

Review article

Autophagy and its link to type II diabetes mellitus

Jai-Sing Yang^{1,†}, Chi-Cheng Lu^{1,†}, Sheng-Chu Kuo², Yuan-Man Hsu³, Shih-Chang Tsai³, Shih-Yin Chen^{4,5},
Yng-Tay Chen^{4,5}, Ying-Ju Lin^{4,5}, Yu-Chuen Huang^{4,5}, Chao-Jung Chen^{4,5}, Wei-De Lin^{4,5}, Wen-Lin Liao^{4,5},
Wei-Yong Lin^{4,5}, Yu-Huei Liu^{4,5}, Jinn-Chyuan Sheu⁶, Fuu-Jen Tsai^{4,5,7,*}

¹Department of Medical Research, China Medical University Hospital, China Medical University, Taichung 404, Taiwan

²School of Pharmacy, China Medical University, Taichung 404, Taiwan

³Department of Biological Science and Technology, China Medical University, Taichung 404, Taiwan

⁴Genetics Center, Department of Medical Research, China Medical University Hospital, Taichung 404, Taiwan

⁵School of Chinese Medicine, China Medical University, Taichung 404, Taiwan

⁶Institute of Biomedical Sciences, National Sun Yat-sen University, Kaohsiung 804, Taiwan

⁷Department of Medical Genetics, China Medical University Hospital, China Medical University, Taichung 404, Taiwan

Received 11th of April 2017 Accepted 2nd of May 2017

© Author(s) 2017. This article is published with open access by China Medical University

Keywords:

Autophagy;
Type 2 diabetes mellitus
(T2DM);
Pancreatic β -cells;
Insulin resistance

ABSTRACT

Autophagy, a double-edged sword for cell survival, is the research object on 2016 Nobel Prize in Physiology or Medicine. Autophagy is a molecular mechanism for maintaining cellular physiology and promoting survival. Defects in autophagy lead to the etiology of many diseases, including diabetes mellitus (DM), cancer, neurodegeneration, infection disease and aging. DM is a metabolic and chronic disorder and has a higher prevalence in the world as well as in Taiwan. The character of diabetes mellitus is hyperglycemia resulting from defects in insulin secretion, insulin action, or both. Type 2 diabetes mellitus (T2DM) is characterized by insulin resistance and failure of producing insulin on pancreatic beta cells. In T2DM, autophagy is not only providing nutrients to maintain cellular energy during fasting, but also removes damaged organelles, lipids and miss-folded proteins. In addition, autophagy plays an important role in pancreatic beta cell dysfunction and insulin resistance. In this review, we summarize the roles of autophagy in T2DM.

1. Introduction

Professor Yoshinori Ohsumi, the 2016 laureate in Physiology or Medicine, discovered the mechanisms for autophagy [1-4]. This pathway plays a crucial role in physiological cellular homeostasis and human diseases [5]. Autophagy has been known to serve as a double-edged sword for promoting survival character and/or activating cell death (Fig. 1) [6-11]. In addition, autophagy, a catabolic process, degrades cellular components and damaged organelles [12, 13]. Recently, autophagic machinery is involved in the pathophysiology of type 2 diabetes mellitus (T2DM) disease, and it regulates normal function of pancreatic beta cells. On the other hand, enhanced autophagy acts as an important protective mechanism against to oxidative stress on insulin-target tissues such as liver, adipose tissue and skeletal muscle [14-19]. In this review, we outline the relationship among autophagy, pancreatic beta cells and T2DM. Furthermore, we highlight recent findings on the novel agents to specifically target autophagy in T2DM.

2. Programmed cell death (PCD)

PCD is an important physiological process during organ development, tissue homeostasis. This process is a protective mechanism against cellular stress, drug, external environment and tumor suppressive mechanism. It is generally divided into three distinct types including: (1) apoptosis; (2) autophagic cell death; and (3) necroptosis. Each type of cell death exhibits the specific morphological, molecular and biochemical characteristics [20]. We summarize the characteristics of the three types as listed in Table 1.

Apoptosis (Type I PCD) is characterized by chromatin condensation, DNA fragmentation and laddering, blebbing of nuclear or cytoplasmic and apoptotic bodies [21]. Apoptotic pathways include death-receptor pathway (extrinsic pathway), mitochondrial pathway (intrinsic pathway), endoplasmic reticulum (ER) stress, caspase-dependent pathway and caspase-independent pathway [22-27]. In the death-receptor pathway (extrinsic pathway), cell death is mediated by the interaction between death receptor

† These authors contributed equally to this work.

* Corresponding author. Department of Medical Research, China Medical University Hospital, China Medical University, No. 2, Yuh-Der Road, Taichung 404, Taiwan.

E-mail address: d0704@mail.cmuh.org.tw (F.-J. Tsai).

Table 1 – The characteristic features of programmed cell death [20].

Programmed cell death (PCD)	Apoptosis (type I PCD)	Autophagic cell death (type II PCD)	Necroptosis (type III PCD)
Feature	Chromatin condensation	Autophagic vesicles	Random DNA degradation
	DNA laddering	Blebbing	Swollen organelles
	Blebbing (nuclear, cytoplasmic)	Degradation of golgi	Cytoplasmic membrane rupture
	Apoptotic bodies		Potent inflammatory response
Key regulators	Caspases	Beclin-1	RIPK1
	Bcl-2 family members	LC III	TRAF2
	Cytochrome <i>c</i>	Atg family proteins	PARP
	AIF		
	Death-receptor proteins	ULK 1	Calpains
Relative pathways	Calpains	mTOR	
	Death-receptor Pathway (extrinsic pathway)	AMPK pathway	Glycosylphosphatidylinositol anchor biosynthesis
	Mitochondrial pathway (Intrinsic pathway)	Akt/mTOR pathway	Type 1 interferon family
	ER stress pathway	MAPK/ERK pathway	
	Caspase-dependent pathway	p53/stress pathway	Toll-like receptor signaling network
	Caspase-independent pathway	ER stress pathway	

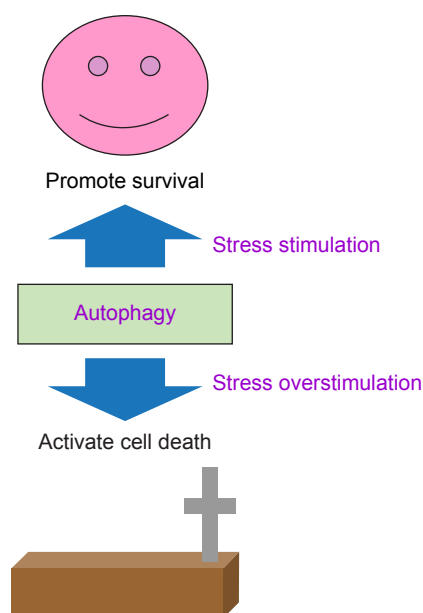


Fig. 1 - Autophagy serves as a double-edged sword. Autophagy promotes survival character when cells undergo stimuli and/or activates cell death when stimuli exceed a threshold.

proteins (such as Fas/CD95, DR4 and DR5) and the ligand (such as FasL and TRAIL), resulting in the staffing of an adaptor protein (FADD) and activation of caspase-8 and caspase-3/7 [22-32]. Mitochondria plays an essential role in the intrinsic pathway, which is inactivated by a drug or stress and then disrupts the mitochondrial membrane potential, causing production of reac-

tive oxygen species (ROS) and release of cytochrome *c*, Apaf-1, procaspase-9, AIF and Endo G signaling. The cytochrome *c*, Apaf-1 and procaspase-9 form an apoptosome complex to activate caspase-9 and caspase-3/-7. In addition, pro-apoptotic Bcl-2 family proteins (such as Bax, Bak, Bim, Bid, *etc.*) and anti-apoptotic proteins (such as Bcl-2, Bcl-xL, Mcl-1, *etc.*) regulate the process of mitochondrial pathway [33-41]. ER stress is induced by accumulation of unfolded/misfolded protein aggregating in ER or by excessive protein traffic. Increasing the proteins level of GADD 153, GRP 78, GRP 94 and ATF6, the hallmarks of ER stress, induce a rise in intracellular Ca^{2+} level, mitochondrial membrane depolarization and activation of calpain and caspase-12 in murine systems and/or caspase-4 in human cells [29, 36, 42-45].

Autophagic cell death (type II PCD) is a process by eliminating intracellular components through the lysosomal degradation in eukaryotic cells. Autophagy was first discovered during the late 1950s and early 1960s [46-48]. In the 1990s, the essential genes of the autophagy pathway were identified and characterized by the genetic screen studies in baker's yeast [49, 50]. Autophagy has been demonstrated to be involved in many biological processes, including maintenance of organelle integrity, protein quality control, regulation of the stress response and immune response [51-62]. Recently, autophagy has been shown to be modulated and to participate in the pathogenesis of human diseases, such as DM, neurodegenerative diseases, aging, pathogen infection diseases, vascular disease, pulmonary disease and cancer (Fig. 2) [13, 63-68]. Dr. Yoshinori Ohsumi discovered autophagy-related genes (ATGs) using a genetic screening approach in *Saccharomyces cerevisiae* and awarded the 2016 Nobel Prize in Physiology or Medicine for his remarkable contribution to autophagy research [1, 4, 69-71].

Autophagy is characterized by an increase of double-membrane vesicles (also known as autophagosomes or autophagic

Table 2 – Assays for monitoring autophagy.

Description	Methods	Reference
Monitor autophagosome number, volume, and content/cargo	Transmission electron microscopy (TEM)	[59, 146, 147]
	Western blotting	[59, 146, 147]
Atg8/LC3 detection and quantification	GFP-Atg8/LC3 fluorescence microscopy	[59, 146, 147]
	Immunohistochemistry	[59, 146, 147]
	Western blotting	[55, 56, 59, 60, 62, 146, 147]
	Real-time PCR	[55, 56, 59, 60, 62, 146, 147]
Additional autophagy-related protein markers	Immunohistochemistry	[55, 56, 59, 60, 62, 146, 147]
	Real-time PCR	[55, 56, 59, 60, 62, 146, 147]
Transcriptional regulation	Monodansylcadaverine (MDC)	[59, 146, 147]
	Acridine orange (AO)	[59, 146, 147]
	Neutral Red	[59, 146, 147]
Acidotropic dyes for identify acidified vesicular compartments	LysoSensor Blue	[59, 146, 147]
	Lyso-Tracker Red	[59, 146, 147]

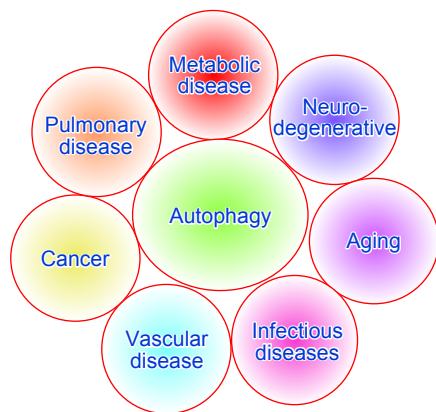


Fig. 2 - Autophagy participates in the pathogenesis of human diseases. These human disorders include DM, neurodegenerative diseases, aging, pathogen infection diseases, vascular disease, pulmonary disease and cancer.

vesicles) and degradation of golgi. Autophagy promotes cell survival in response to stress; however, once autophagy is overstimulated, cells can progress to autophagic cell death (Fig. 1). Here, we propose clearer definitions of the roles on autophagy: (A) the first role of autophagy functions as cell survival or cell protection [72-75]. (B) the second role of autophagy mediates programmed cell death (autophagic PCD). Upon stress, early-onset autophagy triggers cell protection and then late-onset autophagy induces cell death [76-80]. The detailed molecular mechanisms of autophagy will be described later.

Necroptosis (type III PCD), an irreversible cell death [81, 82], is characterized by a gain in cell volume, swollen organelles, DNA degradation, cytoplasmic plasma membrane rupture, subsequent loss of intracellular contents and potent inflammatory response. Relative necroptosis pathways include glycosylphos-

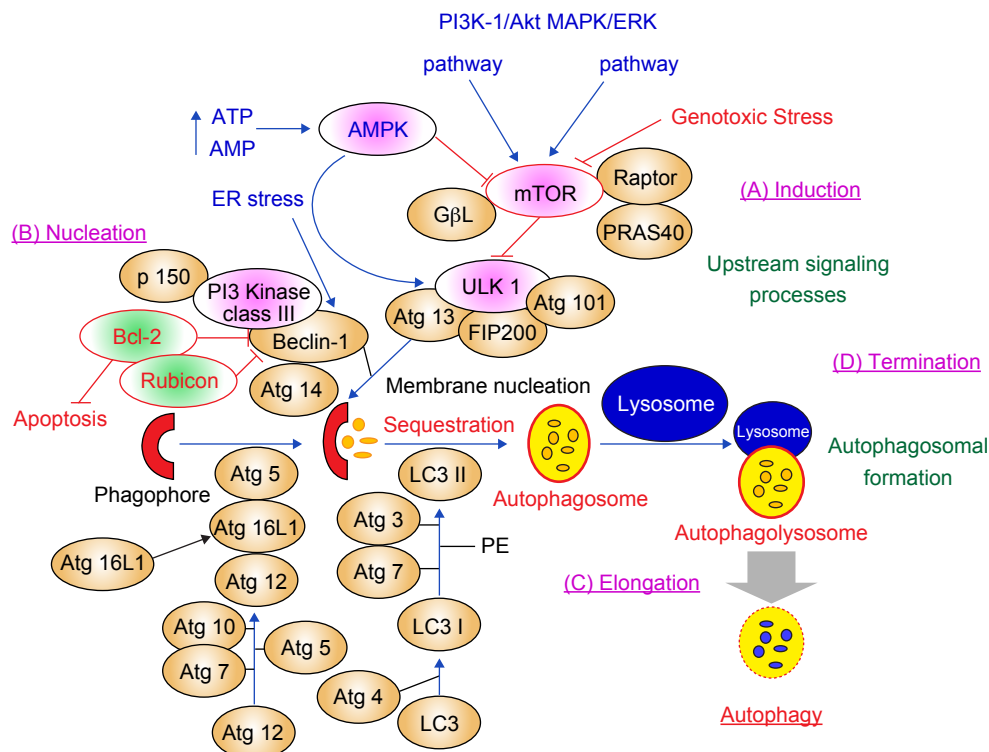
phatidylinositol anchor biosynthesis pathway, type 1 interferon family pathway and toll-like receptor signaling pathway (Table 1) [20]. The protein kinase RIP1 and RIP3 are central molecules in necroptosis. The RIPK1, TRAF2, PARP, calpains and RIPK3 proteins are identified and associated with programmed necrosis [83-89].

3. Assays for monitoring autophagy and pharmacological regulated agents

The features of autophagy are the massive accumulation of autophagic vacuoles (autophagosomes) in the cytoplasm of cells. Hereby, we present a series of methods to monitoring autophagy in Table 2. (1) Transmission electron microscopy (TEM) is used to observe autophagosome number, volume, and content analysis; (2) The lysosomal enzymes activity, assessment of the number, size, and location of lysosomes are examined by the uptake of fluorescent dyes (monodansylcadaverine (MDC), acridine orange (AO), neutral Red, LysoSensor Blue, Lyso-Tracker Red); (3) Autophagy-related proteins such as ATGs and LC3 are detected by western blotting or fluorescent protein tagging; (4) Autophagy-related gene expression levels are measured by western blotting or real-time PCR. Table 3 is a list of the pharmacological agents for assessing autophagy effects such as inhibition of lysosomal enzyme activities, fusion of organelles, or inter-compartmental transfer of molecules. (1) The 3-methyladenine (3-MA) is a PtdIns3K inhibitor and blocks an early stage of autophagy. (2) Bafilomycin A1 is a V-ATPase inhibitor and blocks fusion of autophagosomes with the vacuole. (3) Chloroquine is a lysosomotropic compound that elevates and neutralizes the lysosomal and vacuolar pH. (4) Leupeptin blocks lysosomal protein degradation. (5) Pepstatin A inhibits lysosomal protein degradation. (6) Resveratrol induces autophagy through activation of AMPK and (7) Tunicamycin is a glycosylation inhibitor that induces autophagy [55, 90, 91].

Table 3 – Pharmacological regulation of autophagy.

Method	Comments	Reference
3-Methyladenine (3-MA)	The PtdIns3K inhibitor and blocks an early stage of autophagy	[60, 62, 90, 91]
Bafilomycin A1	The V-ATPase inhibitor and blocks fusion of autophagosomes with the vacuole	[58, 62, 90, 91, 148]
Chloroquine	Lysosomotropic compounds that elevate and neutralize the lysosomal and vacuolar pH	[58, 90, 91]
Leupeptin	Block lysosomal protein degradation	[90, 91]
Pepstatin A	Block lysosomal degradation	[90, 91]
Tunicamycin	The glycosylation inhibitor that induces autophagy	[90, 91]
Resveratrol	Induction of autophagy <i>via</i> activation of AMPK	[55, 90, 91]

**Fig. 3 - There are four stages in the autophagic process: (1) induction, (2) nucleation, (3) elongation and (4) termination.**

4. The molecular mechanisms of autophagy

There are four stages in the autophagic process: (1) induction, (2) vesicle nucleation, (3) autophagosome membrane elongation and (4) termination/ fusion and degradation (Fig. 3) [92, 93]. In the normal status such as adequate nutrition, the mTORC1 complex (mTOR/GβL/Raptor/PRAS40) interacts with the ULK1 complex (ULK1/2-Atg13-FIP200-Atg101) to inhibit autophagy. When the mTORC1 complex senses genotoxic stress from hypoxia, starvation and low energy levels, mTORC1 dissociates from the ULK1 complex and initiates autophagy. Recent evidence suggests that mTORC1 complex is also regulated by PI3K-1/Akt, MAPK/ERK and AMPK signaling pathway. Activated AMPK phosphorylates Raptor and inhibits mTOR, which leads to activation of autophagy [94-98].

Beclin-1 complex (PI3Kinase class III, p150, Beclin-1 and

Atg14) is essential for vesicle nucleation and stimulates the fusion of autophagosomes with lysosomes [94-98]. During the stage of vesicle nucleation, Beclin-1 interacts with Atg14L, Bcl2, Rubicon, p150 and PI3Kinase class III proteins. Several regulators such as Bcl-2 protein (anti-apoptotic protein) and Rubicon bind Beclin-1 and inhibit the vesicle nucleation stage of autophagy.

Autophagosome membrane engagement is executed by the Atg12 and LC3 ubiquitin-like conjugation systems. (1) Atg12 ubiquitin-like conjugation system: ubiquitin-like Atg12 is conjugated to Atg5, Atg7 and Atg10. Atg10 serves as the E2 enzyme. The Atg5-Atg12/Atg16L complex is regulated by the Beclin-1 complex and localizes to the convex surface of the isolation membrane. (2) LC3 ubiquitin-like conjugation system: LC3 is cleaved by the Atg4 cysteine protease, sequentially processed by Atg7 and Atg3 and then conjugated to the membrane lipid phosphatidylethanolamine (the conjugated form is termed LC3-II). The Atg5-Atg12/Atg16L1 complex is necessary to promote the transforma-

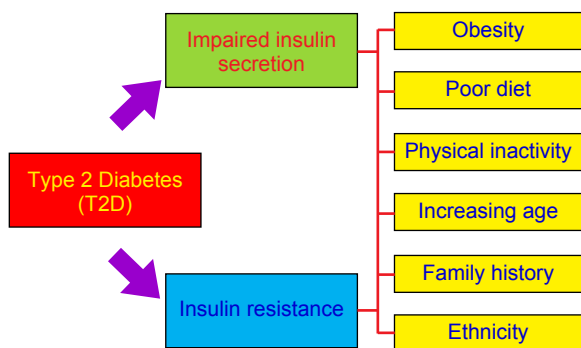


Fig. 4 - Etiology of type 2 DM. Two major physiological defects associated with T2D are reduced insulin sensitivity, insulin resistance and combined with impaired insulin secretion. Obesity, poor diet, physical inactivity, increasing age, family history and ethnicity lead to a higher risk of T2D.

tion of LC3-I to LC3-II [94-98].

At the terminal stage of autophagy, the autophagosome fuses with lysosomes to form autophagolysosomes. Autophagy allows the orderly degradation and recycling of cellular components [99]. The purpose of autophagy is to ensure quality control of organelles and proteins, as well as protection of intracellular homeostasis in stress and nutrient efficiency [94-99].

5. Type 2 diabetes mellitus (T2DM)

Diabetes mellitus (DM), commonly referred to as diabetes, is a metabolic and chronic disease in the world [100, 101]. DM patients have high blood sugar levels over a prolonged period. The character in DM is a relative or absolute lack of insulin, resulting in hyperglycemia [102]. Symptoms of hyperglycemia are frequent urination, increased thirst, and increased hunger. Acute complications of DM can include nonketotic hyperosmolar coma, diabetic ketoacidosis and death. Serious complications of DM include cardiovascular disease, stroke, chronic kidney failure, nephropathy, foot ulcers, neuropathy and damage to the eyes [103-106]. In 2014, approximately 422 million people were diagnosed with DM according to World Health Organization (WHO) report [107, 108]. In Taiwan, DM is ranked as the fifth leading cause of death in 2015 on the basis of statistics by the Ministry of Health and Welfare, R.O.C. (Taiwan) [101, 109].

There are three main types of diabetes mellitus: (1) Type 1 diabetes (T1D): also called insulin-dependent, juvenile or childhood-onset diabetes. T1D is characterized by deficient insulin production in the body. The pathology in T1D is described as an autoimmune disease because the pancreatic beta cells (insulin-producing tissue) are destructed in the islets of Langerhans [110]. T1D is diagnosed most in children and young adults. People with T1D require daily administration of insulin to regulate the amount of glucose in their blood [111]. Environmental factors and genetic influence play an important role in T1D [112, 113]. (2) Type 2 diabetes (T2D): formerly called non-insulin-dependent (NIDDM) or adult onset diabetes. T2D is the most common type of diabetes with prevalence in Taiwan. T2D begins with insulin resistance in which cells fail to respond to and uptake of insulin in the body [114-116]. Insulin resistance can be enhanced by weight reduction and exercise [117]. (3) Gestational diabetes: pregnant

women without a previous history of diabetes develop high blood sugar levels [118, 119].

The physiological defects in T2D that is reduced insulin sensitivity, insulin resistance and combined with impaired insulin secretion (Fig. 4). T2D occurs as a result of obesity, poor diet, physical inactivity, increasing age, family history and ethnicity. The defective or mutant insulin receptor may be caused no response to insulin in body tissues. Controversially, patients with T2D in the early stage often have a normal or high bone mineral density (BMD), associated with obesity and hyperinsulinemia, as well as altered level of insulin. When cells are insensitive to insulin (or insulin resistance), the pancreatic beta cells produce more and more insulin, which leads to the higher insulin concentration in blood (hyperinsulinemia). The pancreatic beta cells desperately secrete insulin and then gradually decline. T2D at late stage is characterized by insufficient secretion of insulin from the pancreatic beta cells, coupled with impaired insulin action in target tissues such as muscle, liver and fat. Hyperglycemia results when insulin secretion is unable to compensate for insulin resistance [120-124]. Mechanisms in the development and pharmacological treatments of T2D are summarized in Fig. 5 and Table 4.

6. Autophagy and type 2 diabetes (T2D)

Autophagy has been known to regulate the function of pancreatic beta cells and insulin-target tissues (skeletal muscle, liver and adipose tissue). T2D progression through impaired pancreatic beta cells function and development of insulin resistance has been associated with autophagy [125-128]. Upon insulin resistance, pancreatic cells enhance their insulin secretion (hyperinsulinemia) to compensate for hyperglycemia on the early onset of T2D (Fig. 5). In contrast, the number of pancreatic cells is progressive diminution through apoptotic cell death on the late onset of T2D [125, 129-131].

Many studies suggest that enhanced autophagy acts as a protective mechanism against oxidative stress in pancreatic beta cells [128, 132]. *In vivo* studies demonstrated that Atg7-deficient mice showed a decrease in the number of pancreatic beta cells, impairment of glucose tolerance and reduction in insulin secretion [133]. The insulin resistant mice (beta-cell-specific Atg7 knockout mice) model has been shown that autophagy plays a crucial role in the development of diabetes and in preserving the structure and function of pancreatic beta cells. Accumulation of autophagosomes in the pancreatic beta cell has been demonstrated in *db/db* mouse model [134-136]. Fujitani *et al.* showed that reduced insulin secretion was associated with pancreatic beta cell degeneration and impaired glucose in autophagy-deficient mice [136-138]. However, constitutively activated autophagy has injurious effects on pancreatic beta cells and chronic activation of autophagy causes autophagic cell death [135, 139-143].

9. Conclusion

The pancreatic beta cells control the releases of insulin and play an important role in the progression of T2D. Autophagy might function as a protective and pro-survival role on pancreatic beta cell death in T2D. Metformin has been widely used in the clinic therapy in T2D and has a protective effect on pancreatic beta cells from injury by activating autophagy through AMPK pathway [144, 145]. Therefore, it is urgent to understand the relationship

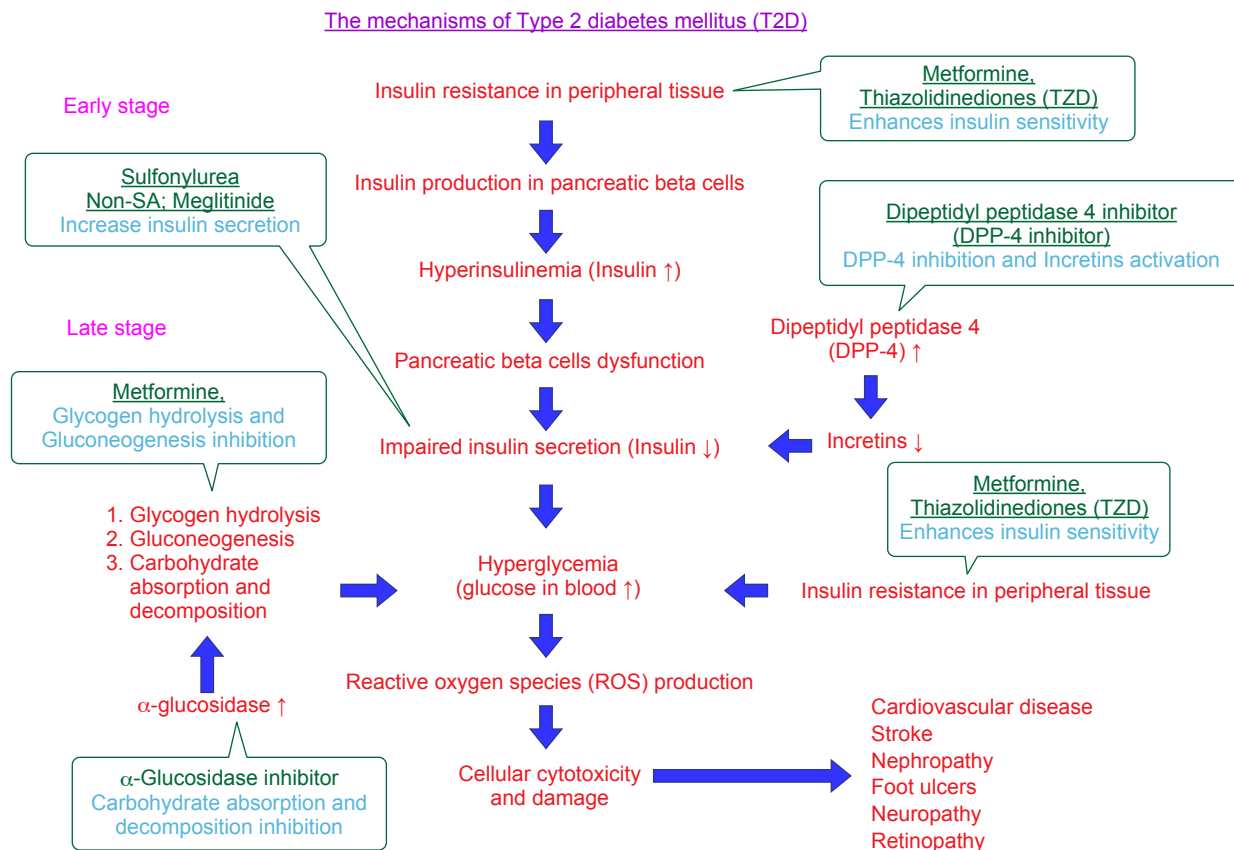


Fig. 5 - Mechanisms in the development and pharmacological treatments of T2D. Details are described in the text.

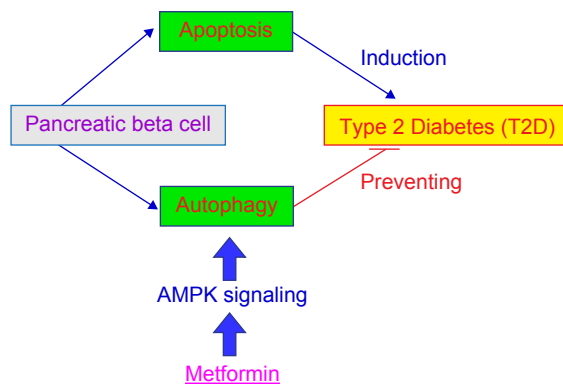


Fig. 6 - The role of autophagy and apoptosis in T2D.

of autophagy and T2D. We summarize the role of autophagy and apoptosis in T2D in Fig. 6. It is expected to develop new drugs and more effective agents targeted in autophagy for the therapy of T2D.

Acknowledgments

This work was supported by the grant from China Medical University Hospital, Taichung, Taiwan (DMR-106-122).

Open Access This article is distributed under terms of the Creative

Commons Attribution License which permits any use, distribution, and reproduction in any medium, provided original author(s) and source are credited.

REFERENCES

- [1] Ke PY. Horning cell self-digestion: Autophagy wins the 2016 Nobel Prize in Physiology or Medicine. *Biomed J* 2017; 40: 5-8.
- [2] Walton EL. Food for thought: Autophagy researcher wins 2016 Nobel Prize in Physiology or Medicine. *Biomed J* 2017; 40: 1-4.
- [3] Cao QH, Liu F, Yang ZL, Fu XH, Yang ZH, Liu Q, *et al.* Prognostic value of autophagy related proteins ULK1, Beclin 1, ATG3, ATG5, ATG7, ATG9, ATG10, ATG12, LC3B and p62/SQSTM1 in gastric cancer. *Am J Transl Res.* 2016; 8: 3831-47.
- [4] Tooze SA, Dikic I. Autophagy Captures the Nobel Prize. *Cell.* 2016; 167: 1433-35.
- [5] Sinha RA, Singh BK, Yen PM. Reciprocal Crosstalk Between Autophagic and Endocrine Signaling in Metabolic Homeostasis. *Endocr Rev.* 2017; 38: 69-102.
- [6] Yoshida GJ. Therapeutic strategies of drug repositioning targeting autophagy to induce cancer cell death: from pathophysiology to treatment. *J Hematol Oncol.* 2017; 10: 67.
- [7] Liu T, Yang P, Chen H, Huang Y, Liu Y, Waqas Y, *et al.* Global analysis of differential gene expression related to long-term sperm storage in oviduct of Chinese Soft-Shell Turtle *Pelodiscus sinensis*. *Sci Rep.* 2016; 6: 33296.

Table 4 – Pharmacological treatment for T2D.

Mechanisms	Type	Drugs	Reference
Increase insulin secretion from pancreatic β -cells.	Sulfonylureas (First generation)	Tolbutamide	[149-158]
		Chlorpropamide Acetohexamide	
	Sulfonylureas (Second generation)	Tolazamide Glibenclamide (Euglucon®)	[149-158]
Glipizide (Glidiab®) Gliclazide (Diamicon®)			
	Meglitinide	Glimepiride (Amaryl®) Repaglinide (Novonorm®) Nateglinide (Starlix®)	[159-162]
Enhances insulin sensitivity in liver and peripheral tissues by activation of AMP activated protein kinase. Glycogen hydrolysis and Gluconeogenesis inhibition.	Biguanide	Metformin	[163-167]
Absorption of glucose is delayed.	α -Glucosidase inhibitor	Acarbose	[168-170]
Enhances insulin sensitivity in peripheral tissues and liver by activation of peroxisome proliferator-activated receptor-gamma receptors.	Thiazolidinedione (TZD)	Rosiglitazone (Avandia®)	[171-174]
		Pioglitazone (Actos)	
Amplifies incretin pathway activation by inhibition of enzymatic breakdown of endogenous GLP-1 and GIP.	DPP-4 inhibitor	Sitagliptin (Januvia)	[150, 175-178]
		Saxagliptin (Onglyza)	
		Linagliptin (Trajenta)	
Activates incretin pathway by utilizing DPP-4 resistant analogue to GLP-1.	GLP-1 receptor agonist	Exenatide (Byetta)	[179-185]
		Liraglutide (Victoza)	
Activates insulin receptors to regulate metabolism of carbohydrate, fat and protein.	Insulin Bolus (prandial) insulins	Aspart (NovoRapid)	[111, 186-191]
		Glulisine (Apidra)	
		Lispro (Humalog)	
	Basal insulins	Detemir (Levemir)	
		Glargine (Lantus)	
	Premixed insulins	NPH (Humulin-N, Novolin ge NPH)	
		Biphasic insulin aspart (NovoMix 30)	
	Insulin lispro/lispro protamine suspension (Humalog Mix25, Mix50)		
	Premixed Regular-NPH (Humulin 30/70; Novolin ge 30/70, 40/60, 50/50)		

- [8] Chen W, Sun Y, Liu K, Sun X. Autophagy: a double-edged sword for neuronal survival after cerebral ischemia. *Neural Regen Res.* 2014; 9: 1210-6.
- [9] Sannigrahi MK, Singh V, Sharma R, Panda NK, Khullar M. Role of autophagy in head and neck cancer and therapeutic resistance. *Oral Dis.* 2015; 21: 283-91.
- [10] Lozy F, Karantza V. Autophagy and cancer cell metabolism. *Semin Cell Dev Biol.* 2012; 23: 395-401.
- [11] Lin LT, Dawson PW, Richardson CD. Viral interactions with macroautophagy: a double-edged sword. *Virology.* 2010; 402: 1-10.
- [12] Morel E, Mehrpour M, Botti J, Dupont N, Hamai A, Nascimbeni AC, *et al.* Autophagy: A Druggable Process. *Annu Rev Pharmacol Toxicol.* 2017; 57: 375-98.
- [13] Lippai M, Szatmari Z. Autophagy-from molecular mechanisms to clinical relevance. *Cell Biol Toxicol.* 2017; 33: 145-68.
- [14] He Q, Sha S, Sun L, Zhang J, Dong M. GLP-1 analogue improves hepatic lipid accumulation by inducing autophagy *via* AMPK/mTOR pathway. *Biochem Biophys Res Commun.* 2016; 476: 196-203.
- [15] Kawamori R. What is the natural history of type 2 diabetes mel-

litis. *Nihon Rinsho*. 2015; 73: 363-72.

- [16] Lin Y, Sun Z. *In vivo* pancreatic beta-cell-specific expression of antiaging gene Klotho: a novel approach for preserving beta-cells in type 2 diabetes. *Diabetes*. 2015; 64: 1444-58.
- [17] Wilson CM, Magnaudeix A, Yardin C, Terro F. Autophagy dysfunction and its link to Alzheimer's disease and type II diabetes mellitus. *CNS Neurol Disord Drug Targets*. 2014; 13: 226-46.
- [18] Kitamura T. The role of FOXO1 in beta-cell failure and type 2 diabetes mellitus. *Nat Rev Endocrinol*. 2013; 9: 615-23.
- [19] Yan J, Feng Z, Liu J, Shen W, Wang Y, Wertz K, *et al*. Enhanced autophagy plays a cardinal role in mitochondrial dysfunction in type 2 diabetic Goto-Kakizaki (GK) rats: ameliorating effects of (-)-epigallocatechin-3-gallate. *J Nutr Biochem*. 2012; 23: 716-24.
- [20] Rikiishi H. Autophagic and apoptotic effects of HDAC inhibitors on cancer cells. *J Biomed Biotechnol*. 2011; 2011: 830260.
- [21] Larsen BD, Sorensen CS. The caspase-activated DNase: apoptosis and beyond. *FEBS J*. 2017; 284: 1160-70.
- [22] Debatin KM. Cell death: From initial concepts to pathways to clinical applications—Personal reflections of a clinical researcher. *Biochem Biophys Res Commun*. 2017; 482: 445-49.
- [23] Siegmund D, Lang I, Wajant H. Cell death-independent activities of the death receptors CD95, TRAILR1, and TRAILR2. *FEBS J*. 2017; 284: 1131-59.
- [24] Maino B, Paparone S, Severini C, Ciotti MT, D'Agata V, Calissano P, *et al*. Drug target identification at the crossroad of neuronal apoptosis and survival. *Expert Opin Drug Discov*. 2017; 12: 249-59.
- [25] Bhat TA, Chaudhary AK, Kumar S, O'Malley J, Inigo JR, Kumar R, *et al*. Endoplasmic reticulum-mediated unfolded protein response and mitochondrial apoptosis in cancer. *Biochim Biophys Acta*. 2017; 1867: 58-66.
- [26] Yin S, Liu X, Fan L, Hu H. Mechanisms of cell death induction by food-borne mycotoxins. *Crit Rev Food Sci Nutr*. 2017.
- [27] Nemazee D. Mechanisms of central tolerance for B cells. *Nat Rev Immunol*. 2017.
- [28] Barnhart BC, Alappat EC, Peter ME. The CD95 type I/type II model. *Semin Immunol*. 2003; 15: 185-93.
- [29] Schultz DR, Harrington WJ, Jr. Apoptosis: programmed cell death at a molecular level. *Semin Arthritis Rheum*. 2003; 32: 345-69.
- [30] Wajant H. The Fas signaling pathway: more than a paradigm. *Science*. 2002; 296: 1635-6.
- [31] Eldadah BA, Faden AI. Caspase pathways, neuronal apoptosis, and CNS injury. *J Neurotrauma*. 2000; 17: 811-29.
- [32] Guchelaar HJ, Vermes A, Vermes I, Haanen C. Apoptosis: molecular mechanisms and implications for cancer chemotherapy. *Pharm World Sci*. 1997; 19: 119-25.
- [33] Tomita T. Apoptosis in pancreatic beta-islet cells in Type 2 diabetes. *Bosn J Basic Med Sci*. 2016; 16: 162-79.
- [34] Kollek M, Muller A, Egle A, Erlacher M. Bcl-2 proteins in development, health, and disease of the hematopoietic system. *FEBS J*. 2016; 283: 2779-810.
- [35] Moe GW, Marin-Garcia J. Role of cell death in the progression of heart failure. *Heart Fail Rev*. 2016; 21: 157-67.
- [36] Green DR, Llamas F. Cell Death Signaling. *Cold Spring Harb Perspect Biol*. 2015; 7.
- [37] Mc Gee MM. Targeting the Mitotic Catastrophe Signaling Pathway in Cancer. *Mediators Inflamm*. 2015; 2015: 146282.
- [38] Anuradha R, Saraswati M, Kumar KG, Rani SH. Apoptosis of beta cells in diabetes mellitus. *DNA Cell Biol*. 2014; 33: 743-8.
- [39] Khan KH, Blanco-Codesido M, Molife LR. Cancer therapeutics: Targeting the apoptotic pathway. *Crit Rev Oncol Hematol*. 2014; 90: 200-19.
- [40] Yang JS, Chen GW, Hsia TC, Ho HC, Ho CC, Lin MW, *et al*. Di-allyl disulfide induces apoptosis in human colon cancer cell line (COLO 205) through the induction of reactive oxygen species, endoplasmic reticulum stress, caspases cascade and mitochondrial-dependent pathways. *Food Chem Toxicol*. 2009; 47: 171-9.
- [41] Liao CL, Lai KC, Huang AC, Yang JS, Lin JJ, Wu SH, *et al*. Gallic acid inhibits migration and invasion in human osteosarcoma U-2 OS cells through suppressing the matrix metalloproteinase-2/-9, protein kinase B (PKB) and PKC signaling pathways. *Food Chem Toxicol*. 2012; 50: 1734-40.
- [42] Harada H, Grant S. Apoptosis regulators. *Rev Clin Exp Hematol*. 2003; 7: 117-38.
- [43] Fil'chenkov AA. Apoptosis-reactivating agents for targeted anti-cancer therapy. *Biomed Khim*. 2013; 59: 119-43.
- [44] Chevet E, Hetz C, Samali A. Endoplasmic reticulum stress-activated cell reprogramming in oncogenesis. *Cancer Discov*. 2015; 5: 586-97.
- [45] Huang WW, Chiu YJ, Fan MJ, Lu HF, Yeh HF, Li KH, *et al*. Kaempferol induced apoptosis *via* endoplasmic reticulum stress and mitochondria-dependent pathway in human osteosarcoma U-2 OS cells. *Mol Nutr Food Res*. 2010; 54: 1585-95.
- [46] Hurley JH, Nogales E. Next-generation electron microscopy in autophagy research. *Curr Opin Struct Biol*. 2016; 41: 211-16.
- [47] Tooze SA. Current views on the source of the autophagosome membrane. *Essays Biochem*. 2013; 55: 29-38.
- [48] Eskelinen EL, Reggiori F, Baba M, Kovacs AL, Seglen PO. Seeing is believing: the impact of electron microscopy on autophagy research. *Autophagy*. 2011; 7: 935-56.
- [49] Delorme-Axford E, Guimaraes RS, Reggiori F, Klionsky DJ. The yeast *Saccharomyces cerevisiae*: an overview of methods to study autophagy progression. *Methods*. 2015; 75: 3-12.
- [50] Guimaraes RS, Delorme-Axford E, Klionsky DJ, Reggiori F. Assays for the biochemical and ultrastructural measurement of selective and nonselective types of autophagy in the yeast *Saccharomyces cerevisiae*. *Methods*. 2015; 75: 141-50.
- [51] Villarejo-Zori B, Boya P. Autophagy and vision. *Med Sci (Paris)*. 2017; 33: 297-304.
- [52] Hurley JH, Young LN. Mechanisms of Autophagy Initiation. *Annu Rev Biochem*. 2017.
- [53] Botbol Y, Guerrero-Ros I, Macian F. Key roles of autophagy in regulating T-cell function. *Eur J Immunol*. 2016; 46: 1326-34.
- [54] Zientara-Rytter K, Subramani S. Autophagic degradation of peroxisomes in mammals. *Biochem Soc Trans*. 2016; 44: 431-40.
- [55] Chang CH, Lee CY, Lu CC, Tsai FJ, Hsu YM, Tsao JW, *et al*. Resveratrol-induced autophagy and apoptosis in cisplatin-resistant human oral cancer CAR cells: A key role of AMPK and Akt/mTOR signaling. *Int J Oncol*. 2017; 50: 873-82.
- [56] Yuan CH, Horng CT, Lee CF, Chiang NN, Tsai FJ, Lu CC, *et al*. Epigallocatechin gallate sensitizes cisplatin-resistant oral cancer CAR cell apoptosis and autophagy through stimulating AKT/STAT3 pathway and suppressing multidrug resistance 1 signaling. *Environ Toxicol*. 2017; 32: 845-55.

- [57] Hsieh MT, Chen HP, Lu CC, Chiang JH, Wu TS, Kuo DH, *et al.* The novel pterostilbene derivative ANK-199 induces autophagic cell death through regulating PI3 kinase class III/beclin 1/Atg-related proteins in cisplatinresistant CAR human oral cancer cells. *Int J Oncol.* 2014; 45: 782-94.
- [58] Huang AC, Lien JC, Lin MW, Yang JS, Wu PP, Chang SJ, *et al.* Tetrandrine induces cell death in SAS human oral cancer cells through caspase activation-dependent apoptosis and LC3-I and LC3-II activation-dependent autophagy. *Int J Oncol.* 2013; 43: 485-94.
- [59] Huang WW, Tsai SC, Peng SF, Lin MW, Chiang JH, Chiu YJ, *et al.* Kaempferol induces autophagy through AMPK and AKT signaling molecules and causes G2/M arrest *via* downregulation of CDK1/cyclin B in SK-HEP-1 human hepatic cancer cells. *Int J Oncol.* 2013; 42: 2069-77.
- [60] Lin C, Tsai SC, Tseng MT, Peng SF, Kuo SC, Lin MW, *et al.* AKT serine/threonine protein kinase modulates baicalin-triggered autophagy in human bladder cancer T24 cells. *Int J Oncol.* 2013; 42: 993-1000.
- [61] Liu CY, Yang JS, Huang SM, Chiang JH, Chen MH, Huang LJ, *et al.* Smh-3 induces G(2)/M arrest and apoptosis through calcium-mediated endoplasmic reticulum stress and mitochondrial signaling in human hepatocellular carcinoma Hep3B cells. *Oncol Rep.* 2013; 29: 751-62.
- [62] Tsai SC, Yang JS, Peng SF, Lu CC, Chiang JH, Chung JG, *et al.* Bufalin increases sensitivity to AKT/mTOR-induced autophagic cell death in SK-HEP-1 human hepatocellular carcinoma cells. *Int J Oncol.* 2012; 41: 1431-42.
- [63] Lv XX, Liu SS, Hu ZW. Autophagy-inducing natural compounds: a treasure resource for developing therapeutics against tissue fibrosis. *J Asian Nat Prod Res.* 2017; 19: 101-08.
- [64] Wu DJ, Adamopoulos IE. Autophagy and autoimmunity. *Clin Immunol.* 2017; 176: 55-62.
- [65] Papandreou ME, Tavernarakis N. Autophagy and the endo/exosomal pathways in health and disease. *Biotechnol J.* 2017; 12.
- [66] Tan YQ, Zhang J, Zhou G. Autophagy and its implication in human oral diseases. *Autophagy.* 2017; 13: 225-36.
- [67] Mowers EE, Sharifi MN, Macleod KF. Autophagy in cancer metastasis. *Oncogene.* 2017; 36: 1619-30.
- [68] Li Q, Liu Y, Sun M. Autophagy and Alzheimer's Disease. *Cell Mol Neurobiol.* 2017; 37: 377-88.
- [69] Zimmermann A, Kainz K, Andryushkova A, Hofer S, Madeo F, Carmona-Gutierrez D. Autophagy: one more Nobel Prize for yeast. *Microb Cell.* 2016; 3: 579-81.
- [70] Munz C. The Autophagic Machinery in Viral Exocytosis. *Front Microbiol.* 2017; 8: 269.
- [71] Palleria C, Leporini C, Maida F, Succurro E, De Sarro G, Arturi F, *et al.* Potential effects of current drug therapies on cognitive impairment in patients with type 2 diabetes. *Front Neuroendocrinol.* 2016; 42: 76-92.
- [72] Xue LY, Chiu SM, Azizuddin K, Joseph S, Oleinick NL. The death of human cancer cells following photodynamic therapy: apoptosis competence is necessary for Bcl-2 protection but not for induction of autophagy. *Photochem Photobiol.* 2007; 83: 1016-23.
- [73] Lavallard VJ, Meijer AJ, Codogno P, Gual P. Autophagy, signaling and obesity. *Pharmacol Res.* 2012; 66: 513-25.
- [74] Meijer AJ, Codogno P. Autophagy: regulation and role in disease. *Crit Rev Clin Lab Sci.* 2009; 46: 210-40.
- [75] Yan L, Sadoshima J, Vatner DE, Vatner SF. Autophagy: a novel protective mechanism in chronic ischemia. *Cell Cycle.* 2006; 5: 1175-7.
- [76] Fu D, Yu JY, Yang S, Wu M, Hammad SM, Connell AR, *et al.* Survival or death: a dual role for autophagy in stress-induced pericyte loss in diabetic retinopathy. *Diabetologia.* 2016; 59: 2251-61.
- [77] Kabbage M, Williams B, Dickman MB. Cell death control: the interplay of apoptosis and autophagy in the pathogenicity of Sclerostin sclerotiorum. *PLoS Pathog.* 2013; 9: e1003287.
- [78] Harr MW, Distelhorst CW. Apoptosis and autophagy: decoding calcium signals that mediate life or death. *Cold Spring Harb Perspect Biol.* 2010; 2: a005579.
- [79] Tyler MA, Ulasov IV, Lesniak MS. Cancer cell death by design: apoptosis, autophagy and glioma virotherapy. *Autophagy.* 2009; 5: 856-7.
- [80] Ricci MS, Zong WX. Chemotherapeutic approaches for targeting cell death pathways. *Oncologist.* 2006; 11: 342-57.
- [81] Mizumura K, Maruoka S, Gon Y, Choi AM, Hashimoto S. The role of necroptosis in pulmonary diseases. *Respir Investig.* 2016; 54: 407-12.
- [82] Inoue H, Tani K. Multimodal immunogenic cancer cell death as a consequence of anticancer cytotoxic treatments. *Cell Death Differ.* 2014; 21: 39-49.
- [83] Kaiser WJ, Sridharan H, Huang C, Mandal P, Upton JW, Gough PJ, *et al.* Toll-like receptor 3-mediated necrosis *via* TRIF, RIP3, and MLKL. *J Biol Chem.* 2013; 288: 31268-79.
- [84] Morgan MJ, Liu ZG. Programmed cell death with a necrotic-like phenotype. *Biomol Concepts.* 2013; 4: 259-75.
- [85] Linkermann A, De Zen F, Weinberg J, Kunzendorf U, Krautwald S. Programmed necrosis in acute kidney injury. *Nephrol Dial Transplant.* 2012; 27: 3412-9.
- [86] Li J, McQuade T, Siemer AB, Napetschnig J, Moriwaki K, Hsiao YS, *et al.* The RIP1/RIP3 necrosome forms a functional amyloid signaling complex required for programmed necrosis. *Cell.* 2012; 150: 339-50.
- [87] Andon FT, Fadeel B. Programmed cell death: molecular mechanisms and implications for safety assessment of nanomaterials. *Acc Chem Res.* 2013; 46: 733-42.
- [88] Galluzzi L, Kepp O, Kroemer G. RIP kinases initiate programmed necrosis. *J Mol Cell Biol.* 2009; 1: 8-10.
- [89] Conrad M, Angeli JP, Vandenabeele P, Stockwell BR. Regulated necrosis: disease relevance and therapeutic opportunities. *Nat Rev Drug Discov.* 2016; 15: 348-66.
- [90] Liu H, He Z, Simon HU. Targeting autophagy as a potential therapeutic approach for melanoma therapy. *Semin Cancer Biol.* 2013; 23: 352-60.
- [91] Belanger M, Rodrigues PH, Dunn WA, Jr., Progluske-Fox A. Autophagy: a highway for *Porphyromonas gingivalis* in endothelial cells. *Autophagy.* 2006; 2: 165-70.
- [92] Yang Y, Liang C. MicroRNAs: an emerging player in autophagy. *ScienceOpen Res.* 2015; 2015.
- [93] Qi W, Liang W, Jiang H, Miuyee Waye M. The function of miRNA in hepatic cancer stem cell. *Biomed Res Int.* 2013; 2013: 358902.
- [94] Salminen A, Kaarniranta K, Kauppinen A. AMPK and HIF signaling pathways regulate both longevity and cancer growth: the good news and the bad news about survival mechanisms. *Biogerontol-*

ogy. 2016; 17: 655-80.

- [95] Kume S, Koya D. Autophagy: A Novel Therapeutic Target for Diabetic Nephropathy. *Diabetes Metab J*. 2015; 39: 451-60.
- [96] Cetrullo S, D'Adamo S, Tantini B, Borzi RM, Flamigni F. mTOR, AMPK, and Sirt1: Key Players in Metabolic Stress Management. *Crit Rev Eukaryot Gene Expr*. 2015; 25: 59-75.
- [97] Dunlop EA, Tee AR. mTOR and autophagy: a dynamic relationship governed by nutrients and energy. *Semin Cell Dev Biol*. 2014; 36: 121-9.
- [98] Sridharan S, Jain K, Basu A. Regulation of autophagy by kinases. *Cancers (Basel)* 2011; 3: 2630-54.
- [99] Mizushima N, Komatsu M. Autophagy: renovation of cells and tissues. *Cell*. 2011; 147: 728-41.
- [100] Hou WH, Li CY, Chen LH, Wang LY, Kuo KN, Shen HN, *et al*. Prevalence of hand syndromes among patients with diabetes mellitus in Taiwan: A population-based study. *J Diabetes*. 2016.
- [101] Chen SY, Hsu YM, Lin YJ, Huang YC, Chen CJ, Lin WD, *et al*. Current concepts regarding developmental mechanisms in diabetic retinopathy in Taiwan. *Biomedicine (Taipei)*. 2016; 6: 7.
- [102] Middleton P, Crowther CA, Simmonds L. Different intensities of glycaemic control for pregnant women with pre-existing diabetes. *Cochrane Database Syst Rev*. 2016CD008540.
- [103] Adeshara KA, Diwan AG, Tupe RS. Diabetes and Complications: Cellular Signaling Pathways, Current Understanding and Targeted Therapies. *Curr Drug Targets*. 2016; 17: 1309-28.
- [104] Zatalia SR, Sanusi H. The role of antioxidants in the pathophysiology, complications, and management of diabetes mellitus. *Acta Med Indones*. 2013; 45: 141-7.
- [105] Resl M, Clodi M. Diabetes and cardiovascular complications. *Wien Med Wochenschr*. 2010; 160: 3-7.
- [106] Hahr AJ, Molitch ME. Diabetes, cardiovascular risk and nephropathy. *Cardiol Clin*. 2010; 28: 467-75.
- [107] Balakumar P, Maung UK, Jagadeesh G. Prevalence and prevention of cardiovascular disease and diabetes mellitus. *Pharmacol Res*. 2016; 113: 600-09.
- [108] Nowotny K, Jung T, Hohn A, Weber D, Grune T. Advanced glycation end products and oxidative stress in type 2 diabetes mellitus. *Biomolecules*. 2015; 5: 194-222.
- [109] Ministry of Health and Welfare ROC (Taiwan). <http://www.mohw.gov.tw/news/572256044>. Published. 2015. Updated Accessed.
- [110] Boldison J, Wong FS. Immune and Pancreatic beta Cell Interactions in Type 1 Diabetes. *Trends Endocrinol Metab*. 2016.
- [111] Kesavadev J. Insulin pump therapy in pregnancy. *J Pak Med Assoc*. 2016; 66: S39-44.
- [112] Zou D, Ye Y, Zou N, Yu J. Analysis of risk factors and their interactions in type 2 diabetes mellitus: A cross-sectional survey in Guilin, China. *J Diabetes Investig*. 2016.
- [113] Hotta-Iwamura C, Tarbell KV. Type 1 diabetes genetic susceptibility and dendritic cell function: potential targets for treatment. *J Leukoc Biol*. 2016; 100: 65-80.
- [114] Sakai S, Tanimoto K, Imbe A, Inaba Y, Shishikura K, Tanimoto Y, *et al*. Decreased beta-Cell Function Is Associated with Reduced Skeletal Muscle Mass in Japanese Subjects without Diabetes. *PLoS One*. 2016; 11: e0162603.
- [115] Honardoost M, Arefian E, Soleimani M, Soudi S, Sarookhani MR. Development of Insulin Resistance through Induction of miRNA-135 in C2C12 Cells. *Cell J*. 2016; 18: 353-61.
- [116] Rezaei S, LoBue S, Henderson CE. Diabetes prevention: Reproductive age women affected by insulin resistance. *Womens Health (Lond)*. 2016; 12: 427-32.
- [117] Arias-Loste MT, Ranchal I, Romero-Gomez M, Crespo J. Irisin, a link among fatty liver disease, physical inactivity and insulin resistance. *Int J Mol Sci*. 2014; 15: 23163-78.
- [118] Viecceli C, Remonti LR, Hirakata VN, Mastella LS, Gnielka V, Oppermann ML, *et al*. Weight gain adequacy and pregnancy outcomes in gestational diabetes: a meta-analysis. *Obes Rev*. 2017; 18: 567-80.
- [119] Logan KM, Gale C, Hyde MJ, Santhakumaran S, Modi N. Diabetes in pregnancy and infant adiposity: systematic review and meta-analysis. *Arch Dis Child Fetal Neonatal Ed*. 2017; 102: F65-F72.
- [120] Tian J, Lian F, Yang L, Tong X. Evaluation of Chinese Herbal Medicine Jinlida in Type 2 Diabetes Patients based on Stratification: Results of Subgroup Analysis from 12-Week Trial. *J Diabetes*. 2017.
- [121] Garabadu D, Krishnamurthy S. Metformin attenuates hepatic insulin resistance in type-2 diabetic rats through PI3K/Akt/GLUT-4 signalling independent to bicuculline-sensitive GABAA receptor stimulation. *Pharm Biol*. 2017; 55: 722-28.
- [122] Eaton SB, Eaton SB. Physical Inactivity, Obesity, and Type 2 Diabetes: An Evolutionary Perspective. *Res Q Exerc Sport*. 2017; 88: 1-8.
- [123] Wong P, Weiner MG, Hwang WT, Yang YX. Insulin therapy and colorectal adenomas in patients with diabetes mellitus. *Cancer Epidemiol Biomarkers Prev*. 2012; 21: 1833-40.
- [124] Yang YX, Hennessy S, Lewis JD. Insulin therapy and colorectal cancer risk among type 2 diabetes mellitus patients. *Gastroenterology*. 2004; 127: 1044-50.
- [125] Marrif HI, Al-Sunousi SI. Pancreatic beta Cell Mass Death. *Front Pharmacol*. 2016; 7: 83.
- [126] Lee MS. Role of islet beta cell autophagy in the pathogenesis of diabetes. *Trends Endocrinol Metab*. 2014; 25: 620-7.
- [127] Wang Y, Li YB, Yin JJ, Wang Y, Zhu LB, Xie GY, *et al*. Autophagy regulates inflammation following oxidative injury in diabetes. *Autophagy*. 2013; 9: 272-7.
- [128] Hur KY, Jung HS, Lee MS. Role of autophagy in beta-cell function and mass. *Diabetes Obes Metab*. 2010; 12 Suppl 2: 20-6.
- [129] Montane J, Cadavez L, Novials A. Stress and the inflammatory process: a major cause of pancreatic cell death in type 2 diabetes. *Diabetes Metab Syndr Obes*. 2014; 7: 25-34.
- [130] Quan W, Jo EK, Lee MS. Role of pancreatic beta-cell death and inflammation in diabetes. *Diabetes Obes Metab*. 2013; 15 Suppl 3: 141-51.
- [131] Lee MS, Kim KA, Kim HS. Role of pancreatic beta-cell death and cell death-associated inflammation in diabetes. *Curr Mol Med*. 2012; 12: 1297-310.
- [132] Rivera JF, Costes S, Gurlo T, Glabe CG, Butler PC. Autophagy defends pancreatic beta cells from human islet amyloid polypeptide-induced toxicity. *J Clin Invest*. 2014; 124: 3489-500.
- [133] Kim J, Cheon H, Jeong YT, Quan W, Kim KH, Cho JM, *et al*. Amyloidogenic peptide oligomer accumulation in autophagy-deficient beta cells induces diabetes. *J Clin Invest*. 2014; 124: 3311-24.
- [134] Lo MC, Chen MH, Lee WS, Lu CI, Chang CR, Kao SH, *et al*. Nepsilon-(carboxymethyl) lysine-induced mitochondrial fission and mitophagy cause decreased insulin secretion from beta-cells.

Am J Physiol Endocrinol Metab. 2015; 309: E829-39.

- [135] Las G, Shirihai OS. The role of autophagy in beta-cell lipotoxicity and type 2 diabetes. *Diabetes Obes Metab.* 2010; 12 Suppl 2: 15-9.
- [136] Fujitani Y, Kawamori R, Watada H. The role of autophagy in pancreatic beta-cell and diabetes. *Autophagy.* 2009; 5: 280-2.
- [137] Abe H, Uchida T, Hara A, Mizukami H, Komiya K, Koike M, *et al.* Exendin-4 improves beta-cell function in autophagy-deficient beta-cells. *Endocrinology.* 2013; 154: 4512-24.
- [138] Ebato C, Uchida T, Arakawa M, Komatsu M, Ueno T, Komiya K, *et al.* Autophagy is important in islet homeostasis and compensatory increase of beta cell mass in response to high-fat diet. *Cell Metab.* 2008; 8: 325-32.
- [139] Demirtas L, Guclu A, Erdur FM, Akbas EM, Ozcicek A, Onk D, *et al.* Apoptosis, autophagy & endoplasmic reticulum stress in diabetes mellitus. *Indian J Med Res.* 2016; 144: 515-24.
- [140] Pabon MA, Ma KC, Choi AM. Autophagy and Obesity-Related Lung Disease. *Am J Respir Cell Mol Biol.* 2016; 54: 636-46.
- [141] Wang S, Sun QQ, Xiang B, Li XJ. Pancreatic islet cell autophagy during aging in rats. *Clin Invest Med.* 2013; 36: E72-80.
- [142] Bartolome A, Guillen C, Benito M. Autophagy plays a protective role in endoplasmic reticulum stress-mediated pancreatic beta cell death. *Autophagy.* 2012; 8: 1757-68.
- [143] Masini M, Lupi R, Bugliani M, Boggi U, Filipponi F, Masiello P, *et al.* A role for autophagy in beta-cell life and death. *Islets.* 2009; 1: 157-9.
- [144] Jiang Y, Huang W, Wang J, Xu Z, He J, Lin X, *et al.* Metformin plays a dual role in MIN6 pancreatic beta cell function through AMPK-dependent autophagy. *Int J Biol Sci.* 2014; 10: 268-77.
- [145] Masini M, Bugliani M, Lupi R, del Guerra S, Boggi U, Filipponi F, *et al.* Autophagy in human type 2 diabetes pancreatic beta cells. *Diabetologia.* 2009; 52: 1083-6.
- [146] Zakeri Z, Melendez A, Lockshin RA. Detection of autophagy in cell death. *Methods Enzymol.* 2008; 442: 289-306.
- [147] Gurusamy N, Das DK. Detection of cell death by autophagy. *Methods Mol Biol.* 2009; 559: 95-103.
- [148] Yu FS, Yu CS, Chen JC, Yang JL, Lu HF, Chang SJ, *et al.* Tetrandrine induces apoptosis *via* caspase-8, -9, and -3 and poly (ADP ribose) polymerase dependent pathways and autophagy through beclin-1/LC3-I, II signaling pathways in human oral cancer HSC-3 cells. *Environ Toxicol.* 2016; 31: 395-406.
- [149] Scheen AJ. Pharmacotherapy of 'treatment resistant' type 2 diabetes. *Expert Opin Pharmacother.* 2017; 18: 503-15.
- [150] Scheen AJ. DPP-4 inhibitor plus SGLT-2 inhibitor as combination therapy for type 2 diabetes: from rationale to clinical aspects. *Expert Opin Drug Metab Toxicol.* 2016; 12: 1407-17.
- [151] Nauck M. Incretin therapies: highlighting common features and differences in the modes of action of glucagon-like peptide-1 receptor agonists and dipeptidyl peptidase-4 inhibitors. *Diabetes Obes Metab.* 2016; 18: 203-16.
- [152] Triplitt C, Solis-Herrera C. GLP-1 Receptor Agonists: Practical Considerations for Clinical Practice. *Diabetes Educ.* 2015; 41: 32S-46S.
- [153] Altaf QA, Barnett AH, Tahrani AA. Novel therapeutics for type 2 diabetes: insulin resistance. *Diabetes Obes Metab.* 2015; 17: 319-34.
- [154] Pawlyk AC, Giacomini KM, McKeon C, Shuldiner AR, Florez JC. Metformin pharmacogenomics: current status and future directions. *Diabetes.* 2014; 63: 2590-9.
- [155] Viollet B, Guigas B, Sanz Garcia N, Leclerc J, Foretz M, Andreelli F. Cellular and molecular mechanisms of metformin: an overview. *Clin Sci (Lond).* 2012; 122: 253-70.
- [156] Schroner Z, Javorsky M, Kozarova M, Tkac I. Pharmacogenetics of oral antidiabetic treatment. *Bratisl Lek Listy.* 2011; 112: 441-6.
- [157] Shah IM, Mackay SP, McKay GA. Therapeutic strategies in the treatment of diabetic nephropathy—a translational medicine approach. *Curr Med Chem.* 2009; 16: 997-1016.
- [158] Caballero AE. Long-term studies of treatments for type 2 diabetes. *Postgrad Med.* 2017; 129: 352-65.
- [159] Mayerson AB, Inzucchi SE. Type 2 diabetes therapy. A pathophysiologically based approach. *Postgrad Med.* 2002; 111: 83-4, 87-92, 95.
- [160] Quillen DM, Kuritzky L. Type 2 diabetes management: a comprehensive clinical review of oral medications. *Compr Ther.* 2002; 28: 50-61.
- [161] Hemmingsen B, Schroll JB, Lund SS, Wetterslev J, Gluud C, Vaag A, *et al.* Sulphonylurea monotherapy for patients with type 2 diabetes mellitus. *Cochrane Database Syst Rev.* 2013CD009008.
- [162] Lamos EL, Stein SA, Davis SN. Sulfonylureas and meglitinides: historical and contemporary issues. *Panminerva Med.* 2013; 55: 239-51.
- [163] Drugs for type 2 diabetes. *Med Lett Drugs Ther.* 2017; 59: 9-18.
- [164] Feng Y, Yang H. Metformin—a potentially effective drug for gestational diabetes mellitus: a systematic review and meta-analysis. *J Matern Fetal Neonatal Med.* 2016:1-8.
- [165] Liu F, Yan L, Wang Z, Lu Y, Chu Y, Li X, *et al.* Metformin therapy and risk of colorectal adenomas and colorectal cancer in type 2 diabetes mellitus patients: A systematic review and meta-analysis. *Oncotarget.* 2017; 8: 16017-26.
- [166] Tan MH, Alquraini H, Mizokami-Stout K, MacEachern M. Metformin: From Research to Clinical Practice. *Endocrinol Metab Clin North Am.* 2016; 45: 819-43.
- [167] Anabtawi A, Miles JM. Metformin: Nonglycemic Effects and Potential Novel Indications. *Endocr Pract.* 2016; 22: 999-1007.
- [168] Katsuta H, Ishida H. Alpha glucosidase inhibitor. *Nihon Rinsho.* 2006; 64 Suppl 9: 637-45.
- [169] Tielmans A, Virally M, Coupaye M, Laloi-Michelin M, Meas T, Guillaumeau PJ. Drug treatment in type 2 diabetes (part 2). *Presse Med.* 2007; 36: 467-74.
- [170] Baba S. Pioglitazone: a review of Japanese clinical studies. *Curr Med Res Opin.* 2001; 17: 166-89.
- [171] Smith JD, Mills E, Carlisle SE. Treatment of Pediatric Type 2 Diabetes. *Ann Pharmacother.* 2016; 50: 768-77.
- [172] Klein J, Charach R, Sheiner E. Treating diabetes during pregnancy. *Expert Opin Pharmacother.* 2015; 16: 357-68.
- [173] Soccio RE, Chen ER, Lazar MA. Thiazolidinediones and the promise of insulin sensitization in type 2 diabetes. *Cell Metab.* 2014; 20: 573-91.
- [174] Hostalek U, Gwilt M, Hildemann S. Therapeutic Use of Metformin in Prediabetes and Diabetes Prevention. *Drugs.* 2015; 75: 1071-94.
- [175] Ryan G. Dipeptidyl peptidase-4 inhibitor use in patients with type 2 diabetes and cardiovascular disease or risk factors. *Postgrad Med.* 2015; 127: 842-54.

- [176] Mamza J, Marlin C, Wang C, Chokkalingam K, Idris I. DPP-4 inhibitor therapy and bone fractures in people with Type 2 diabetes-A systematic review and meta-analysis. *Diabetes Res Clin Pract.* 2016; 116: 288-98.
- [177] Wright JJ, Tylee TS. Pharmacologic Therapy of Type 2 Diabetes. *Med Clin North Am.* 2016; 100: 647-63.
- [178] Sharma SK, Panneerselvam A, Singh KP, Parmar G, Gadge P, Swami OC. Teneeligliptin in management of type 2 diabetes mellitus. *Diabetes Metab Syndr Obes.* 2016; 9: 251-60.
- [179] Tella SH, Rendell MS. Glucagon-like polypeptide agonists in type 2 diabetes mellitus: efficacy and tolerability, a balance. *Ther Adv Endocrinol Metab.* 2015; 6: 109-34.
- [180] Eng C, Kramer CK, Zinman B, Retnakaran R. Glucagon-like peptide-1 receptor agonist and basal insulin combination treatment for the management of type 2 diabetes: a systematic review and meta-analysis. *Lancet.* 2014; 384: 2228-34.
- [181] Rowzee AM, Cawley NX, Chiorini JA, Di Pasquale G. Glucagon-like peptide-1 gene therapy. *Exp Diabetes Res.* 2011; 2011: 601047.
- [182] Gallwitz B. Glucagon-like peptide-1 analogues for Type 2 diabetes mellitus: current and emerging agents. *Drugs.* 2011; 71: 1675-88.
- [183] Tzefos M, Olin JL. Glucagon-like peptide-1 analog and insulin combination therapy in the management of adults with type 2 diabetes mellitus. *Ann Pharmacother.* 2010; 44: 1294-300.
- [184] Mundil D, Cameron-Vendrig A, Husain M. GLP-1 receptor agonists: a clinical perspective on cardiovascular effects. *Diab Vasc Dis Res.* 2012; 9: 95-108.
- [185] Trujillo JM, Nuffer W. GLP-1 receptor agonists for type 2 diabetes mellitus: recent developments and emerging agents. *Pharmacotherapy.* 2014; 34: 1174-86.
- [186] Vos RC, van Avendonk MJ, Jansen H, Goudswaard AN, van den Donk M, Gorter K, *et al.* Insulin monotherapy compared with the addition of oral glucose-lowering agents to insulin for people with type 2 diabetes already on insulin therapy and inadequate glycaemic control. *Cochrane Database Syst Rev.* 2016; 9: CD006992.
- [187] Barnosky A, Shah L, Meah F, Emanuele N, Emanuele MA, Mazhari A. A primer on concentrated insulins: what an internist should know. *Postgrad Med.* 2016; 128: 381-90.
- [188] Millstein R, Becerra NM, Shubrook JH. Insulin pumps: Beyond basal-bolus. *Cleve Clin J Med.* 2015; 82: 835-42.
- [189] Meah F, Juneja R. Insulin tactics in type 2 diabetes. *Med Clin North Am.* 2015; 99: 157-86.
- [190] Keating GM. Insulin detemir: a review of its use in the management of diabetes mellitus. *Drugs.* 2012; 72: 2255-87.
- [191] Meneghini LF. Insulin therapy for type 2 diabetes. *Endocrine.* 2013; 43: 529-34.