, Tsimelzon A, Long W, Le A, Tsai SY, Tsai MJ, et al. Research resource: expression profiling reveals unexpected targets and functions of the human steroid receptor RNA activator (SRA) gene. *Mol Endocrinol* 2010;24:1090–105.
- [90] Lee JT, Davidow LS, Warshawsky D. Tsix, a gene antisense to Xist at the X-inactivation centre. *Nat Genet* 1999;21:400–4.
- [91] Mishra A, Godavarthi SK, Jana NR. UBE3A/E6-AP regulates cell proliferation by promoting proteasomal degradation of p27. *Neurobiol Dis* 2009;36:26–34.
- [92] Numata K, Kohama C, Abe K, Kiyosawa H. Highly parallel SNP genotyping reveals high-resolution landscape of mono-allelic Ube3a expression associated with locus-wide antisense transcription. *Nucleic Acids Res* 2011;39:2649–57.
- [93] Wang F, Li X, Xie X, Zhao L, Chen W. UCA1, a non-protein-coding RNA up-regulated in bladder carcinoma and embryo, influencing cell growth and promoting invasion. *FEBS Lett* 2008;582:1919–27.
- [94] Wang XS, Zhang Z, Wang HC, Cai JL, Xu QW, Li MQ, et al. Rapid identification of UCA1 as a very sensitive and specific unique marker for human bladder carcinoma. *Clin Cancer Res* 2006;12:4851–8.
- [95] Jeon Y, Lee JT. YY1 tethers Xist RNA to the inactive X nucleation center. *Cell* 2011;146:119–33.
- [96] Askarian-Amiri ME, Crawford J, French JD, Smart CE, Smith MA, Clark MB, et al. SNORD-host RNA Zfas1 is a regulator of mammary development and a potential marker for breast cancer. *RNA* 2011;17:878–91.



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## Clinical spotlight

# Unusual rectal submucosal tumor in melanosis coli

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## ARTICLE INFO

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A 60-year-old female patient was referred for polypectomy of multiple colonic flat adenomatous polyps. She suffered no abdominal pain or decreased stool caliber, and tested negative for stool occult blood. Her medical history revealed that she had been taking a laxative for 20 years as a treatment for constipation. The patient denied any systemic disease or taking any medication except for the laxative. Colonoscopy revealed multiple whitish flat polyps in melanosis coli throughout the entire colon, the largest of which was approximately 1.5 cm over the ascending colon (Fig. 1). A submucosal nodule, approximately 5 mm in size (Fig. 2), with multiple black dots on its surface was revealed when the scope was withdrawn to the rectum. Endoscopic ultrasound revealed a tumor originating in the second layer of the rectal wall, which was approximately 5 mm in size (Fig. 3).

A polypectomy of the rectal submucosal tumor was performed (Fig. 4). The mucosa was abnormally dark brown to black, grossly consistent with the presence of melanosis coli. Histopathologic evaluation with hematoxylin and eosin (H&E) staining (Fig. 5) showed that pigment-laden macrophages consistent with melanosis coli were present in the lamina propria and that tumor cells were arranged in

trabecular and glandular patterns, characterized by scanty cytoplasm, round nuclei, and fine chromatin, with rare mitotic figures. Immunohistochemical staining results were positive for markers of synaptophysin, chromogranin A (Fig. 6), and neuron-specific enolase. Rectal carcinoid in melanosis coli was identified.

The overall incidence of carcinoid tumors is difficult to determine because many are asymptomatic. They are discovered incidentally during routine colonoscopy and are usually less than 13 mm in size. Small rectal carcinoids are rarely malignant, and endoscopic resection is curative. The prevalence of rectal carcinoid is on the rise, particularly in the United States, where its age-adjusted incidence has increased by 800–1000% in the last 35 years [1]. The prevalence of rectal carcinoid in adults, as revealed by colonoscopy, is 0.05–0.07% [2].

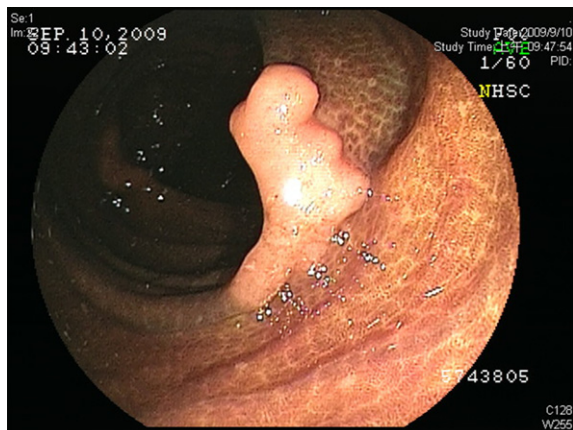
Melanosis coli refers to an abnormal brown or black pigmentation of colonic mucosa caused by the presence of lipofuscin produced by the breakdown of apoptotic colonic epithelial cells in macrophages within the lamina propria. Melanosis coli is usually related to chronic use of laxative agents. Endoscopic observation reveals blackening over the

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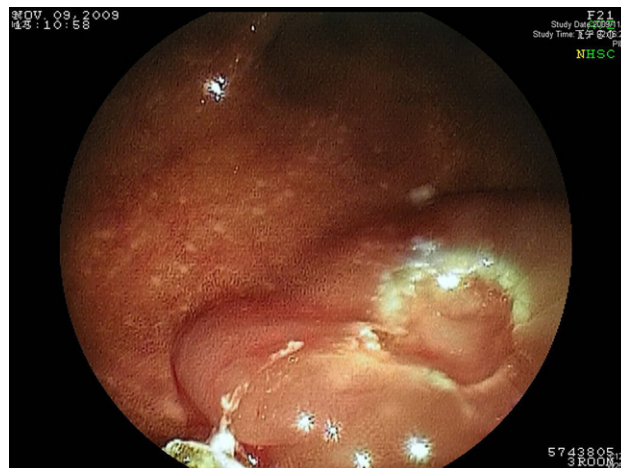
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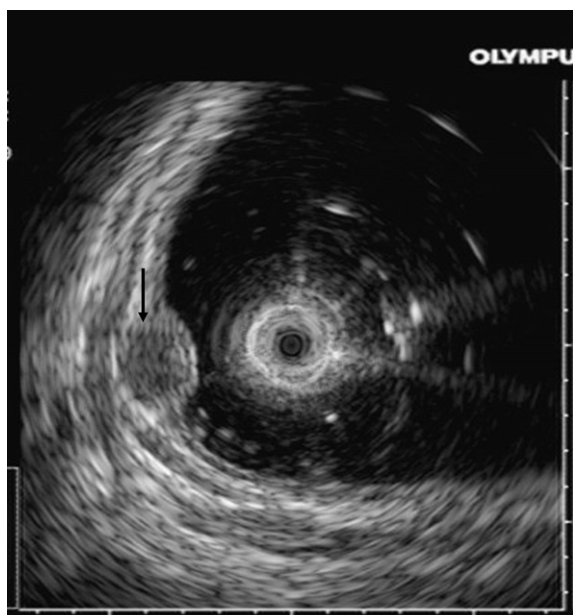
**Fig. 1 – Colonoscopic image showing whitish flat polyps in melanosis coli of ascending colon.**



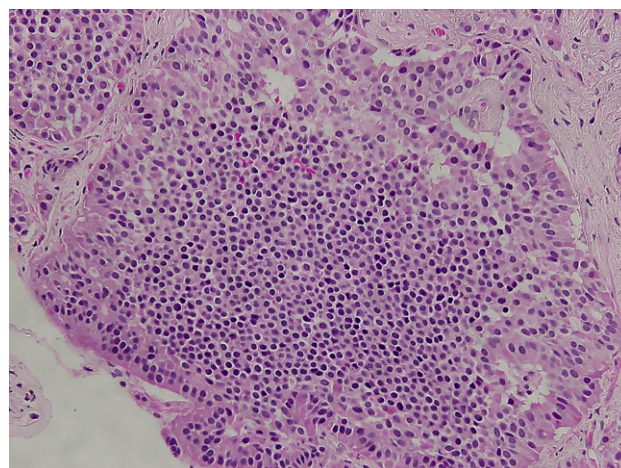
**Fig. 4 – Postpolypectomy of rectal submucosal tumor.**



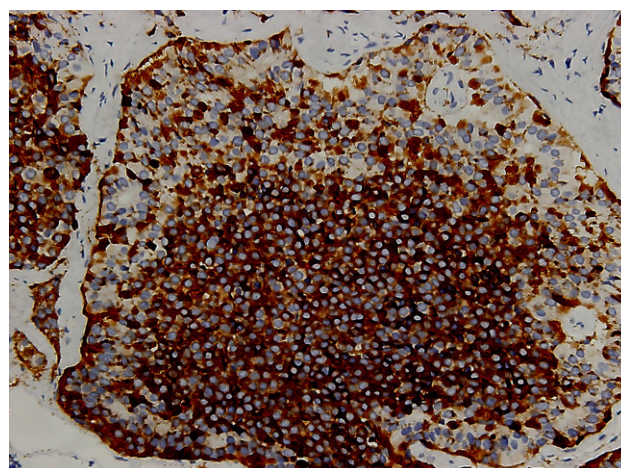
**Fig. 2 – Colonoscopic image revealing a submucosal nodule with multiple black dots in rectum.**



**Fig. 3 – Endoscopic ultrasound shows a tumor, approximately 5 mm in size, originating in the second layer of rectal wall.**



**Fig. 5 – H&E staining shows that melanosis coli is present in the lamina propria, with tumor cells arranged in trabecular and glandular patterns characterized by scanty cytoplasm, round nuclei, and fine chromatin. H&E = hematoxylin and eosin.**



**Fig. 6 – Synaptophysin staining shows positive findings.**

entire colonic mucosa [3]. Epithelial neoplasms in melanosis coli are easily observed because of their whitish color against a black background, as in the present case.

Rectal carcinoid usually presents a yellowish color change in the submucosal tumor, but this color change is often covered by black colonic mucosa. Submucosal tumor will be distal to mucosa, revealing multiple black dots in a diffuse black background under the endoscopic observation, as in this case. In conclusion, caution must be exercised in detecting submucosal tumor in melanosis coli.

## REFERENCES

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- [1] Modlin IM, Oberg K, Chung DC, Jensen RT, de Herder WW, Thakker RV, et al. Gastroenteropancreatic neuroendocrine tumours. *Lancet Oncol* 2008;9:61–72.
- [2] Modlin IM, Kidd M, Latich I, Zikusoka MN, Shapiro MD. Current status of gastrointestinal carcinoids. *Gastroenterology* 2005;128:1717–51.
- [3] Freeman HJ. "Melanosis" in the small and large intestine. *World J Gastroenterol* 2008;14:4296–9.

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### 8.3. Case Reports

These are short discussions of a case or case series with unique features not previously described that make an important teaching point or scientific observation. They may describe novel techniques, novel use of equipment, or new information on diseases of importance. Section headings should be: Abstract, Introduction, Case Report, Discussion, Acknowledgments (if applicable), Conflicts of Interest (if any), and References.

The Introduction should describe the purpose of the report, the significance of the disease and its specificity, and briefly review the relevant literature.

The Case Report should include the general data of the case, medical history, family history, chief complaint, present illness, clinical manifestation, methods of diagnosis and treatment, and outcome.

The Discussion should compare, analyze and discuss the similarities and differences between the reported case and similar previously reported cases. The importance or specificity of the case should be restated when discussing the differential diagnoses. Suggest the prognosis of the disease and possibility of prevention. Typical length: no more than 1500 words, 20–40 references.

### 8.4. Short Communications

These should be concise presentations of clinical or preliminary experimental results. Section headings should be: Abstract, Introduction, Methods, Results, Discussion, Acknowledgments (if applicable), Conflicts of Interest (if any), and References.

Typical length: no more than 1000 words, 20–40 references, with no more than four figures or tables. The Editors reserve the right to decide what constitutes a Short Communication.

### 8.5. Letters to the Editor

Letters are welcome in response to previously published articles, and may also include interesting cases that do not meet the requirement of being truly exceptional, as well as other communications of general interest. Letters should have a title and include appropriate references, and include the corresponding author's mailing and e-mail addresses. Letters are edited, sometimes extensively, to sharpen their focus. They may be sent for peer review at the discretion of the Editors. Letters are selected based on clarity, significance, and space. Typical length: no more than 600 words, 5–10 references; 1 table and/or 1 figure may be included.

### 8.6. Editorials

Editorials are invited articles or comments concerning a specific paper in the Journal or a topical issue in the field. While normally invited, unsolicited editorials may be submitted. Typical length: no more than 1500 words, 15–30 references.

## 9. Manuscript Preparation

Text should be typed double-spaced on one side of white A4 (297 × 210 mm) paper, with outer margins of 2.5 cm. A manuscript should include a title page, abstract, text, acknowledgments (if any), conflicts of interest statement (if any), references, and figures and tables as appropriate. Each section of the manuscript should begin on a new page. Pages should be numbered consecutively, beginning with the title page.

### 9.1. Title Page

The title page should contain the following information (in order, from the top to bottom of the page):

- category of paper
- article title
- names (spelled out in full)\* of all the authors, and the institutions with which they are affiliated; indicate all affiliations with a superscripted lowercase letter after the author's name and in front of the appropriate affiliation
- corresponding author details (name, e-mail, mailing address, telephone and fax numbers)

*\*The name of each author should be written with the family name last, e.g., Jing-Lin Chang. Authorship is restricted only to direct participants who have contributed significantly to the work.*

### 9.2. Abstract and Keywords

Abstracts should be no more than 300 words in length. Abstracts for Original Articles should be structured, with the section headings: Background/Introduction, Purpose(s)/Aim(s), Methods, Results, Conclusion. Abstracts for Case Reports are unstructured, but should include the significance and purpose of the case presentation, the diagnostic methods of the case, the key data, and brief comments and suggestions with regard to the case. Abstracts for Review Articles and Short Communications should also be unstructured. No abstract is required for Letters to the Editor and Editorials. For the article categories that require an abstract, 3–5 relevant keywords should also be provided in alphabetical order.

### 9.3. Main Text

The text for Original Articles should be organized into the following sections: Background/Introduction, Purpose(s)/Aim(s), Methods, Results and Discussion. Sections for Case Reports are: Introduction, Case Report, and Discussion. Each section should begin on a new page.

#### 9.3.1. Abbreviations

Where a term/definition will be continually referred to, it must be written in full when it first appears in the text, followed by the subsequent abbreviation in parentheses. Thereafter, the abbreviation may be used. An abbreviation should not be first defined in any section heading; if an abbreviation has previously been defined in the text, then the abbreviation may be used in a subsequent section heading. Restrict the number of abbreviations to those that are absolutely necessary.

#### 9.3.2. Units

Système International (SI) units must be used, with the exception of blood pressure values which are to be reported in mmHg. Please use the metric system for the expression of length, area, mass, and volume. Temperatures are to be given in degrees Celsius.

#### 9.3.3. Names of drugs, devices and other products

Use the Recommended International Non-proprietary Name for medicinal substances, unless the specific trade name of a drug is directly relevant to the discussion. For devices and other products, the generic term should be used, unless the specific trade name is directly relevant to the discussion. If the trade name is given, then the manufacturer name and the city, state and country location of the manufacturer must be provided the first time it is mentioned in the text, for example, "...SPSS version 11 was used (SPSS Inc., Chicago, IL, USA)."

#### 9.3.4. Statistical requirements

Statistical analysis is essential for all research papers except case reports. Use correct nomenclature of statistical methods (e.g., two sample t test, not unpaired t test). Descriptive statistics should follow the scales used in data description. Inferential statistics are important for interpreting results and should be described in detail.

All  $p$  values should be expressed to 2 digits to the right of the decimal point, unless  $p < 0.01$ , in which case the  $p$  value should be expressed to 3 digits to the right of the decimal point. The smallest  $p$  value that should be expressed is  $p < 0.001$ , since additional zeros do not convey useful information; the largest  $p$  value that should be expressed is  $p > 0.99$ .

#### 9.3.5. Personal communications and unpublished data

These sources cannot be included in the references list but may be described in the text. The author(s) must give the full name and highest academic degree of the person, the date of the communication, and indicate whether it was in oral or written (letter, fax, e-mail) form. A signed statement of permission should be included from each person identified as a source of information in a personal communication or as a source for unpublished data.

### **9.4. Acknowledgments and Conflicts of Interest Statement**

General acknowledgments for consultations, statistical analysis, etc., should be listed concisely at the end of the text, including the names of the individuals who were directly involved. Consent should be obtained from those individuals before their names are listed in this section. All financial and material support for the research and work from internal or external agencies, including commercial companies, should be clearly and completely identified. Ensure that any conflicts of interest (financial and/or non-financial) are explicitly declared.

### **9.5. Abbreviation list**

A term that appears more than three times in a paper should be abbreviated. Spell out the term on first mention, followed by the abbreviated form in parentheses. Thereafter, please use the abbreviated form. Supply a list of nonstandard abbreviations used in the paper at the end of the main text, in alphabetical order, giving each abbreviation followed by its spelled-out version.

### **9.6. References**

#### 9.6.1. In the main text, tables, figure legends

- References should be indicated by numbers in square brackets in line with the text, and numbered consecutively in order of appearance in the text.
- References cited in tables or figure legends should be included in sequence at the point where the table or figure is first mentioned in the main text.
- Do not cite uncompleted work or work that has not yet been accepted for publication (i.e., "unpublished observation", "personal communication") as references. Also see Section 9.3.5. above.
- Do not cite abstracts unless they are the only available reference to an important concept.

#### 9.6.2. In the references section

- References should be limited to those cited in the text and listed in numerical order, NOT alphabetical order.
- References should include, in order, author surnames and initials, article title, abbreviated journal name, year, volume and inclusive page numbers. The last names and initials of all the authors up to 6 should be included, but when authors number 7 or more, list the first 6 authors only followed by "et al". Abbreviations for journal names should conform to those used in MEDLINE.
- If citing a website, provide the author information, article title, website address and the date you accessed the information.

- Reference to an article that is in press must state the journal name and, if possible, the year and volume.

*Authors are responsible for the accuracy and completeness of their references and for correct text citation.*

Examples are given below.

*Standard journal article*

Chen Z, Fan M, Bian Z, Zhang Q, Zhu Q, Lu P. Immunolocalization of heat shock protein 70 during reparative dentinogenesis. *Chin J Dent Res* 2000;3:50–5.

*Journal supplement*

Kaplan NM. The endothelium as prognostic factor and therapeutic target: what criteria should we apply? *J Cardiovasc Pharmacol* 1998;32(Suppl 3):S78–80.

*Journal article not in English but with English abstract*  
Nakayama H, Ishikawa T, Yamashita S, Fukui I, Mutoh T, Hikichi K, et al. CSF leakage and anosmia in aneurysm clipping of anterior communicating artery by basal interhemispheric approach. *No Shinkei Geka* 2011;39:263–8. [In Japanese, English abstract]

*Book*

Bradley EL. Medical and surgical management. Philadelphia: Saunders; 1982, p. 72–95.

*Book chapter in book with editor and edition*

Greaves M, Culligan DJ. Blood and bone marrow. In: Underwood JCE, editor. General and systematic pathology. 4th ed. London: Churchill Livingstone; 2004, p. 615–72.

*Bulletin*

World Health Organization. World health report 2002: reducing risk, promoting healthy life. Geneva, Switzerland: World Health Organization; 2002.

*Company/manufacturer publication/pamphlet*

Eastman Kodak Company, Eastman Organic Chemicals. Catalog no. 49. Rochester, NY: Eastman Kodak; 1977, p. 2–3.

*Electronic publications*

Duchin JS. Can preparedness for biological terrorism save us from pertussis? *Arch Pediatr Adolesc Med* 2004;158:106–7. Available from: <http://archpedi.ama-assn.org/cgi/content/full/158/2/106>. Accessed June 5, 2004.

Smeeth L, Iliffe S. Community screening for visual impairment in the elderly. *Cochrane Database Syst Rev* 2002(2):CD001054. doi:10.1002/14651858.CD1001054.

*Items presented at a meeting but not yet published*

Durbin D, Kallan M, Elliott M, Arbogast K, Cornejo R,

Winston F. Risk of injury to restrained children from passenger air bags. Paper presented at: 46th Annual Meeting of the Association for the Advancement for Automotive Medicine; September 2002; Tempe, AZ.

Greenspan A, Eerdeken M, Mahmoud R. Is there an increased rate of cerebrovascular events among dementia patients? Poster presented at: 24th Congress of the Collegium Internationale Neuro-Psychopharmacologicum (CINP); June 20–24, 2004; Paris, France.

Khuri FR, Lee JJ, Lippman SM. Isotretinoin effects on head and neck cancer recurrence and second primary tumors. In: Proceedings from the American Society of Clinical Oncology; May 31–June 3, 2003; Chicago, IL. Abstract 359.

*Item presented at a meeting and published*

Cionni RJ. Color perception in patients with UV- or blue-light-filtering IOLs. In: Symposium on Cataract, IOL, and Refractive Surgery. San Diego, CA: American Society of Cataract and Refractive Surgery; 2004. Abstract 337.

*Material accepted for publication but not yet published*

Carrau RL, Khidr A, Crawley JA, Hillson EM, Davis JK, Pashos CL. The impact of laryngopharyngeal reflux on patient-reported quality of life. *Laryngoscope*. In press.

Ofri D. Incidental findings: Lessons from my patients in the art of medicine. Boston, MA: Beacon Press. In press.

*Theses and dissertations*

Undeman C. Fully automatic segmentation of MRI brain images using probabilistic diffusion and a watershed scale-space approach [master's thesis]. Stockholm, Sweden: NADA, Royal Institute of Technology; 2001.

Ayers AJ. Retention of resin restorations by means of enamel etching and by pins [dissertation]. Indianapolis: Indiana University; 1971.

*Website*

American Association of Oral and Maxillofacial Surgeons. Wisdom teeth. AAOMS Web site. [http://www.aaoms.org/wisdom\\_teeth.php](http://www.aaoms.org/wisdom_teeth.php). Published January 23, 2008. Updated March 9, 2009. Accessed November 15, 2009.

## 9.7. Tables

Tables should supplement, not duplicate, the text. They should have a concise table heading, be self-explanatory, and numbered consecutively in the order of their citation in the text. Information requiring explanatory footnotes should be denoted using superscripted lowercase letters in alphabetical order (a, b, c, etc.). Asterisks (\*, \*\*) are



used only to indicate the probability level of tests of significance. Abbreviations used in the table must be defined and placed after the footnotes. If you include a block of data or table from another source, whether published or unpublished, you must acknowledge the original source.

## 9.8. Figures

### 9.8.1. General guidelines

The number of figures should be restricted to the minimum necessary to support the textual material. They should have an informative figure legend and be numbered in the order of their citation in the text. All symbols and abbreviations should be defined in the legend. Patient identification should be obscured. All lettering should be done professionally and should be in proportion to the drawing, graph or photograph. Photomicrographs must include an internal scale marker, and the legend should state the type of specimen, original magnification and stain.

Figures must be submitted as separate picture files at the correct resolution (see Section 9.7.2. below). The files should be named according to the figure number, e.g., "Article1\_Fig1", "Article1\_Fig2".

### 9.8.2. Formats

Regardless of the application used, when your electronic artwork is finalized, please "save as" or convert the images to one of the following formats (note the resolution requirements for line drawings, halftones, and line/halftone combinations given below):

- EPS: Vector drawings. Embed the font or save the text as "graphics".
- TIFF: Color or grayscale photographs (halftones): always use a minimum of 300 dpi.
- TIFF: Bitmapped line drawings: use a minimum of 1000 dpi.
- TIFF: Combination of bitmapped line/half-tone (color or grayscale): a minimum of 600 dpi is required.
- DOC, XLS or PPT: If your electronic artwork is created in any of these Microsoft Office applications, please supply "as is".

Please do not:

- Supply files that are optimized for screen use (like GIF, BMP, PICT, WPG); the resolution is too low;
- Supply files that are too low in resolution;
- Submit graphics that are disproportionately large for the content.

A detailed guide on electronic artwork is available at <http://www.elsevier.com/artworkinstructions>.

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As a general rule, the receipt of a manuscript will be acknowledged within 1 week of submission, and authors will be provided with a manuscript reference number for future correspondence.

If such an acknowledgment is not received in a reasonable period of time, the author should contact the Editorial Office.

Submissions are reviewed by the Editorial Office to ensure that it contains all parts. The Editorial Office will not accept a submission if the author has not supplied all the material and documents as outlined in these author instructions.

Manuscripts are then forwarded to the Editor-in-Chief, who makes an initial assessment of it. If the manuscript does not appear to be of sufficient merit or is not appropriate for the Journal, then the manuscript will be rejected without review.

Manuscripts that appear meritorious and appropriate for the Journal are reviewed by at least two Editorial Board members or expert consultants assigned by the Editor-in-Chief. Authors will usually be notified within 6 weeks of whether the submitted article is accepted for publication, rejected, or subject to revision before acceptance. However, do note that delays are sometimes unavoidable.

## 11. Preparation for Publication

Once a manuscript has been accepted for publication, the authors should submit the final version of the manuscript in MS Word format, with all tables/figures as applicable, to the Editorial Office.

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- ☐ 3–5 relevant keywords in alphabetical order: required for Review Article, Original Article, Case Report, Short Communication (MeSH terms are recommended; see <http://www.ncbi.nlm.nih.gov/mesh?term>)
- ☐ Main text
- ☐ References in the correct format, cited in numerical order, and all references in the List are cited in the Text/Tables/Figures, and vice versa

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- ☐ Acknowledgments
- ☐ Conflicts of interest statement
- ☐ Table headings and tables, each on a new page
- ☐ Figure legends, on a new page
- ☐ Electronic picture files of all figures; resolution of 300 dpi for halftone images, 600 dpi for combination art (halftone + line art), and 1000 dpi for line art

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### Acknowledgments

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